



Precision Oncology in Lung Cancer – Diagnostics

ESMO DEEP DIVE WEBINAR SERIES: LUNG CANCER

Thomas John, *Chair*

*Peter MacCallum Cancer Centre,
University of Melbourne, Australia*

5 February 2025

| | |
|--------|--|
| 5 min | Welcome and introduction Tom John |
| 25 min | Precision through genomics and beyond (multi-omics) Celine Mascaux |
| 25 min | Harnessing liquid biopsy for diagnosis, monitoring Christian Rolfo |
| 25 min | Beyond Xray vision – novel imaging technologies and radiomics Raquel Lopez |
| 15 min | QnA and Discussion All speakers |



Thomas John

Chair

Peter MacCallum Cancer
Centre, University of
Melbourne



Celine Mascaux

Speaker

Strasbourg University
Hospital



Christian Rolfo

Speaker

The Ohio State University
Wexner Medical Center



Raquel Perez Lopez

Speaker

Vall d'Hebron Institute of
Oncology

Learning objectives

- To acquire a deeper understanding of the clinical course of lung cancer.
- To understand biological hypotheses on classification and risk stratification, ongoing/required research in therapeutics and knowledge of use of omics technologies for biomarker-enabled precision medicine for lung cancer.
- To develop skills and abilities for critical analysis, interpretation of research data and therapeutic strategies.
- To become better equipped for informed, innovative thinking and engagement in ongoing or new research projects.



Precision Oncology in Lung cancer – Diagnostics (genomics, transcriptomics (SCLC), plasma, radiomics)

Celine Mascaux

04 février 2025



Precision medicine through genomics and beyond (multi-omics)

Celine Mascaux

04 février 2025

WHY

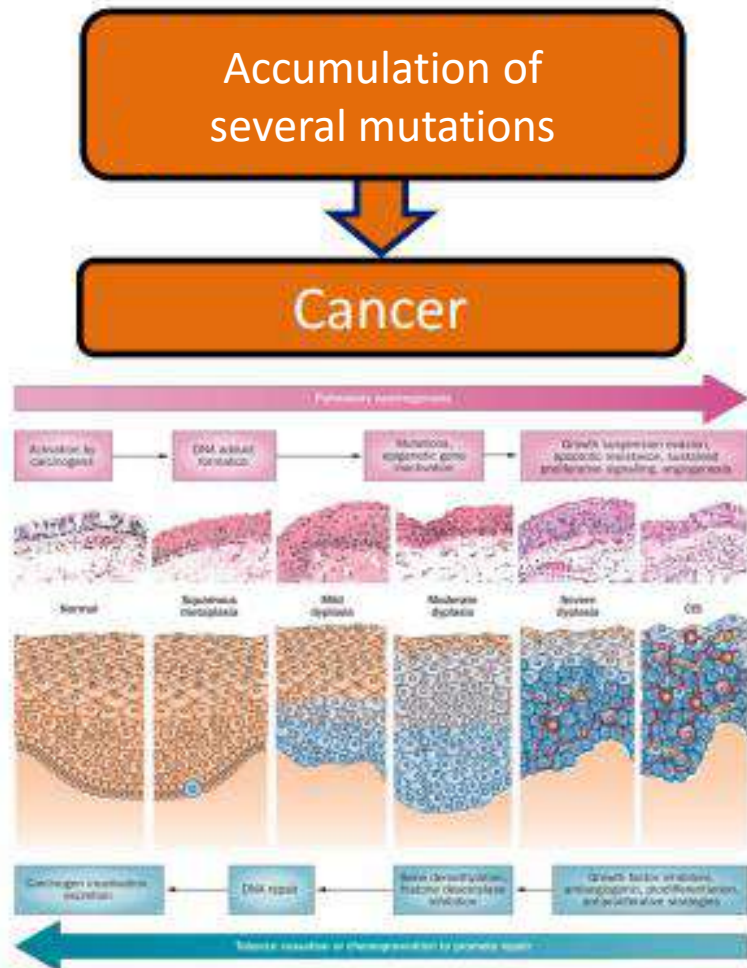
IN WHOM do we analyse tumours for molecular abnormalities ?

HOW

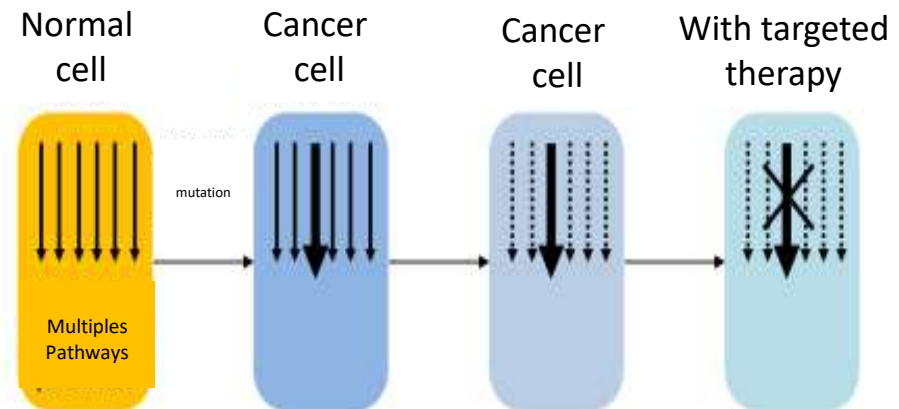
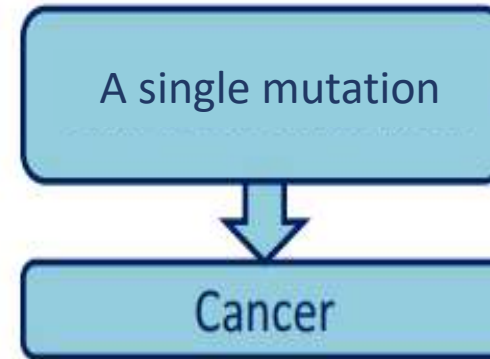
WHY do we analyse tumours for molecular abnormalities ?

Two models of carcinogenesis

Multistep carcinogenesis

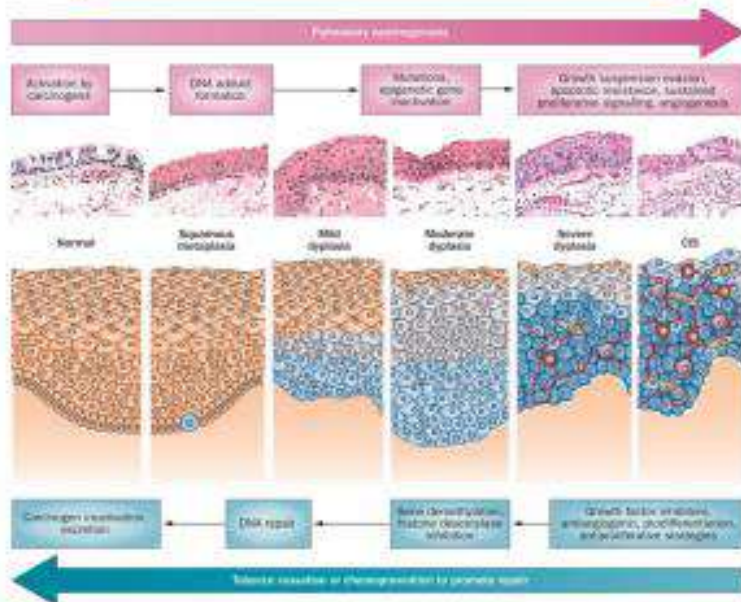
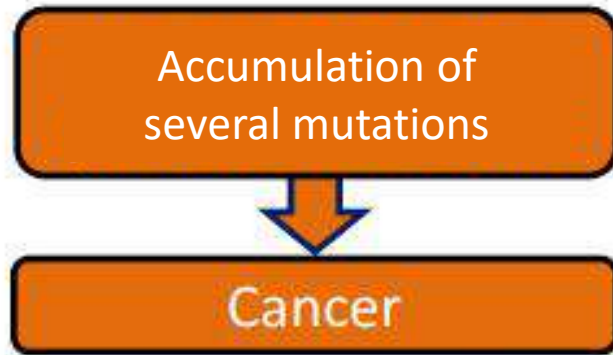


Oncogenic addiction



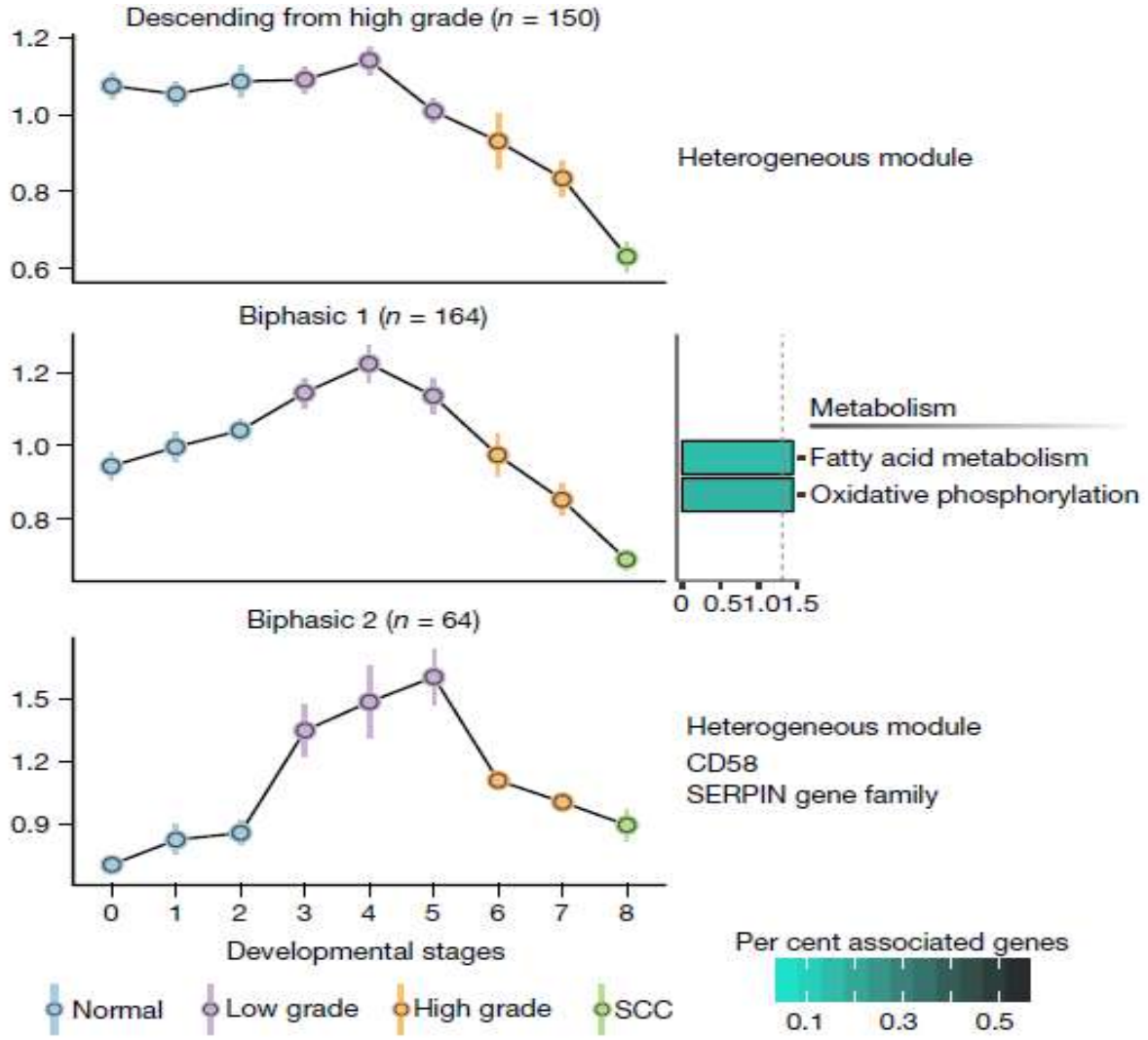
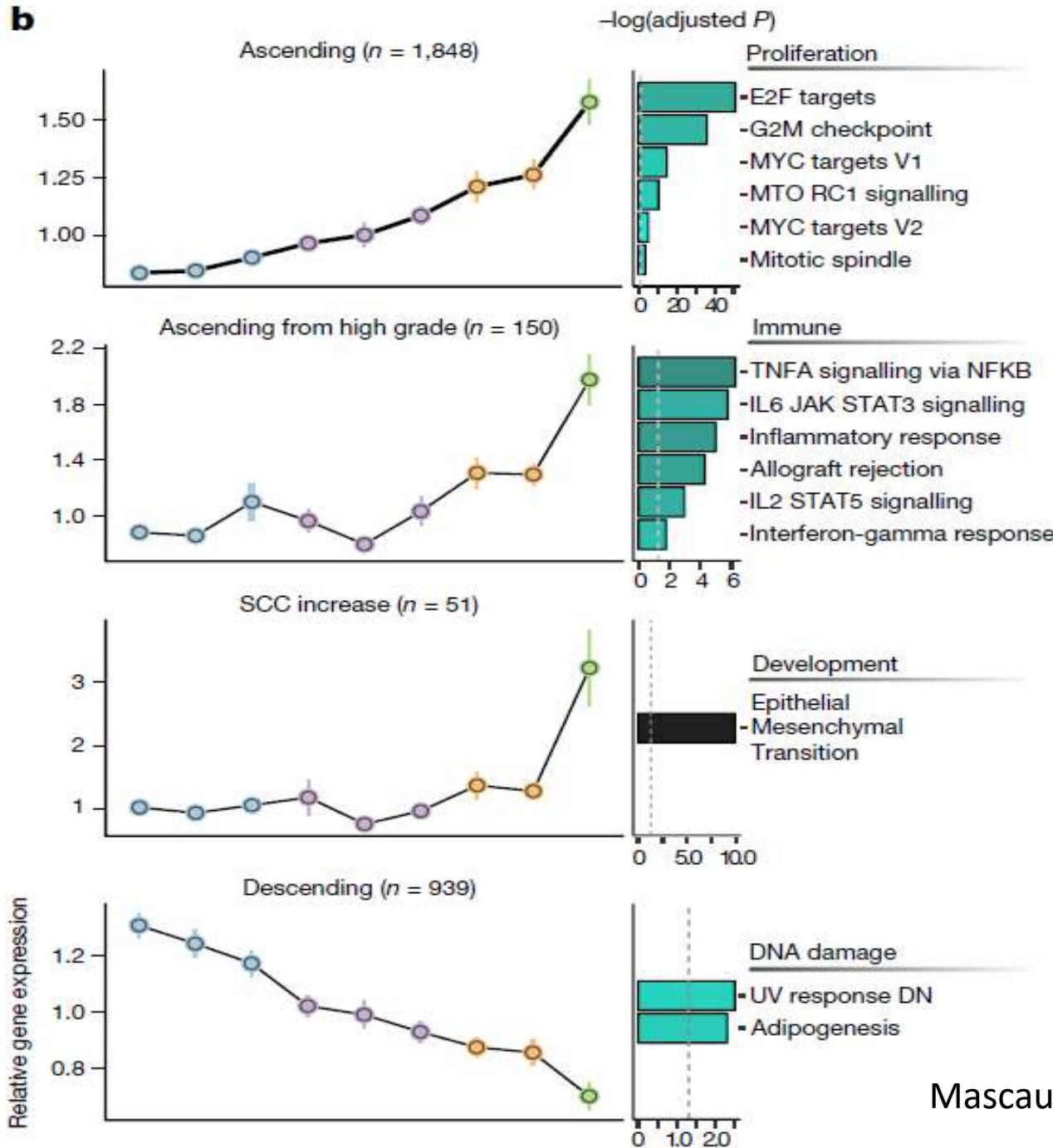
Two models of carcinogenesis

Multistep carcinogenesis



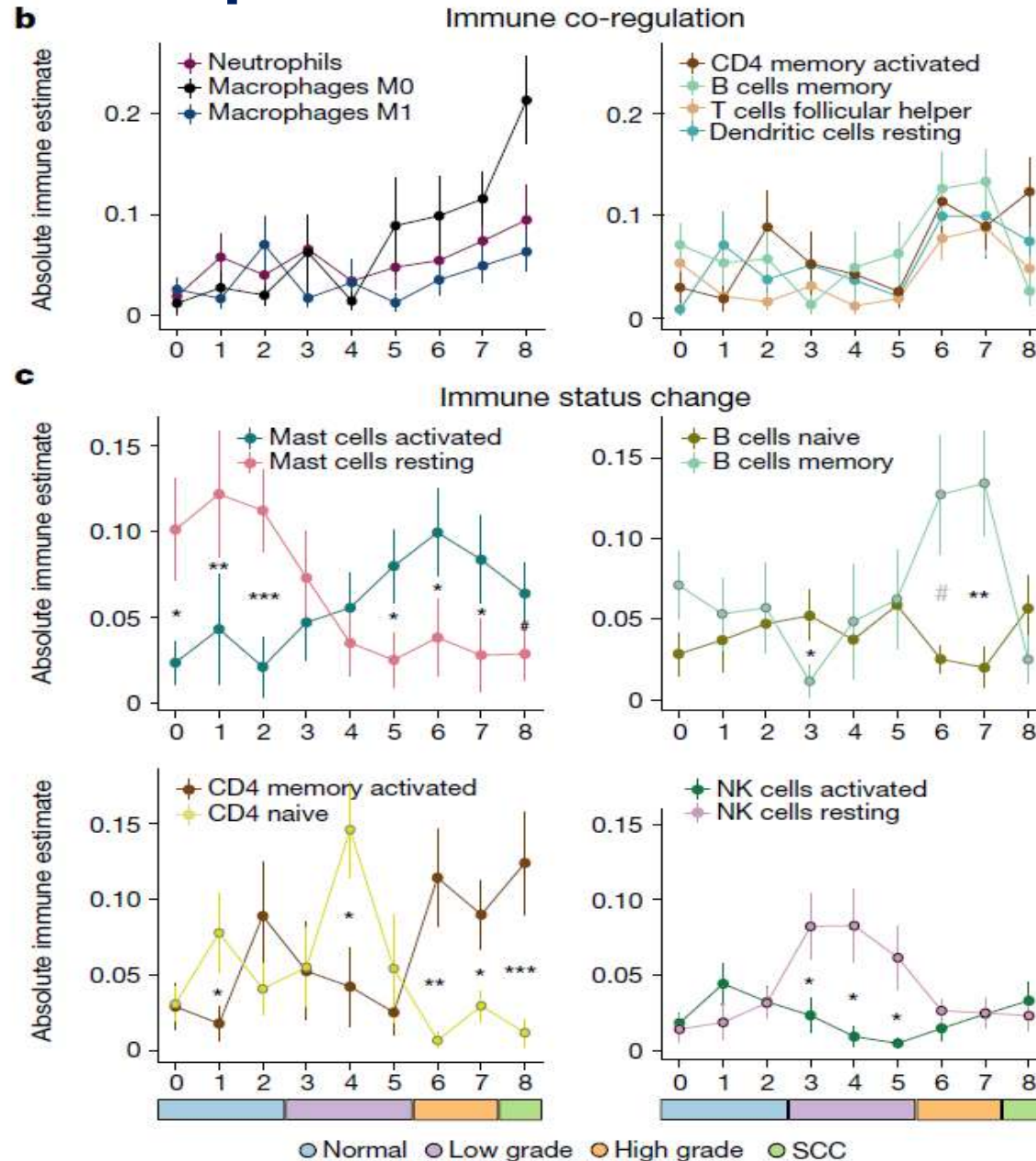
Multi-step carcinogenesis

b



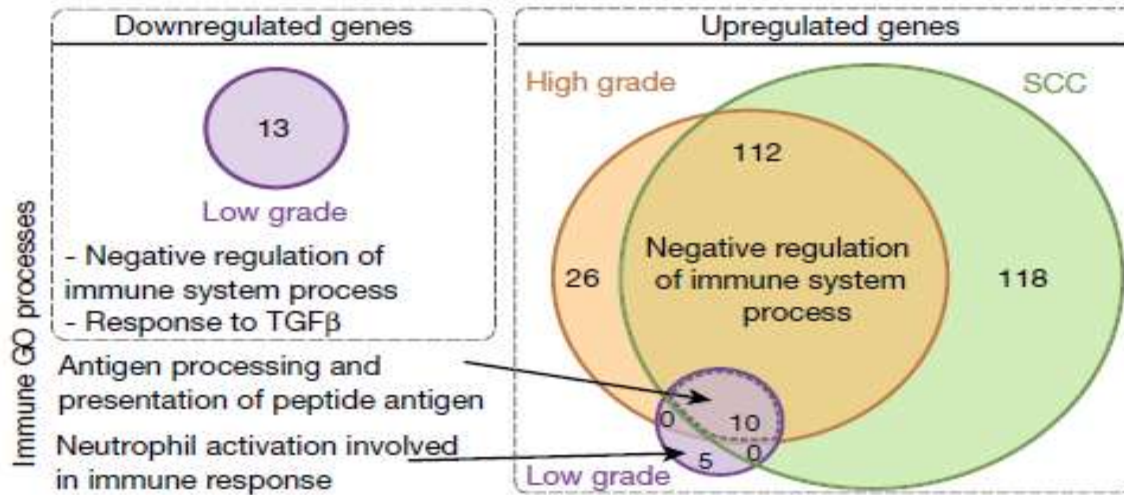
Mascaux et al, Nature 2019

Active immune response

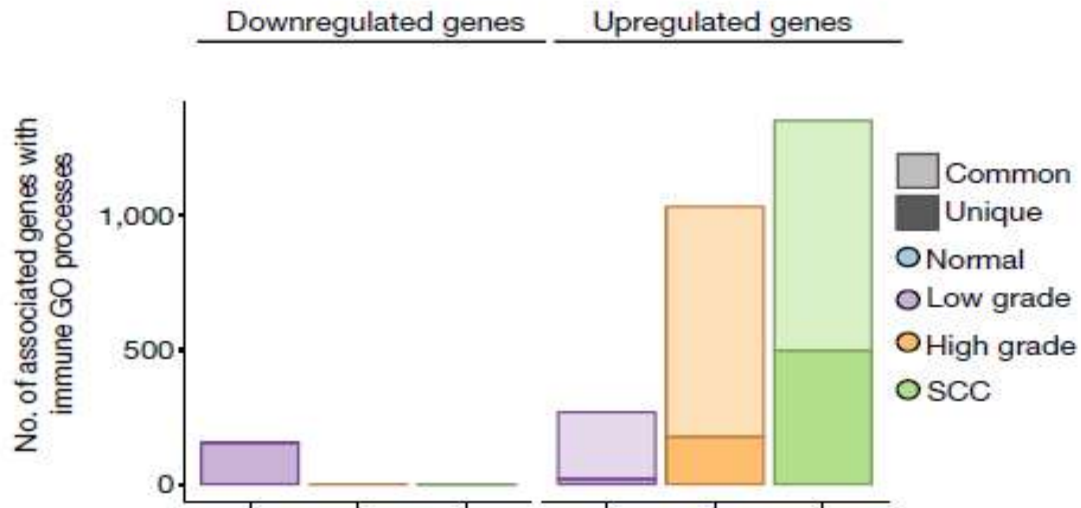
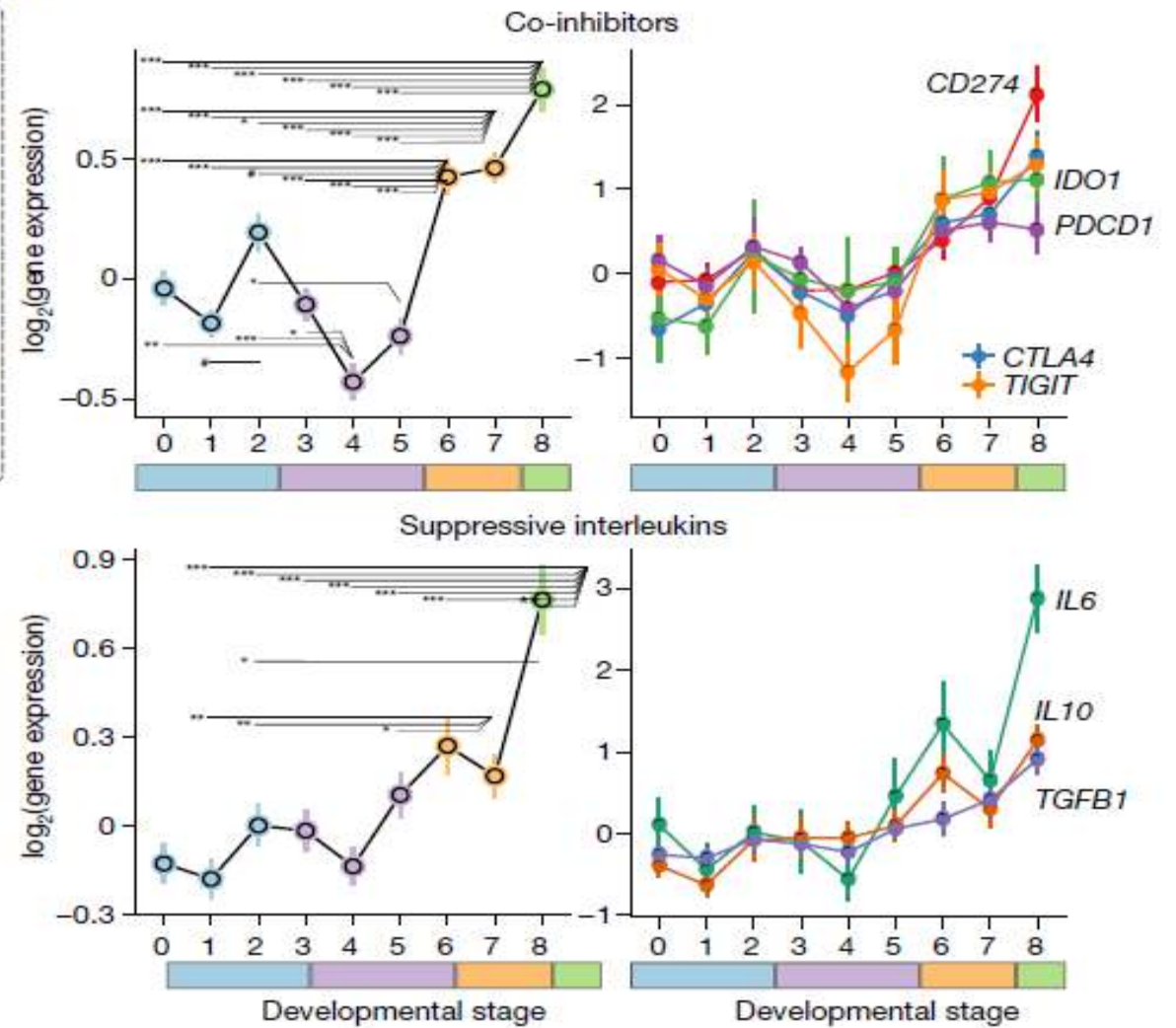


Early inhibition of immune response

a

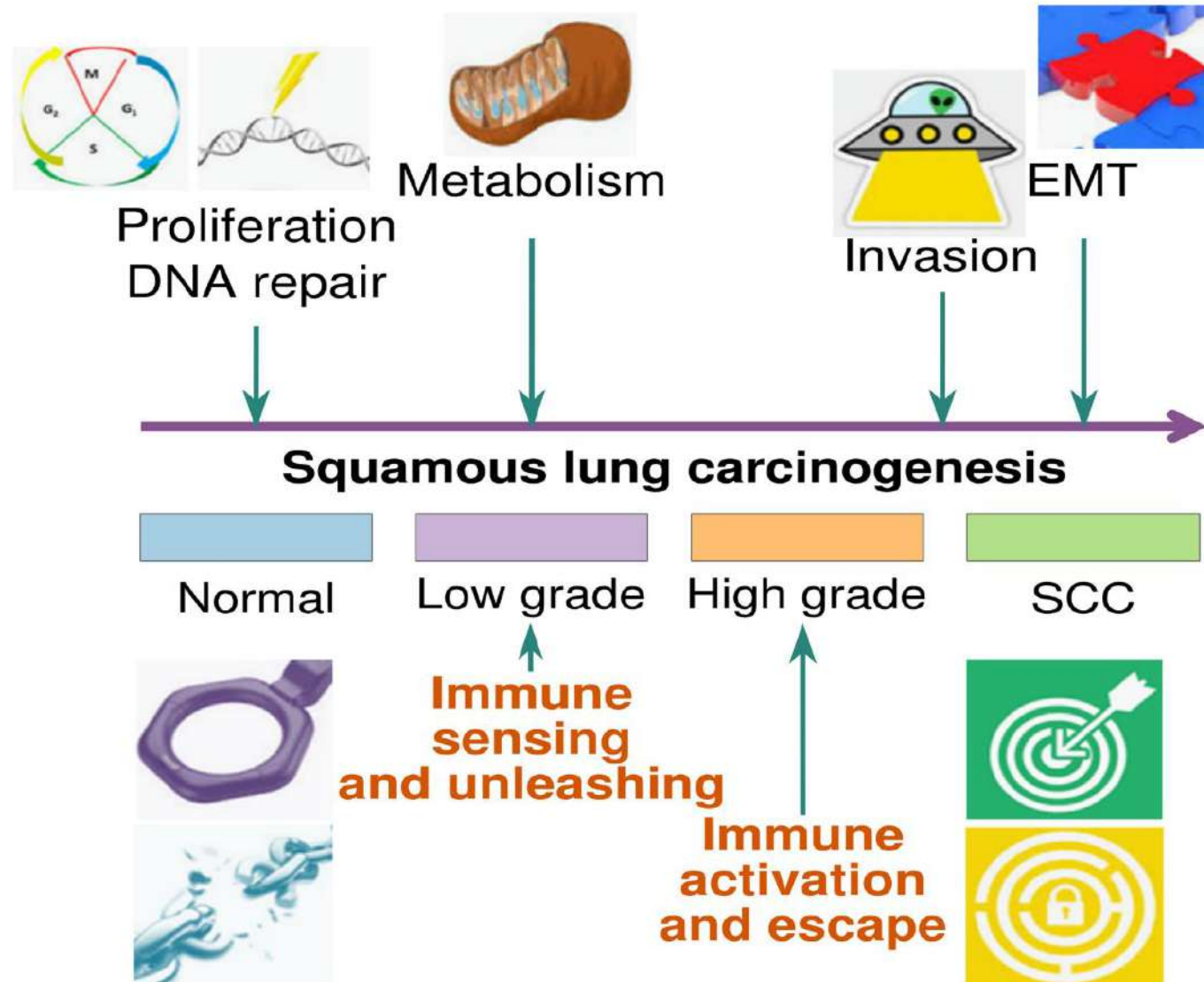


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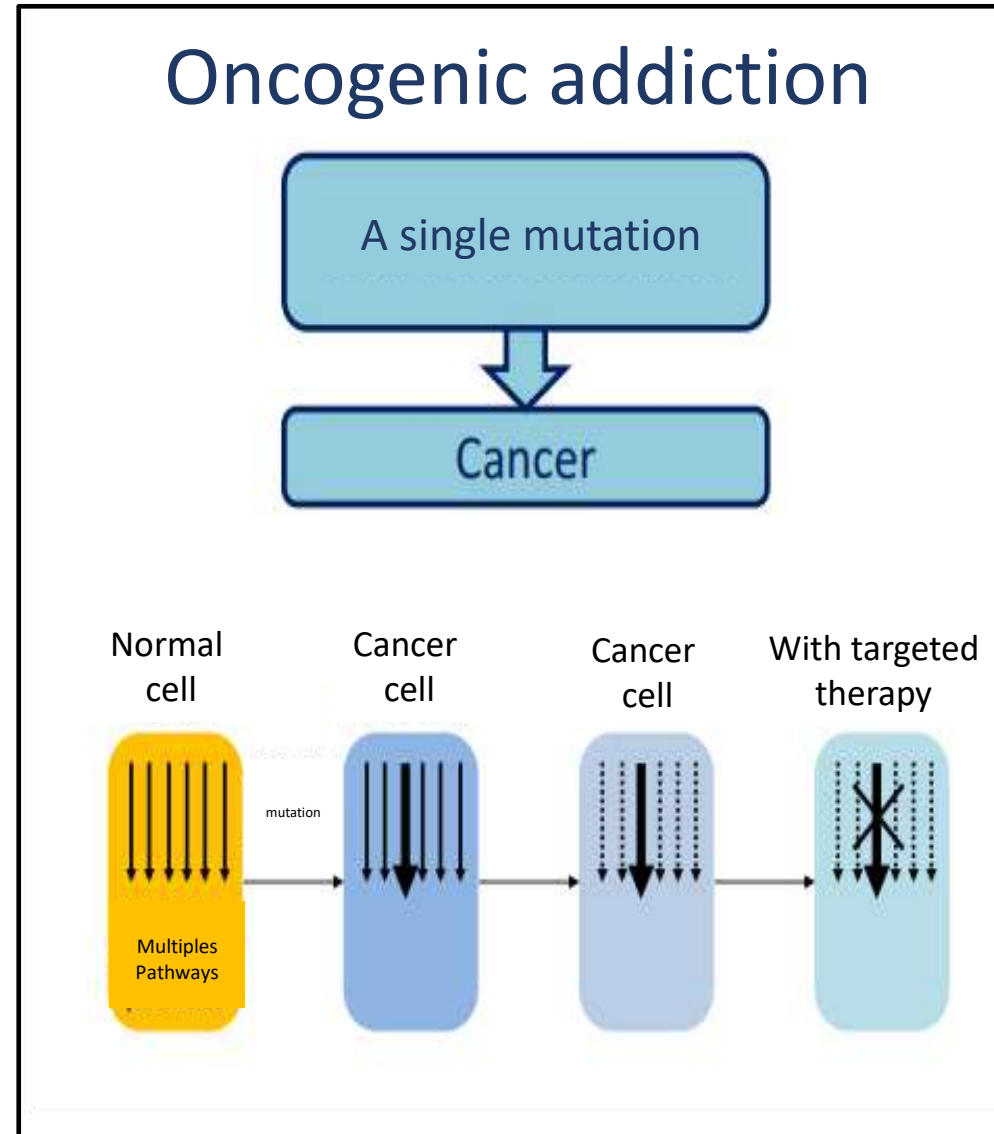


Mascaux et al, Nature 2019

Chronology of molecular pathway of lung carcinogenesis

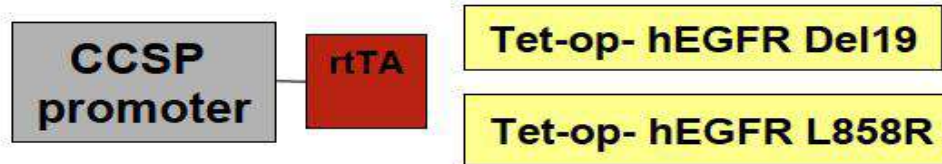


Two models of carcinogenesis



Genetically engineered models of lung cancer

Cell specific inducible hEGFR mutant

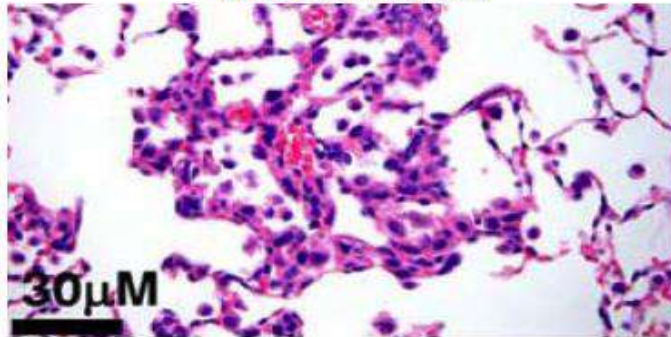


✓ Doxycycline administration
3-4 weeks

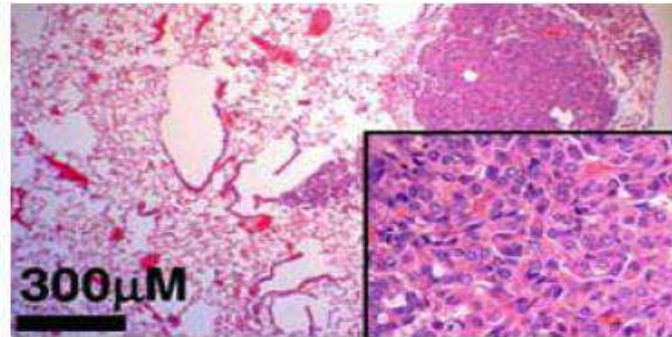
5-6 weeks

8-10 weeks

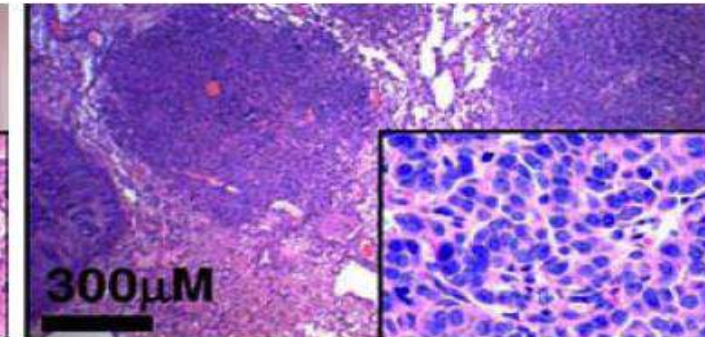
ATYPICAL ADENOMATOUS
HYPERPLASIA



ADENOMA



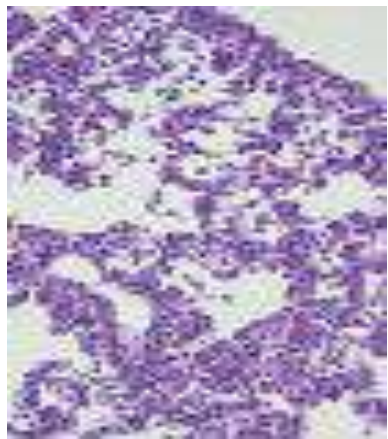
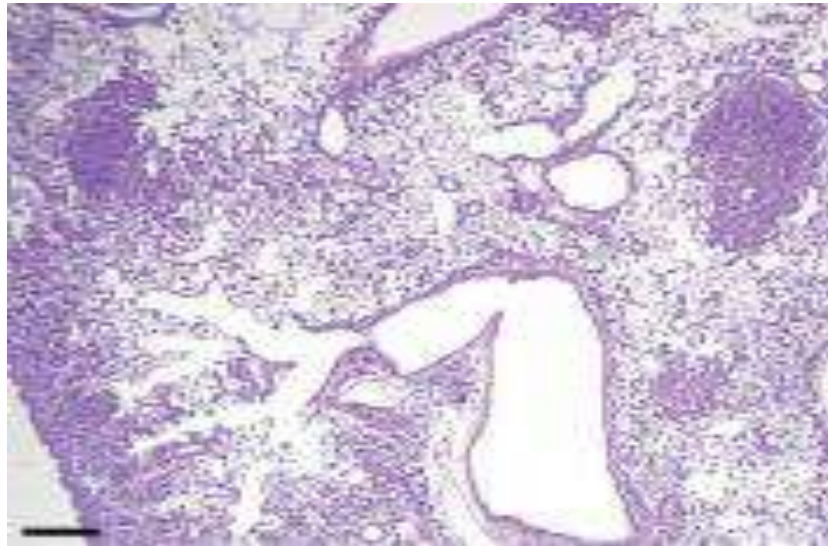
ADENOCARCINOMA



- 100% developed adenocarcinoma with BAC features after 8 or more weeks (SPC+)
- Expression of the hEGFR mutants is essential for tumor maintenance (After 3 weeks of doxycycline withdrawal a complete regression was observed)
- Treatment with TKis (erlotinib or HKI-272) or long term anti-hEGFR antibody (cetuximab) led to tumor regression.

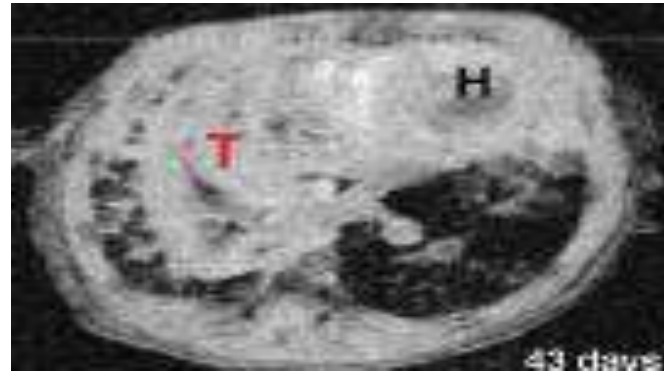
Mouse Lung Adenocarcinomas Induced by Sensitive EGFR Mutations Respond to Down-regulation of the Receptors or Erlotinib

Histopathology



Tumor Response (MRI)

7 weeks ON dox



9 weeks on dox

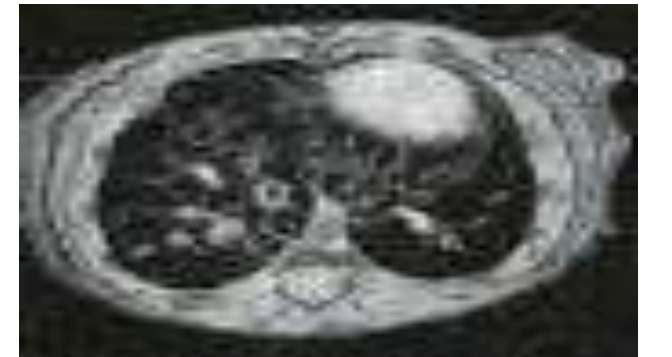


1 week OFF dox

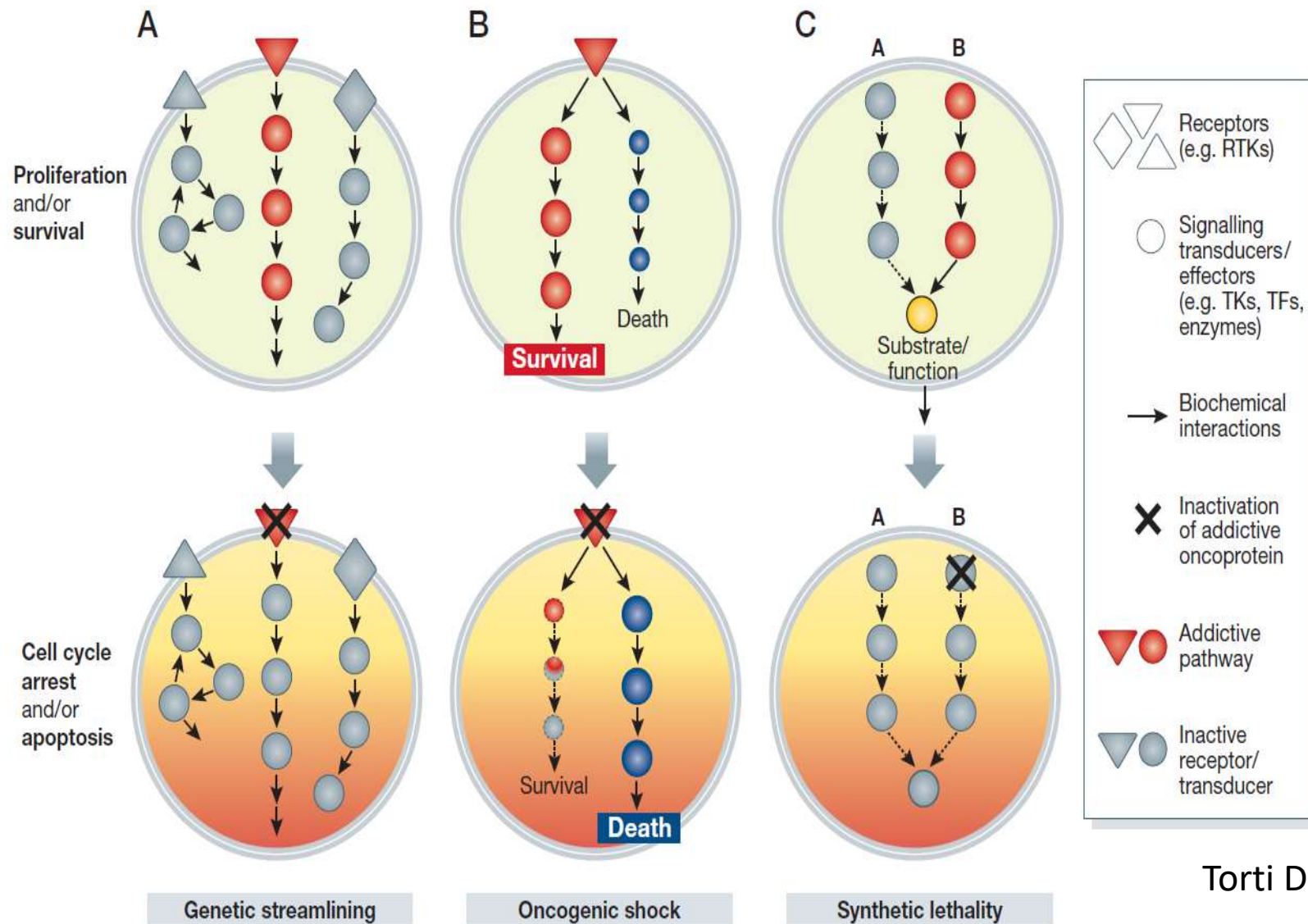


10 weeks on dox

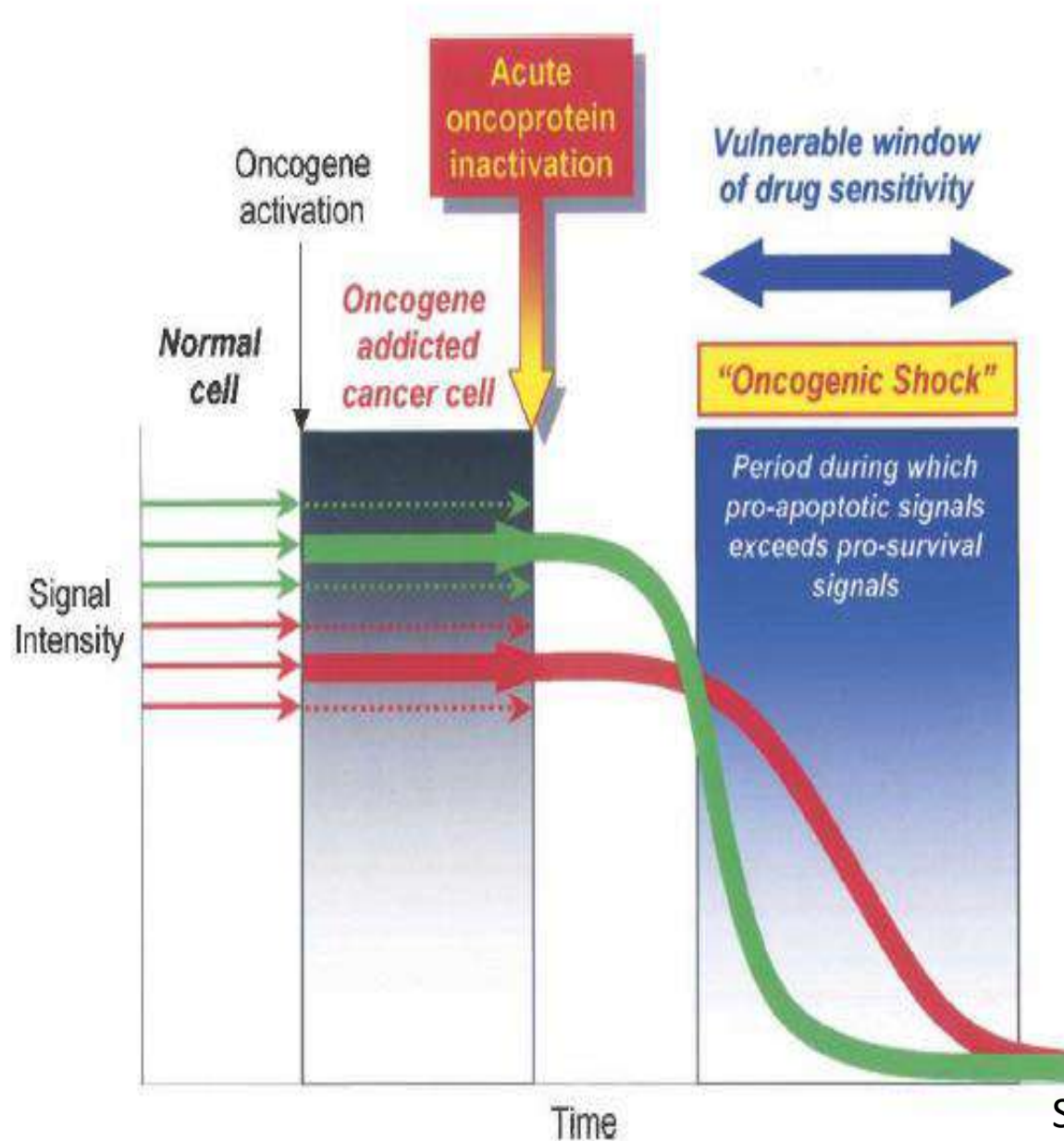
1 week on Erlotinib



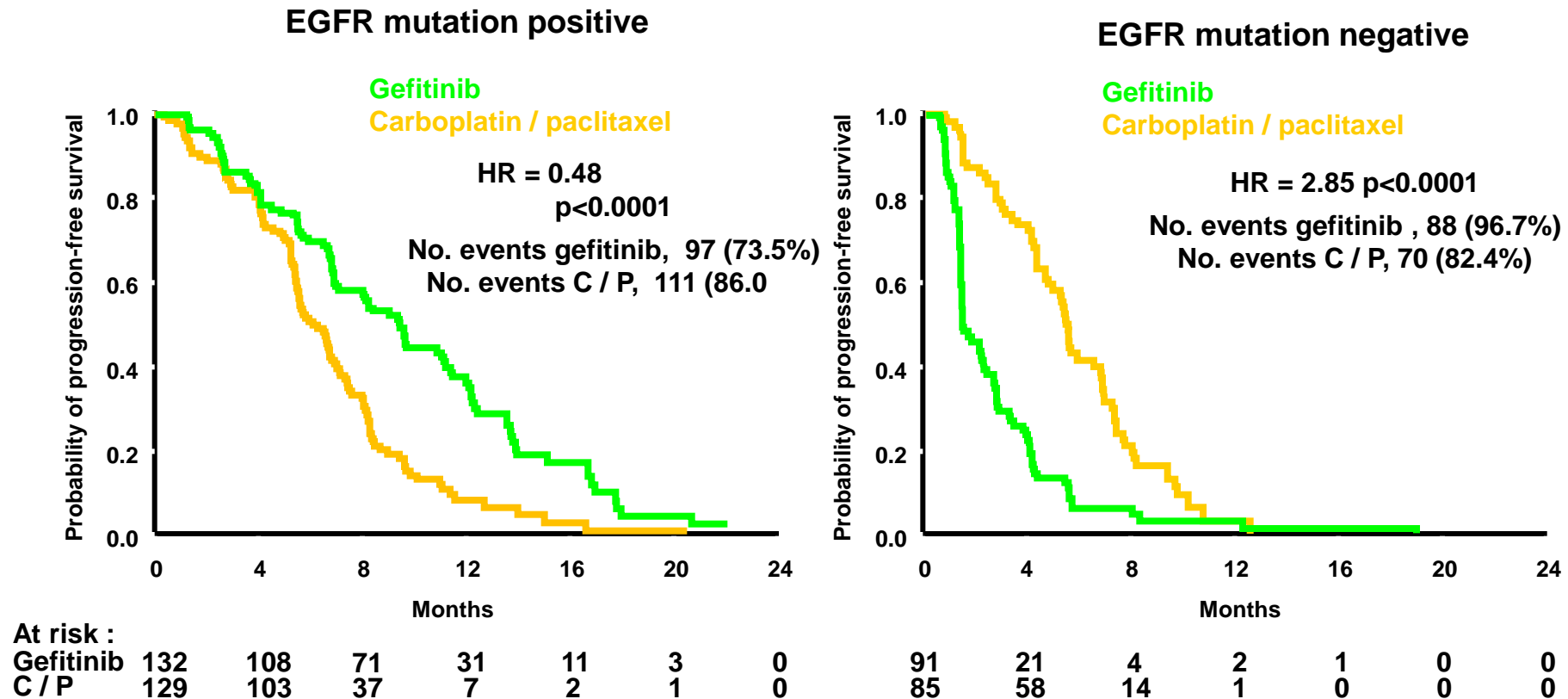
Models of oncogenic addiction



The oncogenic shock



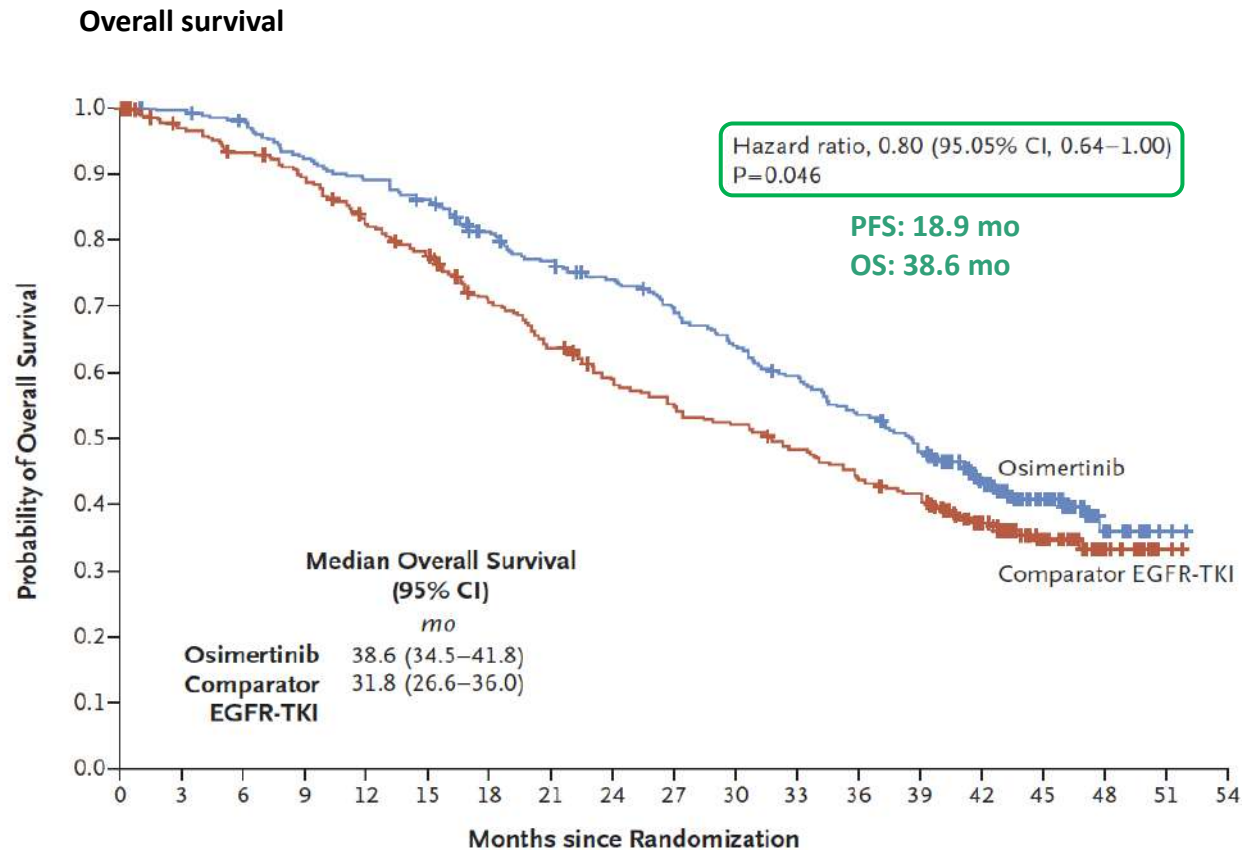
IPASS: Progression-Free Survival in *EGFR* Mutation Positive and Negative Patients



Treatment by subgroup interaction test, p < 0.0001

FLAURA Phase III 1st line treatment, osimertinib vs gefitinib/erlotinib

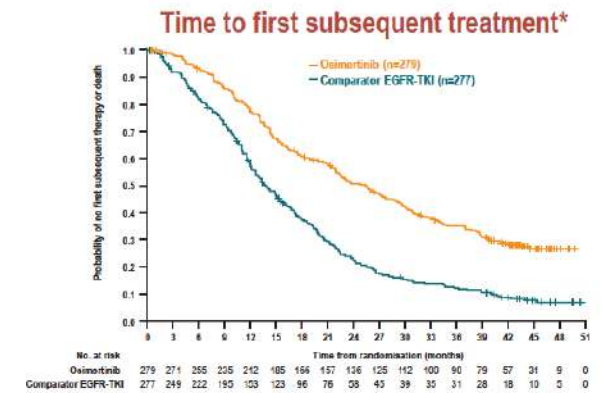
Advanced NSCLC, common EGFR mutation, double blind, amendment for osimertinib cross-over



| No. at Risk | 0 | 3 | 6 | 9 | 12 | 15 | 18 | 21 | 24 | 27 | 30 | 33 | 36 | 39 | 42 | 45 | 48 | 51 | 54 |
|---------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|----|----|----|----|----|
| Osimertinib | 279 | 276 | 270 | 254 | 245 | 236 | 217 | 204 | 193 | 180 | 166 | 153 | 138 | 123 | 86 | 50 | 17 | 2 | 0 |
| Comparator EGFR-TKI | 277 | 263 | 252 | 239 | 219 | 205 | 182 | 165 | 148 | 138 | 131 | 121 | 110 | 101 | 72 | 40 | 17 | 2 | 0 |

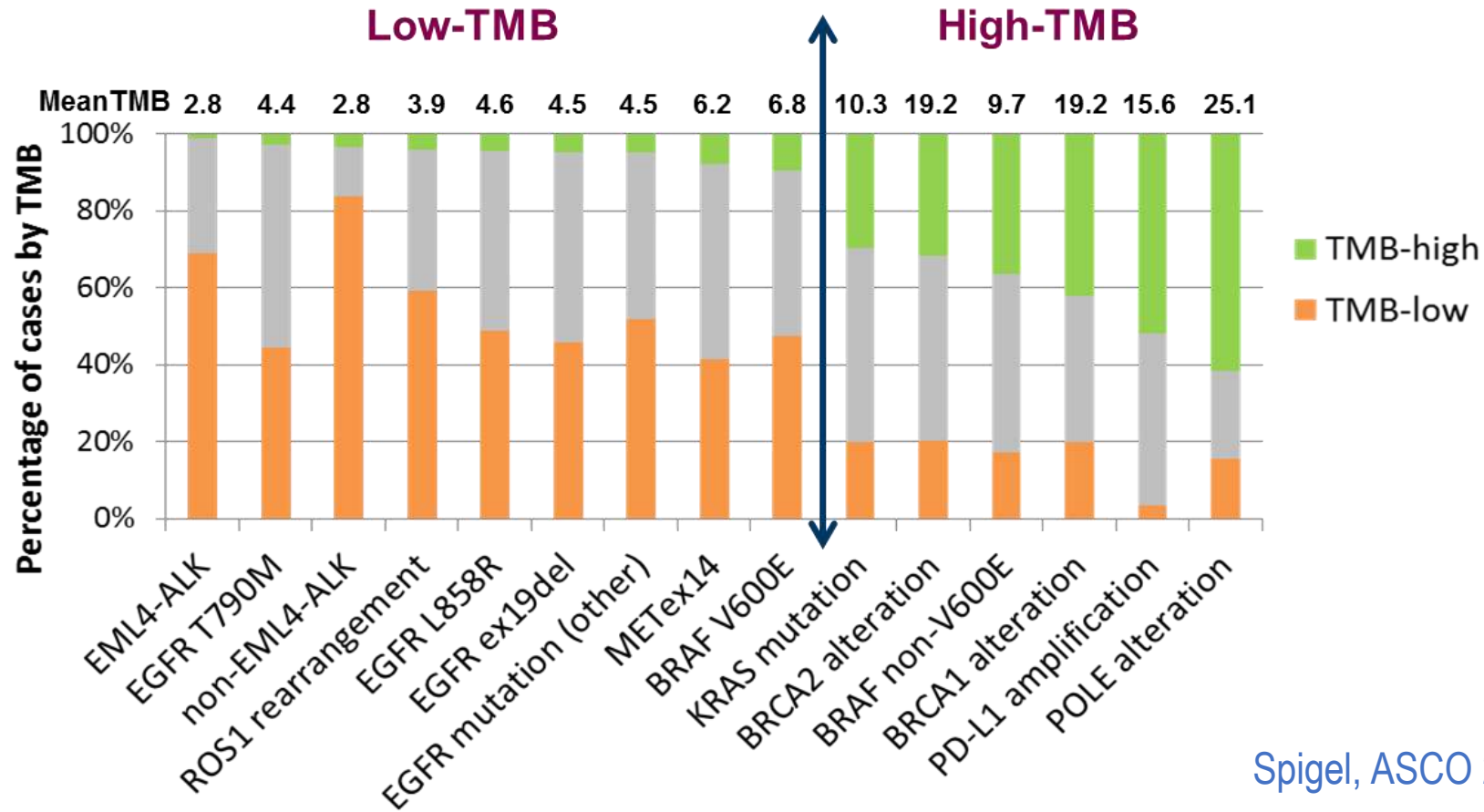
Table 1. Overall Survival and Continuation of First-Line Trial Drug.*

| Variable | Osimertinib (N=279) | Comparator EGFR-TKI (N=277) |
|--|---------------------|-----------------------------|
| Patients continuing to receive first-line trial drug — no. (%) | | |
| At 12 mo | 194 (70) | 131 (47) |
| At 24 mo | 118 (42) | 45 (16) |
| At 36 mo | 78 (28) | 26 (9) |



| Time to first subsequent therapy or death | Events | Median, months (95% CI) |
|---|--------|-------------------------------|
| Osimertinib | 194 | 25.5 (22.0, 29.1) |
| Comparator EGFR-TKI | 242 | 13.7 (12.3, 15.7) |
| HR (95% CI) | | 0.478 (0.393, 0.581) p<0.0001 |

TMB high versus low and oncogenic addiction



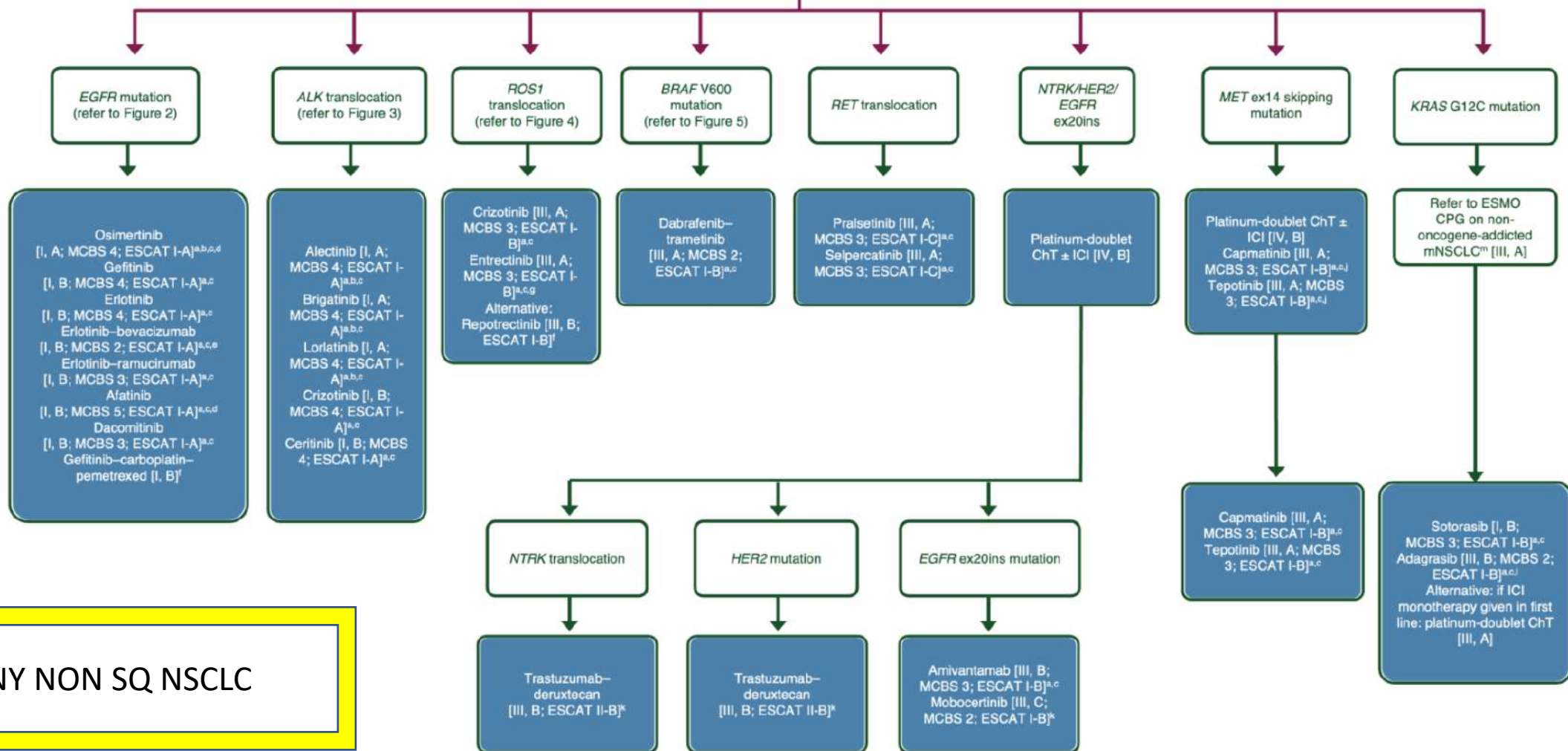
Spigel, ASCO 2016

IN WHOM do we analyse tumours for molecular abnormalities ?

ADVANCED STAGES NSCLC

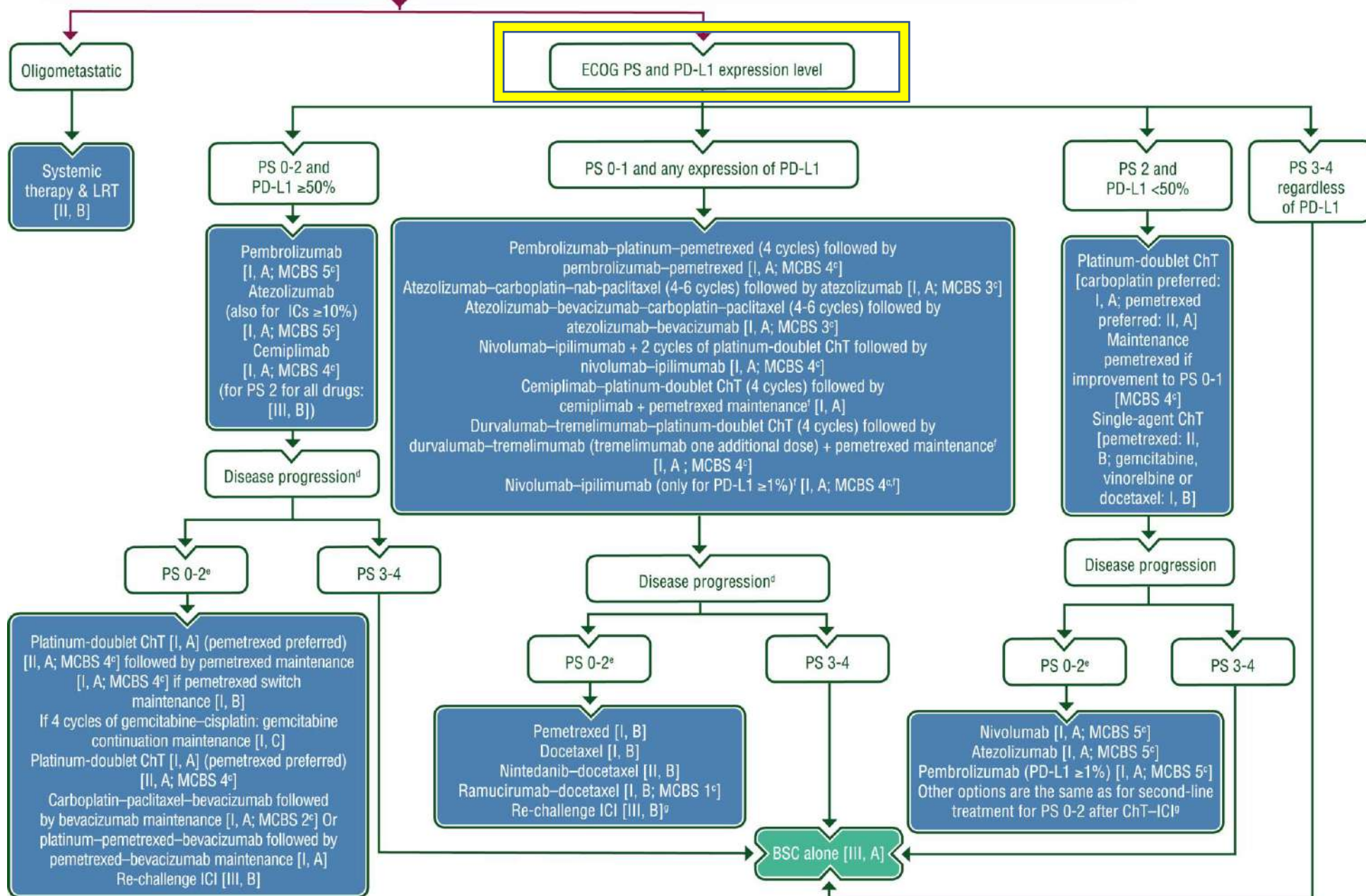
Targeted Therapies in Advanced NSCLC: ESMO guidelines

Stage IV mNSCLC, molecular tests positive (EGFR/ALK/ROS1/BRAF/RET/NTRK/MET/HER2/EGFRex20ins/KRAS G12C)

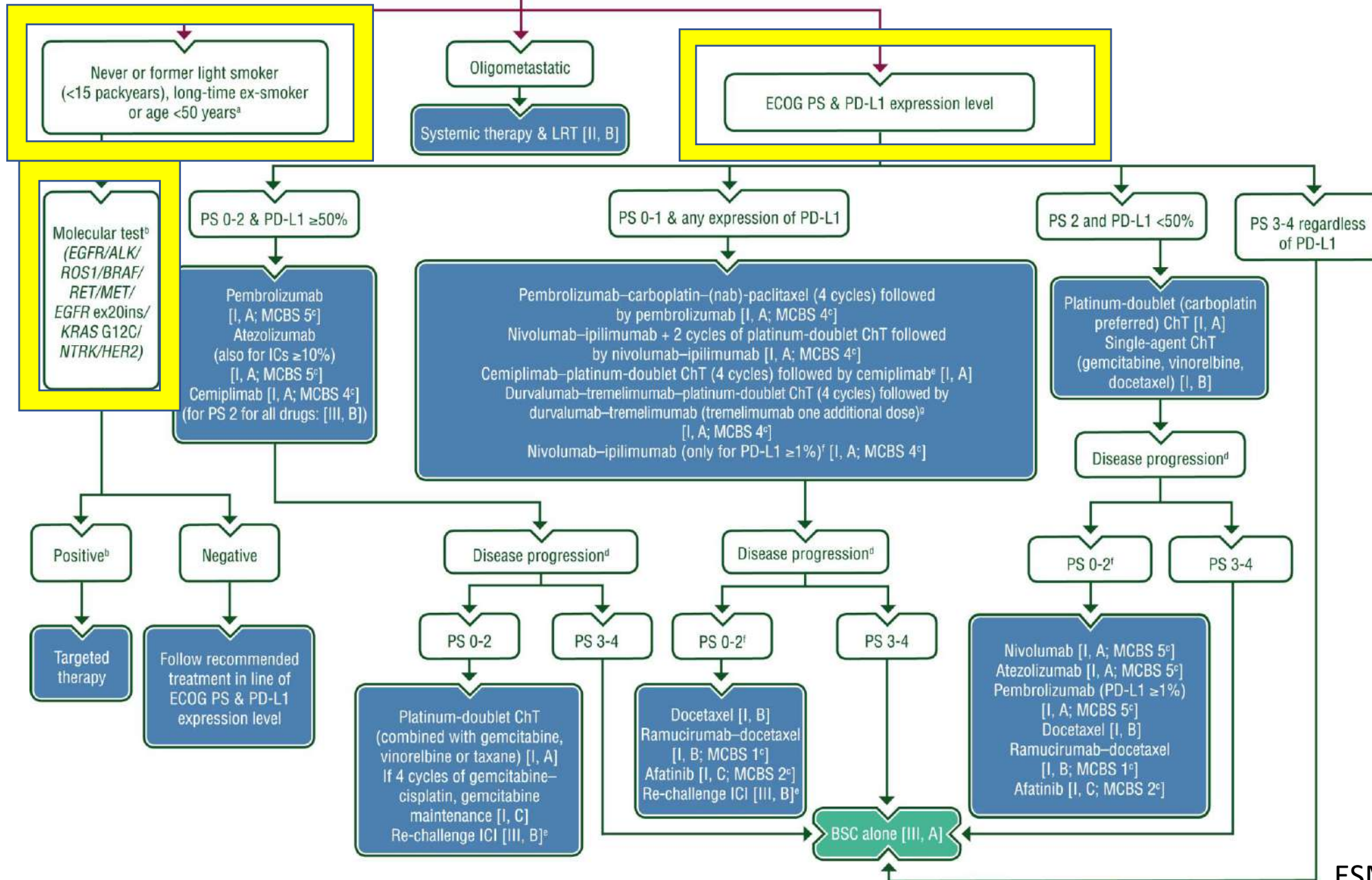


ANY NON SQ NSCLC

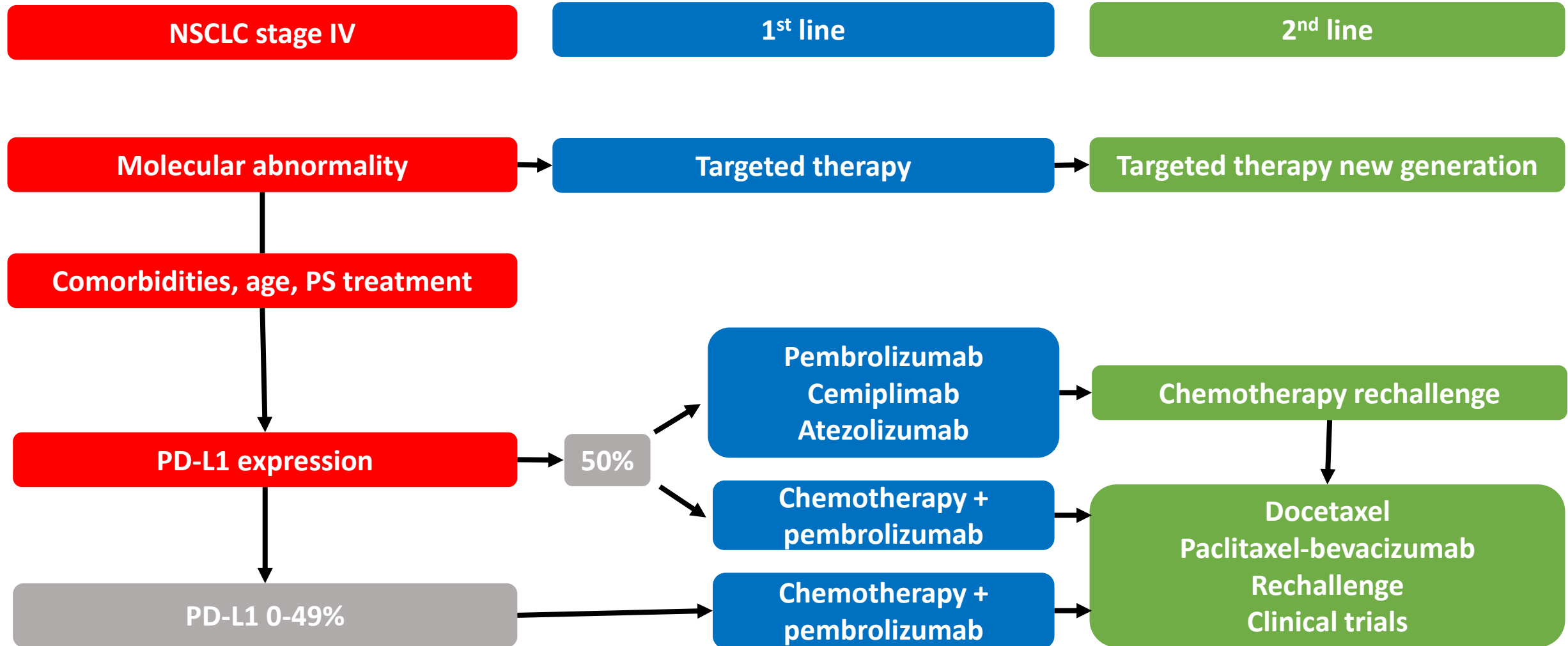
Stage IV NSqNSCC, molecular tests negative (EGFR/ALK/ROS1/BRAF/RET/MET/EGFR ex20ins/KRAS G12C/NTRK/HER2)^{a,b} without contraindication for immunotherapy



Stage IV SqCC without contraindication for immunotherapy



ALGORITHM IN STAGES IV NSCLC



EARLY STAGES NSCLC

Neoadjuvant and Adjuvant Treatments for Early Stage Resectable NSCLC: Consensus Recommendations From the International Association for the Study of Lung Cancer

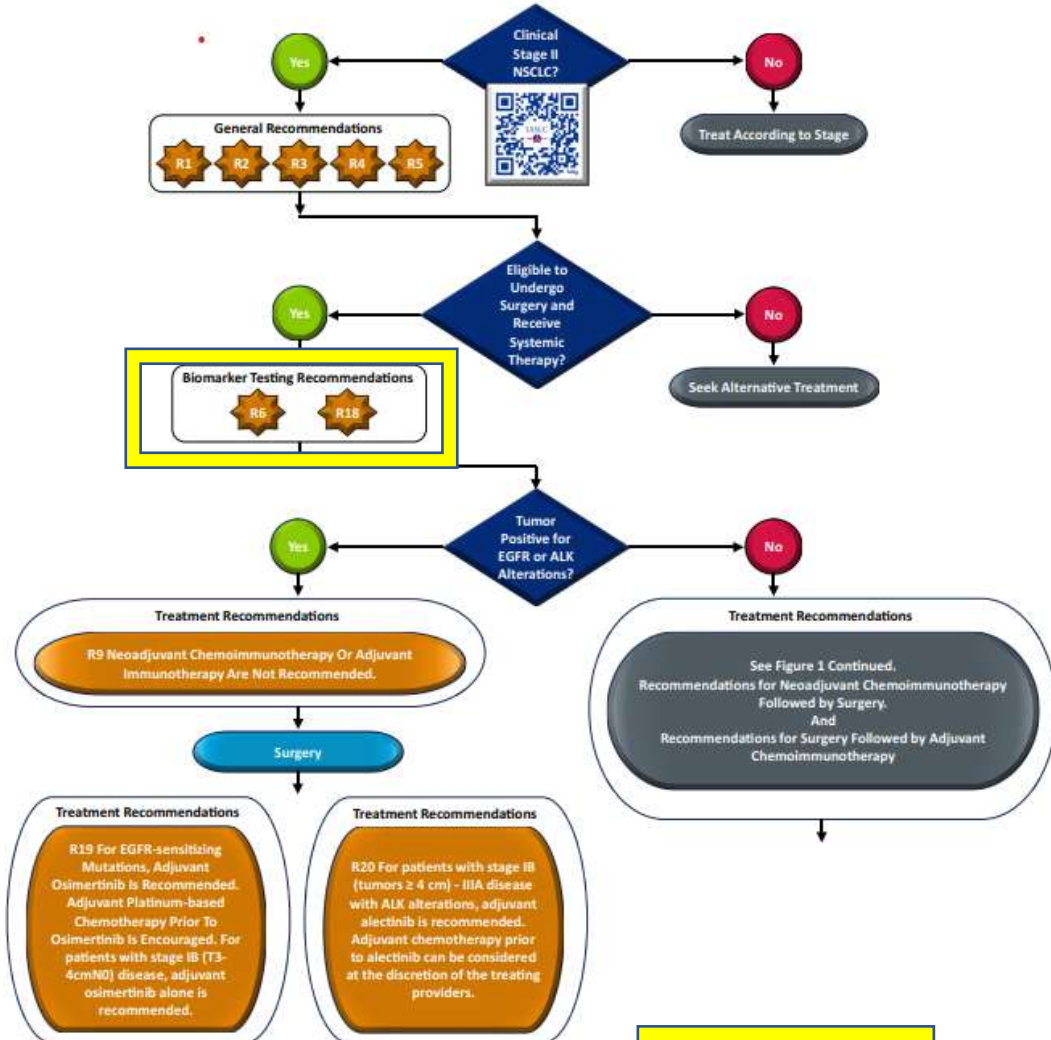
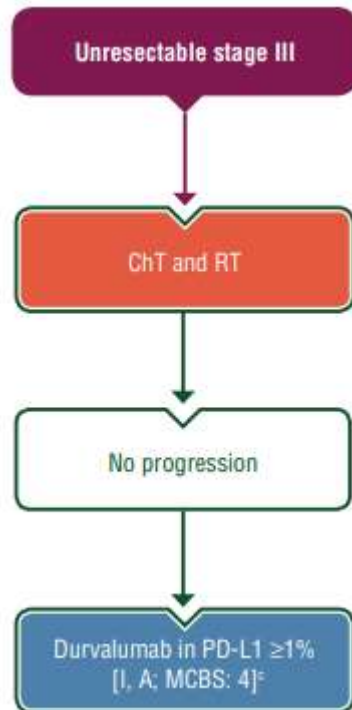


Figure 1. Workflow for neoadjuvant or adjuvant therapy for clinical stage II NSCLC.

Figure 2. Workflow for neoadjuvant or adjuvant therapy for clinical stage III NSCLC.

| | Expert Panel Agreement, % | Open Comment Agreement, % |
|---|---------------------------|---------------------------|
| For patients being considered for neoadjuvant or adjuvant systemic therapy, at a minimum, determination of <i>EGFR</i> and <i>ALK</i> alteration status is required. Tumor proportion score measurement for determination of PD-L1 status should also be considered. | 100 | 93 |
| For patients with TKI-sensitizing <i>EGFR</i> or <i>ALK</i> alterations, neoadjuvant chemoimmunotherapy or adjuvant immunotherapy is not recommended. | 95 | 89 |
| In the light of ongoing trials in populations with specific driver alterations and with extrapolation of the limited efficacy of PD-1 and PD-L1 inhibitors in patients with driver alterations, in addition to assessing <i>EGFR</i> and <i>ALK</i> alteration status, biomarker testing for other oncogenic drivers is highly encouraged in patients with early stage disease. | 94 | 91 |
| For patients with stage II or IIIA disease with <i>EGFR</i> -sensitizing mutations, adjuvant osimertinib is recommended. Adjuvant platinum-based chemotherapy before osimertinib is encouraged. For patients with stage IB (T3-4cmN0) disease, adjuvant osimertinib alone is recommended. | 94 | 92 |
| For patients with stages IB (tumors ≥ 4 cm) to IIIA disease with <i>ALK</i> alterations, adjuvant alectinib is recommended. Adjuvant chemotherapy before alectinib can be considered at the discretion of the treating providers. | 95 | ND |

Unresectable stage III



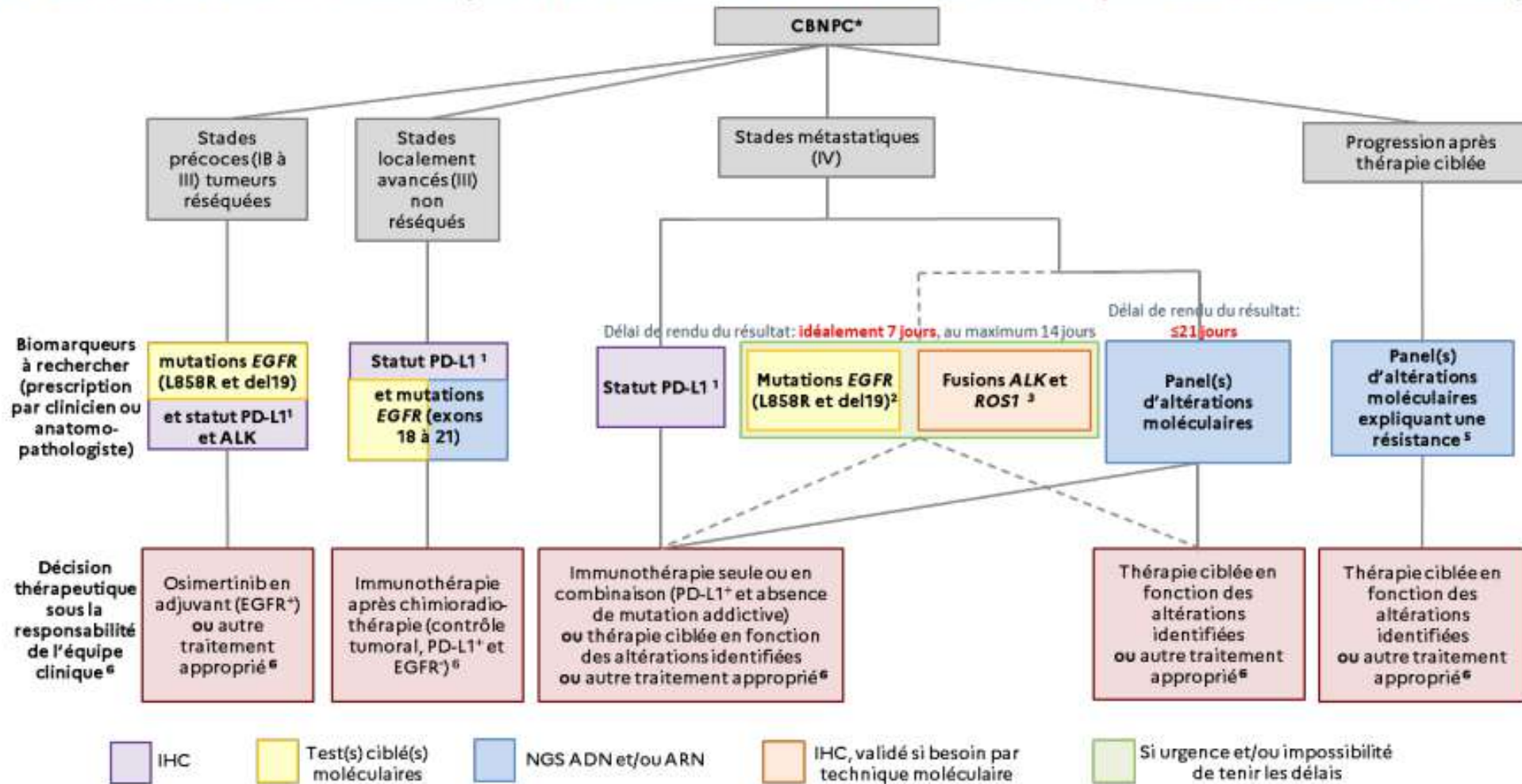
Unresectable locally advanced NSCLC

The phase III PACIFIC trial randomised (2 : 1) 713 patients with unresectable, locally-advanced NSCLC without disease progression within the first 42 days after concurrent chemoradiotherapy, to consolidative durvalumab for 1 year or placebo.⁸ After a median follow-up of 34.2 months, the median OS for durvalumab was reached (47.5 months versus 29.1 months for placebo, HR 0.72, 95% CI 0.59-0.89), and the estimated 5-year OS rates were 42.9% versus 33.4% for durvalumab versus placebo, respectively. The median PFS was 16.9 months for durvalumab and 5.6 months for placebo (HR 0.55, 95% CI 0.45-0.68) with a 5-year PFS rate of 33.1% versus 19.0%, respectively.⁸

A *post hoc* exploratory analysis of the mature survival data requested by licensing European authorities observed that the benefit with durvalumab was not evident in patients with PD-L1 expression <1%. The significance of this observation is disputed.⁹

In France

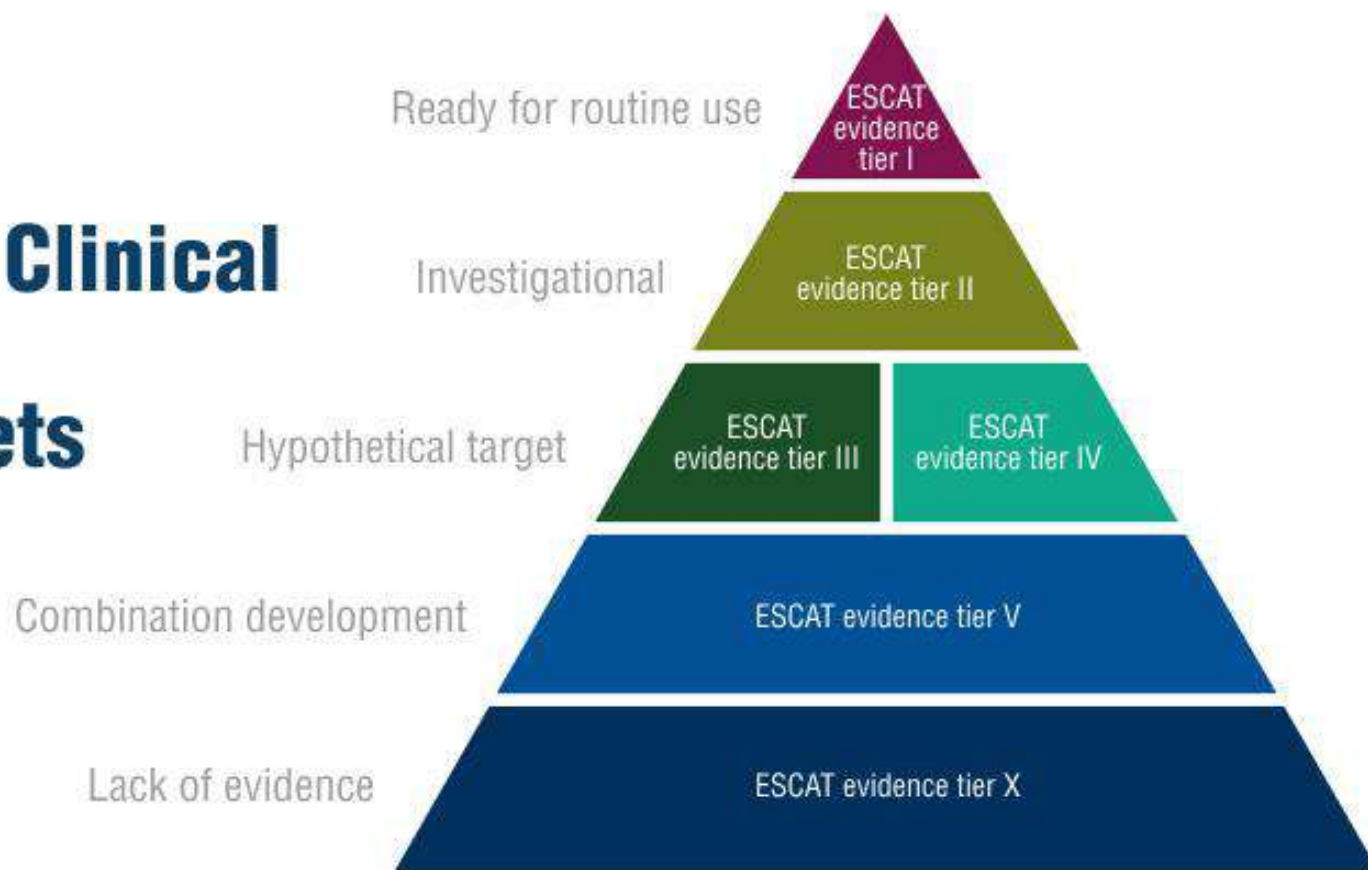
Arbre décisionnel : biomarqueurs nécessaires au traitement des patients atteints de cbnpc



HOW do we analyse tumours for molecular abnormalities ?

ESCAT

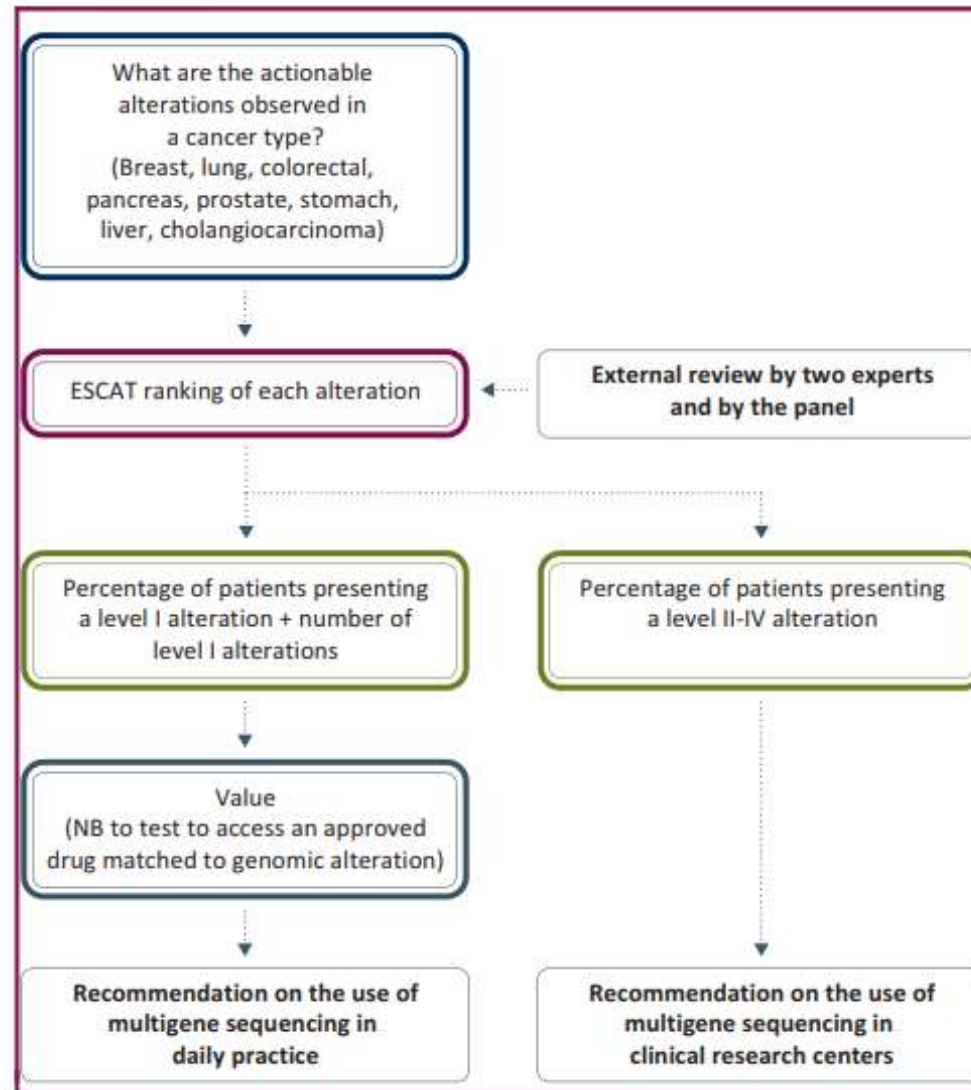
ESMO Scale for Clinical Actionability of Molecular Targets



ESMO GOOD SCIENCE
BETTER MEDICINE
BEST PRACTICE

Ensure precision medicine based on biomarkers analyzed in NGS with a sufficient level of evidence (ESCAT I//II).

Methods to develop recommendation about NGS in daily practice



Which biomarkers do we test in first line for advanced NSCLC

Table 3A. List of genomic alterations level I/II/III according to ESCAT in advanced non-squamous non-small-cell lung cancer (NSCLC)

| Gene | Alteration | Prevalence | ESCAT | References |
|-----------------------|--|--------------------------|-------|---|
| EGFR | Common mutations (<i>Del19, L858R</i>) | 15% (50%–60% Asian) | IA | Midha A, et al. <i>Am J Cancer Res.</i> 2015 ²⁶ |
| | Acquired <i>T790M</i> exon 20 | 60% of EGFR mutant NSCLC | IA | Mok T, et al. <i>J Clin Oncol.</i> 2018 ²⁷ |
| | Uncommon EGFR mutations (<i>G719X</i> in exon 18, <i>L861Q</i> in exon 21, <i>S768I</i> in exon 20) | 10% | IB | Soria J-C, et al. <i>N Engl J Med.</i> 2018 ²⁸ |
| | Exon 20 insertions | 2% | IIB | Ramalingam S, et al. <i>N Engl J Med.</i> 2020 ²⁹ Mok T, et al. <i>N Engl J Med.</i> 2017 ³⁰ Yang J-C-H, et al. <i>Lancet Oncol.</i> 2015 ³¹ Cho J, et al. <i>J Thorac Oncol.</i> 2018 ³² Cardona A, et al. <i>Lung Cancer.</i> 2018 ³³ Heymach J, et al. <i>J Thorac Oncol.</i> 2018 ³⁴ |
| ALK | Fusions (mutations as mechanism of resistance) | 5% | IA | Solomon B, et al. <i>J Clin Oncol.</i> 2018 ³⁵ Soria J-C, et al. <i>Lancet.</i> 2017 ³⁶ Peters S, et al. <i>N Engl J Med.</i> 2017 ³⁷ Zhou C, et al. <i>Ann Oncol.</i> 2018 ³⁸ Camidge D, et al. <i>N Engl J Med.</i> 2018 ³⁹ |
| MET | Mutations <i>ex 14 skipping</i> | 3% | IB | Tong J, et al. <i>Clin Cancer Res.</i> 2016 ⁴⁰ |
| | Focal amplifications (acquired resistance on EGFR TKI in EGFR-mutant tumours) | 3% | IIB | Drilon A, et al. <i>Nat Med.</i> 2020 ⁴¹ Camidge D, et al. <i>J Clin Oncol.</i> 2018 ⁵² |
| BRAF ^{V600E} | Mutations | 2% | IB | Planchard D, et al. <i>Lancet Oncol.</i> 2016 ⁴² Planchard D, et al. <i>Lancet Oncol.</i> 2017 ⁴³ Planchard D, et al. <i>J Clin Oncol.</i> 2017 ⁴⁴ |
| ROS1 | Fusions (mutations as mechanism of resistance) | 1%–2% | IB | Shaw A, et al. <i>N Engl J Med.</i> 2014 ⁴⁵ Shaw A, et al. <i>Ann Oncol.</i> 2019 ⁴⁶ Drilon A, et al. <i>Lancet Oncol.</i> 2020 ⁴⁷ |
| NTRK | Fusions | 0.23%–3% | IC | Drilon A, et al. <i>N Engl J Med.</i> 2018 ⁴⁸ Hong D, et al. <i>Lancet Oncol.</i> 2020 ⁴⁹ Doebele RC, et al. <i>Lancet Oncol.</i> 2020 ⁵⁰ |
| RET | Fusions | 1%–2% | IC | Drilon A, et al. <i>J Thorac Oncol.</i> 2019 ⁵¹ |
| KRAS ^{G12C} | Mutations | 12% | IIB | Barlesi F, et al. <i>Lancet.</i> 2016 ⁵³ Fakih M, et al. <i>J Clin Oncol.</i> 2019 ⁵⁴ |
| ERBB2 | Hotspot mutations Amplifications | 2%–5% | IIB | Hyman D, et al. <i>Nature.</i> 2018 ⁵⁵ Wang Y, et al. <i>Ann Oncol.</i> 2018 ⁵⁶ Tsurutani J, et al. <i>J Thorac Oncol.</i> 2018 ⁵⁷ |
| BRCA 1/2 | Mutations | 1.2% | IIIA | Balasubramaniam S, et al. <i>Clin Cancer Res.</i> 2017 ⁶³ |
| PIK3CA | Hotspot mutations | 1.2%–7% | IIIA | Cancer Genome Atlas Research Network. <i>Nature.</i> 2014 ⁶⁰ Vansteenkiste J, et al. <i>J Thorac Oncol.</i> 2015 ⁶² |
| NRG1 | Fusions | 1.7% | IIIB | Duruisseaux M, et al. <i>J Clin Oncol.</i> 2019 ⁵⁹ |

Which biomarkers do we test in first line for advanced NSCLC

Table 3B. List of genomic alterations level I/II/III according to ESCAT in advanced squamous NSCLC

| Gene | Alteration | Prevalence | ESCAT | References |
|-----------------|-------------------|------------|-------|---|
| <i>NTRK</i> | Fusions | 0.23%–3% | IC | Dylon A, et al. <i>N Engl J Med.</i> 2018 ⁴⁸ Hong D, et al. <i>Lancet Oncol.</i> 2020 ⁴⁹ Doebele RC, et al. <i>Lancet Oncol.</i> 2020 ⁵⁰ |
| <i>PIK3CA</i> | Hotspot mutations | 16% | IIIA | Cancer Genome Atlas Research Network, <i>Nature.</i> 2012 ⁶¹ Vansteenkiste J, et al. <i>J Thorac Oncol.</i> 2015 ⁶² |
| <i>BRCA 1/2</i> | Mutations | 1.2% | IIIA | Balasubramaniam S, et al. <i>Clin Cancer Res.</i> 2017 ⁶³ |

ESCAT. European Society for Medical Oncology (ESMO) Scale for Clinical Actionability of molecular targets.

SPECIAL ARTICLE

Recommendations for the use of next-generation sequencing (NGS) for patients with advanced cancer in 2024: a report from the ESMO Precision Medicine Working Group

M. F. Mosele^{1,2}, C. B. Westphalen³, A. Stenzinger⁴, F. Barlesi^{1,2,5}, A. Bayle^{5,6,7,8}, I. Bièche⁹, J. Bonastre^{7,8}, E. Castro¹⁰, R. Dienstmann^{11,12,13}, A. Krämer^{14,15}, A. M. Czarnecka^{16,17}, F. Meric-Bernstam¹⁸, S. Michiels^{7,8}, R. Miller^{19,20}, N. Normanno²¹, J. Reis-Filho^{22†}, J. Remon², M. Robson²³, E. Rouleau²⁴, A. Scarpa²⁵, C. Serrano¹¹, J. Mateo¹¹ & F. André^{1,2,5*}

Summary of recommendations

No changes have been made to the indication of carrying out tumour NGS in patients with advanced non-squamous NSCLC in daily practice, as the working group has already recommended tumour NGS in these patients. However, with the inclusion of new genomic alterations categorised as ESCAT level I, it is crucial to carefully consider the optimal approach for tumour NGS implementation in the clinical management of patients with advanced non-squamous NSCLC.

ESCAT I/II

| Table 2. List of genomic alterations level I/II according to ESCAT in advanced non-squamous non-small-cell lung cancer | | | | | |
|--|---|---|-------------|---|--|
| Gene | Alteration | Estimated prevalence | ESCAT score | Drug class matched | References |
| EGFR | Common mutations (deletion exon 19, p.L858R) | 15% Caucasian 50% Asian 30% LATAM | IA | First-, second- and third-generation EGFR TKIs EGFR-MET bispecific antibodies + third-generation EGFR TKIs EGFR-MET bispecific antibodies + chemotherapy ± EGFR TKIs (after PD on third-generation EGFR TKIs) | Midha et al., <i>Am J Can Res</i> 2015 ¹² Arrieta et al., <i>J Thorac Oncol</i> 2015 ¹³ Soria et al., <i>N Engl J Med</i> 2018 ¹⁴ Ramalingam et al., <i>N Engl J Med</i> 2020 ¹⁵ Cho et al., <i>Ann Oncol</i> 2023 ¹⁶ |
| | Acquired p.T790M mutation in exon 20 | 60% after first- or second-generation EGFR TKIs | IA | Third-generation EGFR TKIs | Passaro et al., <i>Ann Oncol</i> 2024 ¹⁷ Mok et al., <i>N Engl J Med</i> 2017 ¹⁸ Papadimitrakopoulou et al., <i>Ann Oncol</i> 2020 ¹⁹ |
| | Exon 20 insertions | 2% | IA | EGFR-MET bispecific antibodies or TKIs | Park et al., <i>J Clin Oncol</i> 2021 ²⁰ Zhou et al., <i>N Engl J Med</i> 2023 ²¹ |
| | Uncommon mutations (p.G719 variants in exon 18, p.L861Q in exon 21, p.S768I in exon 20) | 10% | IB | Second- and third-generation EGFR TKIs | Cho et al., <i>J Clin Oncol</i> 2020 ²² Yang et al., <i>Front Oncol</i> 2022 ²³ |
| ALK | Fusions | 5% | IA | ALK TKIs | Mok et al., <i>Ann Oncol</i> 2020 ²⁴ Shaw et al., <i>N Engl J Med</i> 2020 ²⁵ Camidge et al., <i>J Thorac Oncol</i> 2021 ²⁶ Horn et al., <i>JAMA Oncol</i> 2021 ²⁷ Solomon et al., <i>Lancet Respir Med</i> 2023 ²⁸ |
| KRAS | Mutations (p. G12C) | 12% | IA | KRAS ^{G12C} TKIs | Jänne et al., <i>N Engl J Med</i> 2022 ²⁹ de Langen et al., <i>Lancet</i> 2023 ³⁰ |
| RET | Fusions | 1%-2% | IA | RET TKIs | Subbiah et al., <i>Clin Can Res</i> 2021 ³¹ Griesinger et al., <i>Ann Oncol</i> 2022 ³² Drilon et al., <i>J Clin Oncol</i> 2023 ³³ Zhou et al., <i>N Engl J Med</i> 2023 ³⁴ |
| ROS1 | Fusions | 1%-2% | IB | ROS1 TKIs | Shaw et al., <i>Ann Oncol</i> 2019 ³⁵ Shaw et al., <i>Lancet Oncol</i> 2019 ³⁶ Drilon et al., <i>JTO Clin Res Rep</i> 2022 ³⁷ |
| BRAF | Mutations (p. V600E) | 2% | IB | BRAF TKIs + MEK TKIs | Planchard et al., <i>J Thorac Oncol</i> 2022 ³⁸ Riely et al., <i>J Clin Oncol</i> 2023 ³⁹ |
| MET | Mutations exon 14 skipping | 3% | IB | MET TKIs | Drilon et al., <i>Nat Med</i> 2020 ⁴⁰ Wolf et al., <i>J Clin Oncol</i> 2021 ⁴¹ Lu et al., <i>Lancet Respir</i> 2021 ⁴² Thomas et al., <i>J Thorac Oncol</i> 2022 ⁴³ Wolf et al., <i>Ann Oncol</i> 2022 ⁴⁴ |
| | Focal amplifications | 5% as primary 15% as mechanism of acquired resistance on EGFR TKIs | IIB | MET TKIs + third-generation EGFR TKIs EGFR-MET bispecific antibodies + third-generation EGFR TKIs | Yu et al., <i>Ann Oncol</i> 2021 ⁴⁵ Bauml et al., <i>J Clin Oncol</i> 2021 ⁴⁶ Shu et al., <i>J Clin Oncol</i> 2022 ⁴⁷ Marmarelis et al., <i>J Thorac Oncol</i> 2022 ⁴⁸ Hartmaier et al., <i>Cancer Discov</i> 2023 ⁴⁹ Tan et al., <i>J Clin Oncol</i> 2023 ⁵⁰ |
| ERBB2 | Hotspot mutations | 3% | IIB | Pan-HER TKIs Anti-HER2 ADCs | Hyman et al., <i>Nature</i> 2018 ⁵¹ Li et al., <i>N Engl J Med</i> 2022 ⁵² |
| NRG1 | Fusions | <1% | IIB | Anti-HER2/HER3 bispecific antibody | Schram et al., <i>JCO</i> 2022 ⁵³ |

ADC, antibody–drug conjugates; EGFR, epidermal growth factor receptor; ESCAT, ESMO Scale for Clinical Actionability of molecular Targets; HER, human epidermal growth factor receptor; LATAM, Latin America; PD, progressive disease; TKIs, tyrosine kinase inhibitors.

ESCAT I/II

| Table 2. List of genomic alterations level I/II according to ESCAT in advanced non-squamous non-small-cell lung cancer | | | | | |
|---|--|---|-------------|---|--|
| Gene | Alteration | Estimated prevalence | ESCAT score | Drug class matched | References |
| EGFR | Common mutations (deletion exon 19, L858R) | 15% Caucasian 50% Asian 30% LATAM | IA | First-, second- and third-generation EGFR TKIs EGFR-MET bispecific antibodies + third-generation EGFR TKIs | Midha et al., <i>Am J Can Res</i> 2015 ¹² Arrieta et al., <i>J Thorac Oncol</i> 2015 ¹³ Soria et al., <i>N Engl J Med</i> 2018 ¹⁴ |
| <h2 style="text-align: center;">What do the ESMO Guidelines recommend?</h2> <div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <h3>1. <u>Molecular Testing</u></h3> <ul style="list-style-type: none"> - Broad based NGS testing and PD-L1 tumor expression at stage IV diagnosis¹ - RNA-based NGS preferred - Targets should include <i>EGFR</i>, <i>KRAS</i>^{G12C}, <i>BRAF</i>^{V600E}, <i>ERBB2</i>, <i>MET</i> (ex14, amplification), <i>ALK</i>, <i>ROS1</i>, <i>TRK</i>, <i>RET</i> translocation - Plasma cfDNA testing recommended²; reflex to tissue testing if negative </div> <div style="width: 45%;"> <h3>2. <u>Treatment</u>^{3,4}</h3> <p style="text-align: center;">ICI + Chemo (PD-L1 1%)</p> </div> </div> | | | | | |
| | Focal amplifications | 5% as primary 15% as mechanism of acquired resistance on EGFR TKIs | IIB | MET TKIs + third-generation EGFR TKIs EGFR-MET bispecific antibodies + third-generation EGFR TKIs | Wong et al., <i>Ann Oncol</i> 2022 ⁴⁵ Yu et al., <i>Ann Oncol</i> 2021 ⁴⁵ Bauml et al., <i>J Clin Oncol</i> 2021 ⁴⁶ Shu et al., <i>J Clin Oncol</i> 2022 ⁴⁷ Marmarelis et al., <i>J Thorac Oncol</i> 2022 ⁴⁸ Hartmaier et al., <i>Cancer Discov</i> 2023 ⁴⁹ Tan et al., <i>J Clin Oncol</i> 2023 ⁵⁰ |
| | ERBB2 Hotspot mutations | 3% | IIB | Pan-HER TKIs Anti-HER2 ADCs | Hyman et al., <i>Nature</i> 2018 ⁵¹ Li et al., <i>N Engl J Med</i> 2022 ⁵² |
| | NRG1 Fusions | <1% | IIB | Anti-HER2/HER3 bispecific antibody | Schram et al., <i>JCO</i> 2022 ⁵³ |

ADC, antibody–drug conjugates; EGFR, epidermal growth factor receptor; ESCAT, ESMO Scale for Clinical Actionability of molecular Targets; HER, human epidermal growth factor receptor; LATAM, Latin America; PD, progressive disease; TKIs, tyrosine kinase inhibitors.

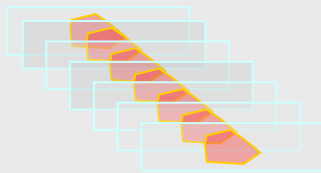


Bloc selection by the pathologist

HES : % CT



Small pieces 10µ



slides

Nucleic acid extraction
Molecular biology

FISH

DNA

DNA
RNA

IHC

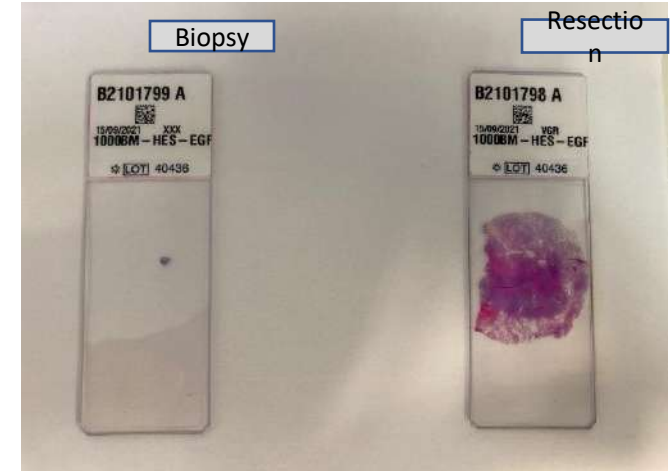
Protein expression

Analyses of mutations/fusions/amplifications

Quality of the tissue (fixation, treatment, time since sampling)

Quantity of available material

Tumour cellularity (%)



With sequential approach of testing
Risk of lack of tissue

For small sample, prefer high throughput techniques (NGS +++)

Biomarkers, molecular targets and methods for testing according to ESCAT scores

| Biomarker or genomic alteration | Method of detection | Drug match | ESCAT score ^{a,b} |
|--|------------------------------|---|----------------------------|
| <i>EGFR</i> mutations (ex19 deletion or exon 21 L858R) ⁵⁵⁻⁶⁷ | Sanger sequencing or NGS | EGFR TKI | I-A |
| <i>EGFR</i> T790M resistance mutation ⁶⁸ | Sanger sequencing or NGS | EGFR TKI | I-A |
| Non-exon 19 deletion/exon 21 L858R sensitising <i>EGFR</i> mutations ^{69, 70} | Sanger sequencing or NGS | EGFR TKI | I-B |
| <i>EGFR</i> exon 20 insertion mutations ^{71, 72} | Sanger sequencing or NGS | EGFR TKI EGFR MET Bispecific antibody | I-B |
| <i>ALK</i> rearrangements ^{70, 73-77} | IHC FISH RNA-based NGS | ALK TKI | I-A |

| | | | |
|---|--|-------------------------|------|
| <i>ROS1</i> rearrangements ⁷⁸⁻⁸¹ | IHC (only screening) FISH RNA-based NGS | ROS1 TKI | I-B |
| <i>BRAF</i> V600 mutations ⁸² | Sanger sequencing or NGS | BRAF + MEK TKI | I-B |
| <i>RET</i> rearrangements ^{83, 84} | FISH RNA-based NGS (preferably) | RET TKI | I-C |
| <i>MET</i> exon 14 skipping mutations ^{85, 86} | NGS (RNA-based NGS might detect more cases compared with DNA-based NGS) | MET TKI | I-B |
| <i>MET</i> amplification ⁸⁵ | IHC ISH | MET TKI | II-B |
| <i>HER2</i> mutations ^{87, 88} | NGS | Antibody–drug conjugate | II-B |

| | | | |
|--|------------------------------------|----------------------------|-------|
| <i>KRAS</i> G12C mutations ⁸⁹ | NGS | <i>KRAS</i> G12C inhibitor | I-B |
| <i>NTRK</i> rearrangements ^{90, 91} | IHC for screening RNA-based NGS | NTRK inhibitors | I-C |
| <i>NRG1</i> fusions ⁹² | RNA-based NGS | HER inhibitors | III-A |

Limits of detection

Variants detected

Real-Time PCR



1%–5%

Only hotspot mutations (probe-based)




Digital PCR

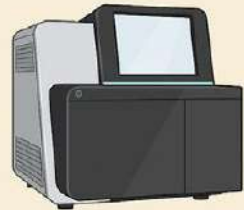


0.1%–1%

Only hotspot mutations (probe-based)




NGS



0.01%–5%

All the mutations present in the analyzed gene regions



Only if NGS not available

Recommended

REVIEW

Recommendations for the use of next-generation sequencing (NGS) for patients with metastatic cancers: a report from the ESMO Precision Medicine Working Group

F. Mosele¹, J. Remon², J. Mateo³, C. B. Westphalen⁴, F. Barlesi¹, M. P. Lolkema⁵, N. Normanno⁶, A. Scarpa⁷, M. Robson⁸, F. Meric-Bernstam⁹, N. Wagle¹⁰, A. Stenzinger¹¹, J. Bonastre^{12,13}, A. Bayle^{1,12,13}, S. Michiels^{12,13}, I. Bièche¹⁴, E. Rouleau¹⁵, S. Jezdic¹⁶, J-Y. Douillard¹⁶, J. S. Reis-Filho¹⁷, R. Dienstmann¹⁸ & F. André^{1,19,20*}

Table 2. Summary recommendations

| Tumour types | General recommendations for daily practice | Recommendation for clinical research centres | Special considerations for patients |
|----------------------------|---|---|--|
| Lung adenocarcinoma | <u>Tumour multigene NGS</u> to assess level I alterations. <u>Larger panels</u> can be used only on the basis of <u>specific agreements with payers</u> taking into account the overall cost of the strategy (drug included ^a) and if they report accurate ranking of alterations. NGS can either be done on RNA or DNA, if it includes level I fusions in the panel. | It is highly recommended that clinical research centres perform multigene sequencing in the context of molecular screening programmes in order to increase access to innovative drugs and to speed up clinical research. This is particularly relevant in breast, pancreatic and hepatocellular cancers where level II–IV alterations are numerous. | Using large panels of genes could lead to few clinically meaningful responders, not detected by small panels or standard testings. In this context and outside the diseases where large panels of genes are recommended, ESMO acknowledges that a patient and a doctor could decide together to order a large panel of genes, pending no extra cost for the public health care system, and if the patient is informed about the low likelihood of benefit. |
| Squamous cell lung cancers | No current indication for tumour multigene NGS | | |
| Breast cancers | No current indication for tumour multigene | | |

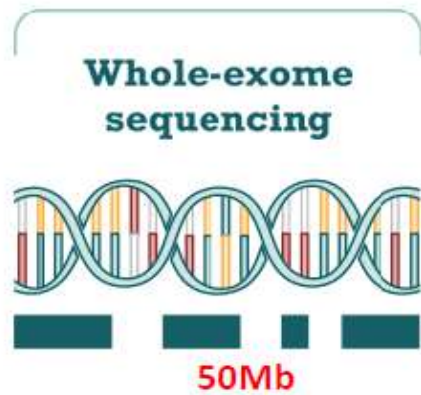
NGS approaches

ADN

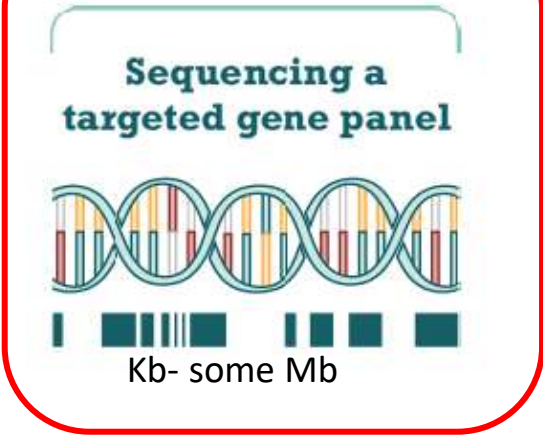
WGS



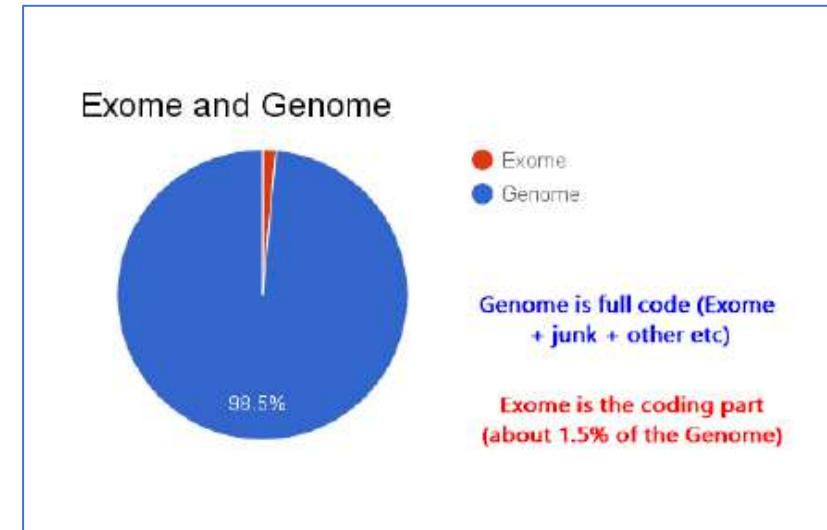
WES



Panel of genes of DNA

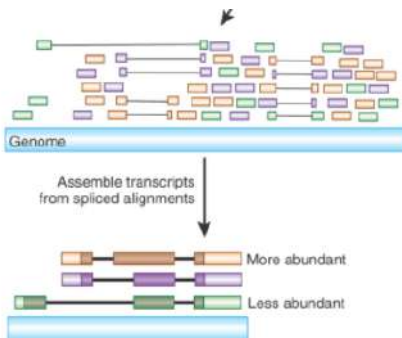


Variable size
(30-500 genes)



ARN

RNASeq



Targetted RNASeq

Panel of genes

Gene targets

| | | | | | | |
|-------|-------|-------|-------|-------|--------|--------|
| AKT1 | ALK | AXL | BRAF | CALCA | CCND1 | CTNNB1 |
| DDR2 | EGFR | ENBR2 | FGFR1 | FGFR2 | FGFR3 | GNAS |
| HRAS | IDH1 | IDH2 | KRAS | KRT20 | KRT7 | MAP2K1 |
| MET | NF1AS | NRG1 | NTRK1 | NTRK2 | NTRK3 | PBCICA |
| PPARG | PTH | RAF1 | RET | ROS1 | SLCSA5 | THADA |
| TTF1 | | | | | | |

Legend:

- SNV/indel
- Expression
- Fusion, splicing or exon-skipping
- Internal tandem duplication (ITD)
- CNV

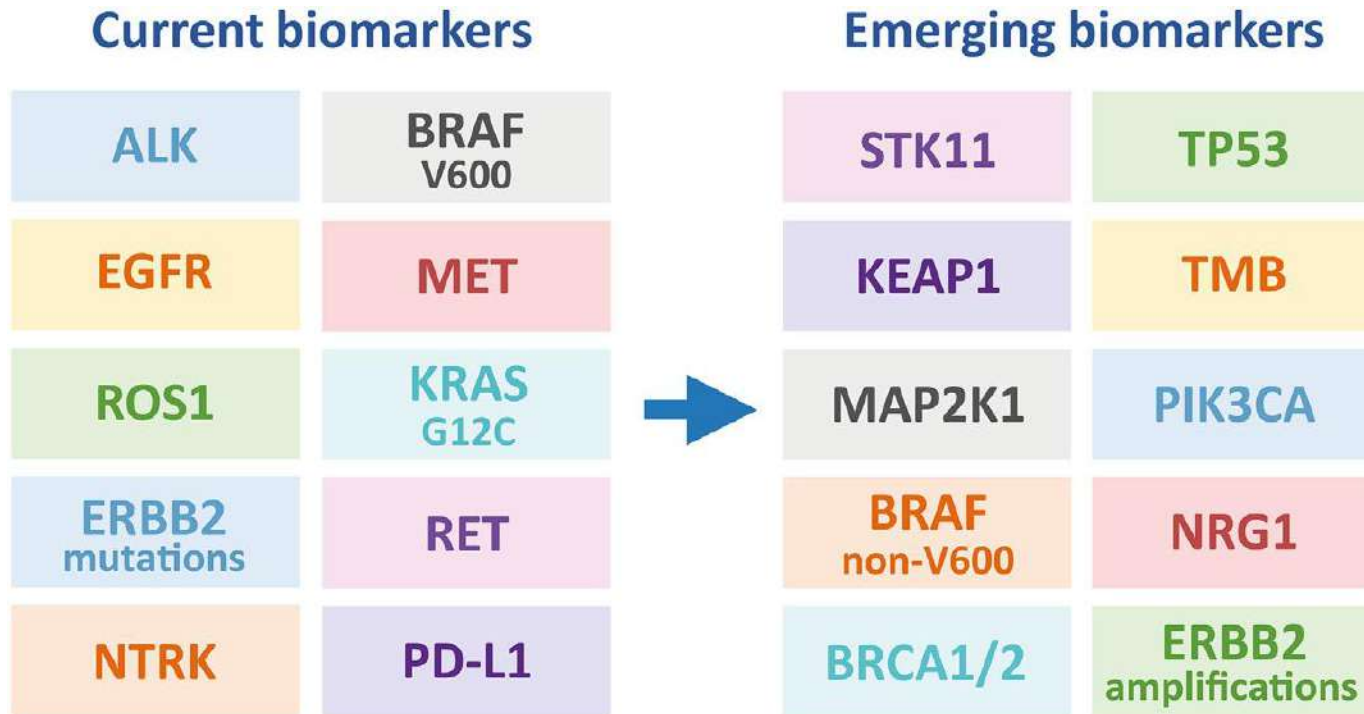
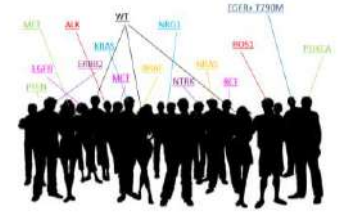
Advantages and disadvantages of the different next-generation sequencing methods for detection of fusion genes

| Library | Nucleic acid | Input* | Genetic alterations detected |
|-------------------------------|--------------|----------------------|--|
| Hybrid capture based | DNA | High (≥ 50 ng) | Break points of translocations (limited if long introns) |
| Hybrid capture based | RNA | High (≥ 40 ng) | Fusions (partner independent) |
| Amplicon based | RNA | Low (≥ 10 ng) | Fusions (only known fusion partners) |
| Anchored Multiplex Sequencing | RNA | Low (≥ 10 ng) | Fusions (partner independent) |

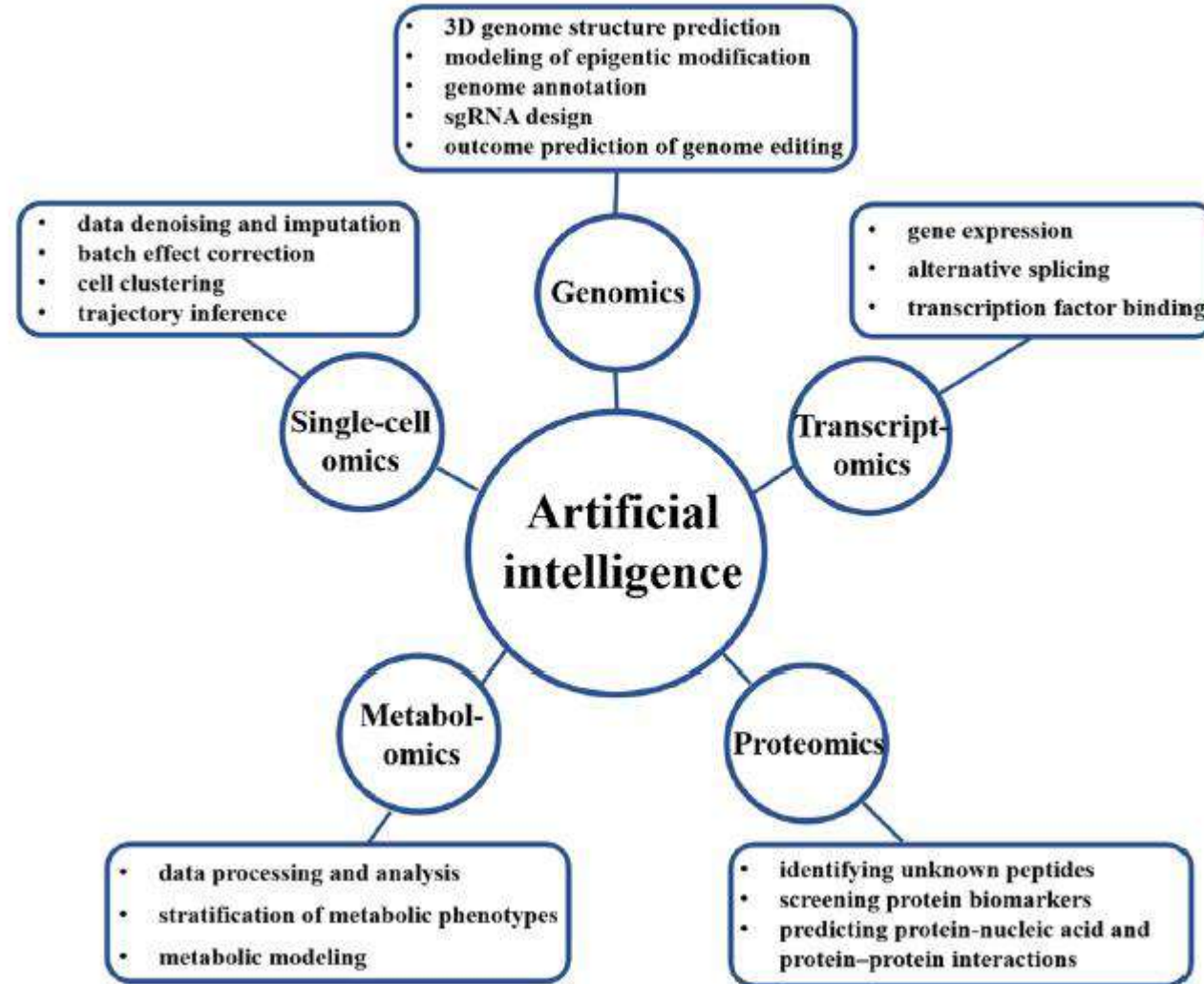
* Minimum nucleic acid input suggested by manufacturers. Higher input is recommended when possible to increase the sensitivity of the test.

And the future ?

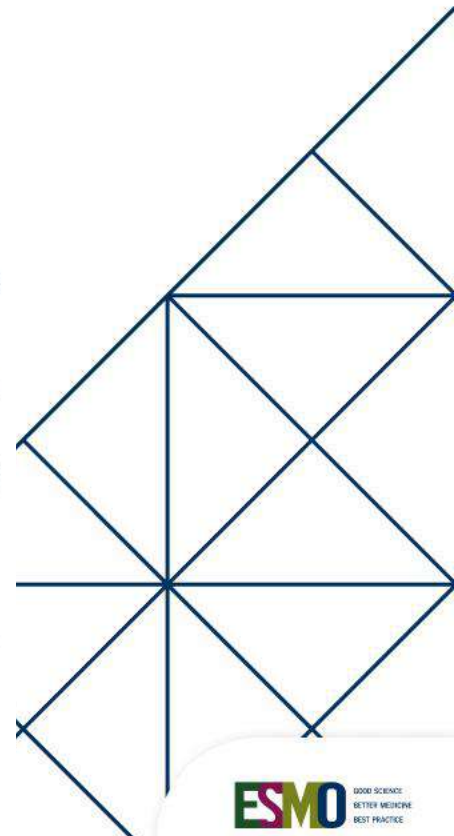
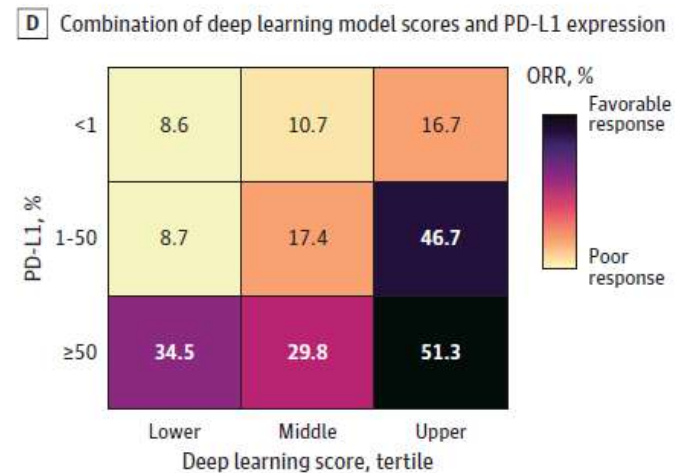
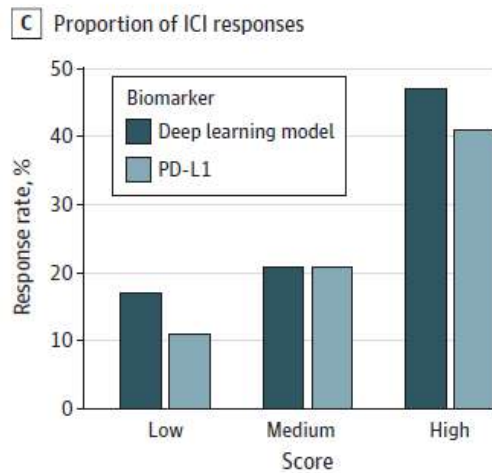
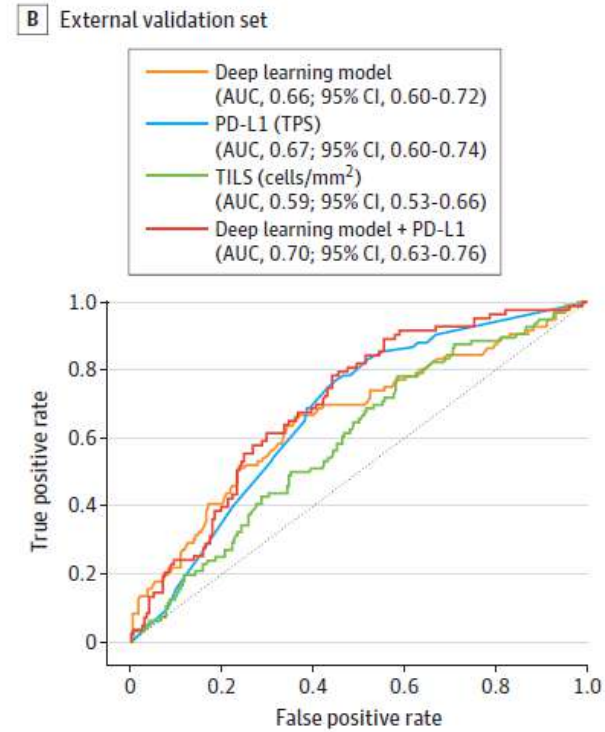
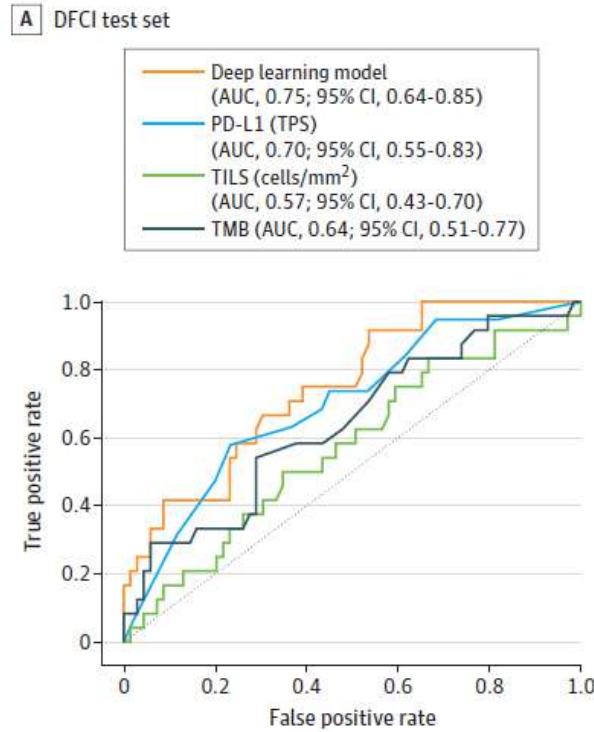
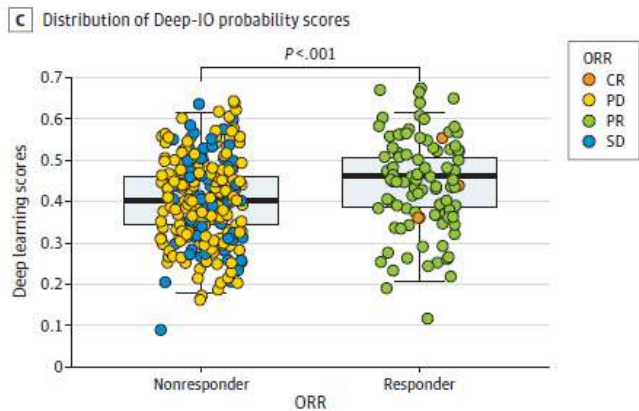
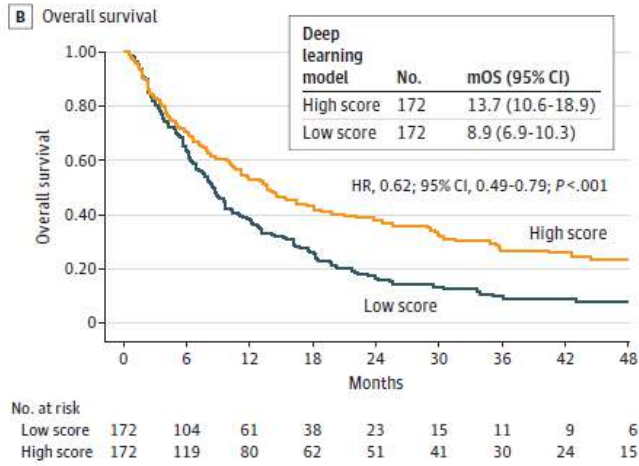
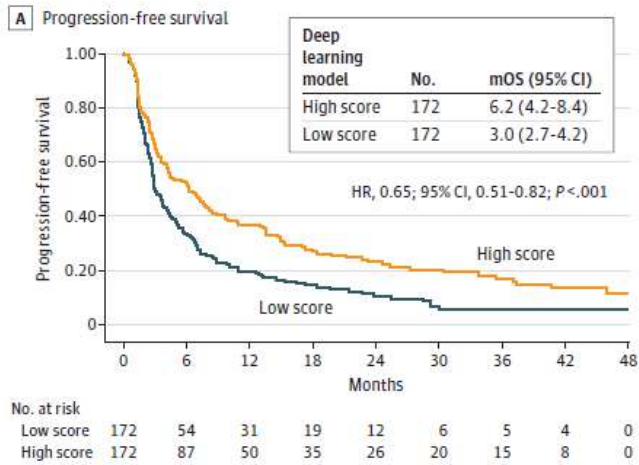
Emergent biomarkers



Application of IA technologies to single-omics analysis



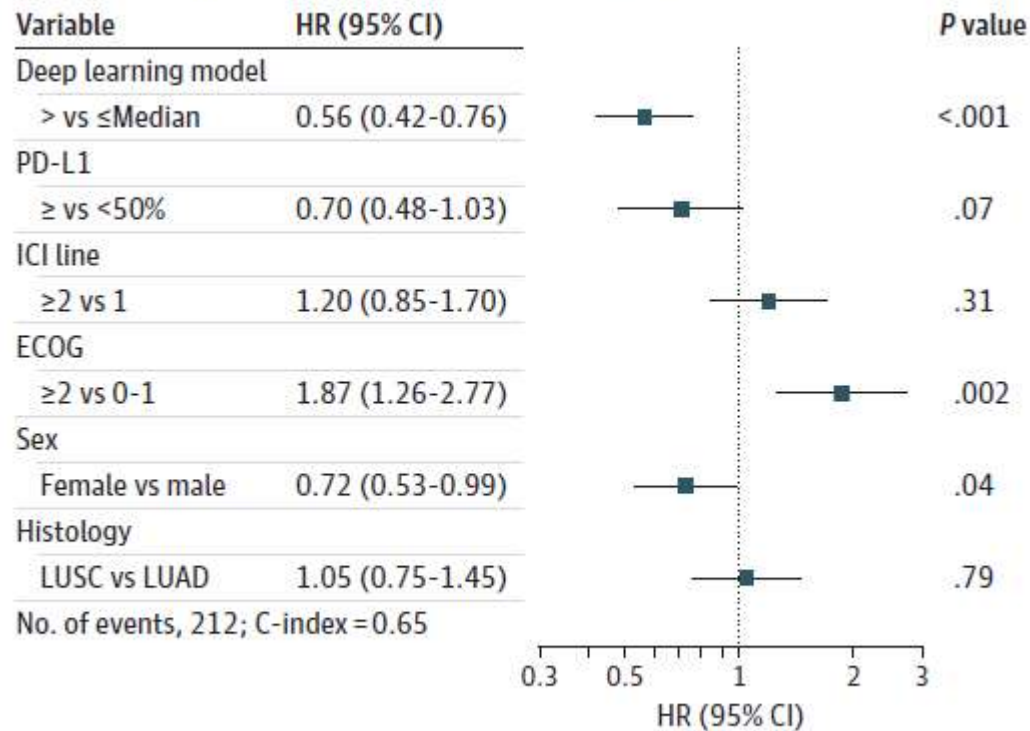
IA Guided precision medicine



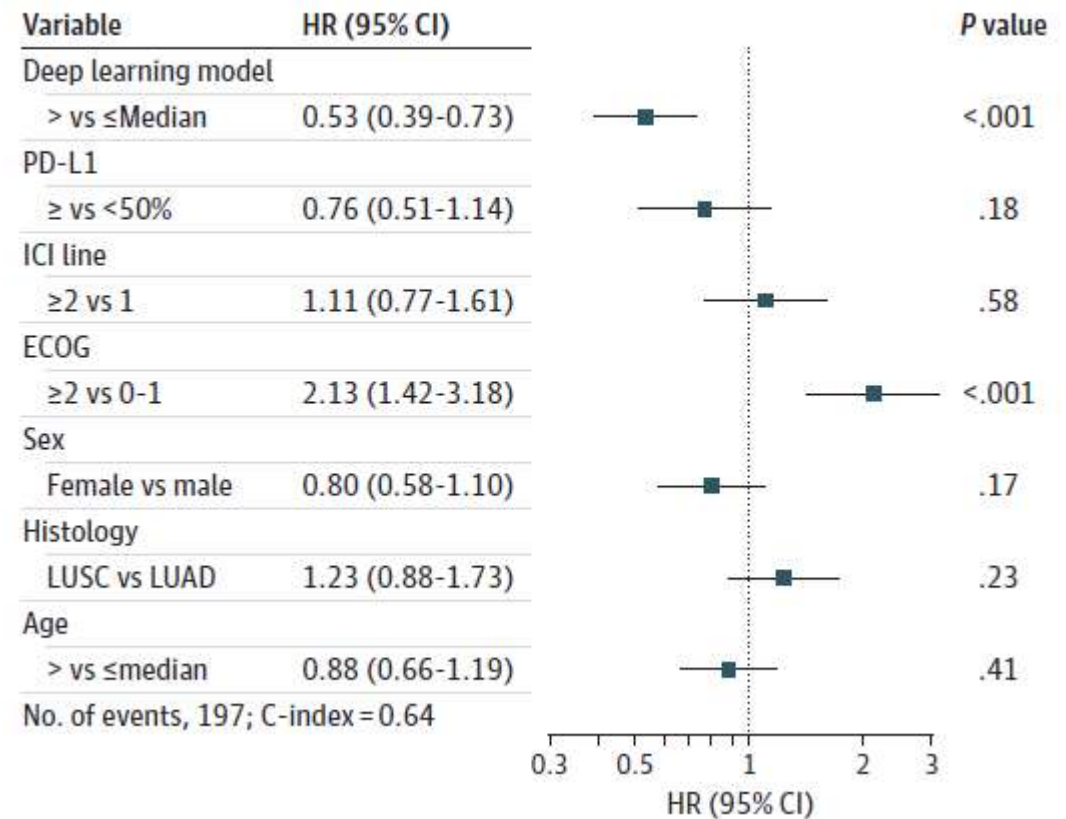
IA Guided precision medicine

Multivariable Analysis in the Validation Cohort

A Cox proportional hazard model of PFS in validation cohort



B Cox proportional hazard model of OS in validation cohort



Multi-omics



Single-cell omics

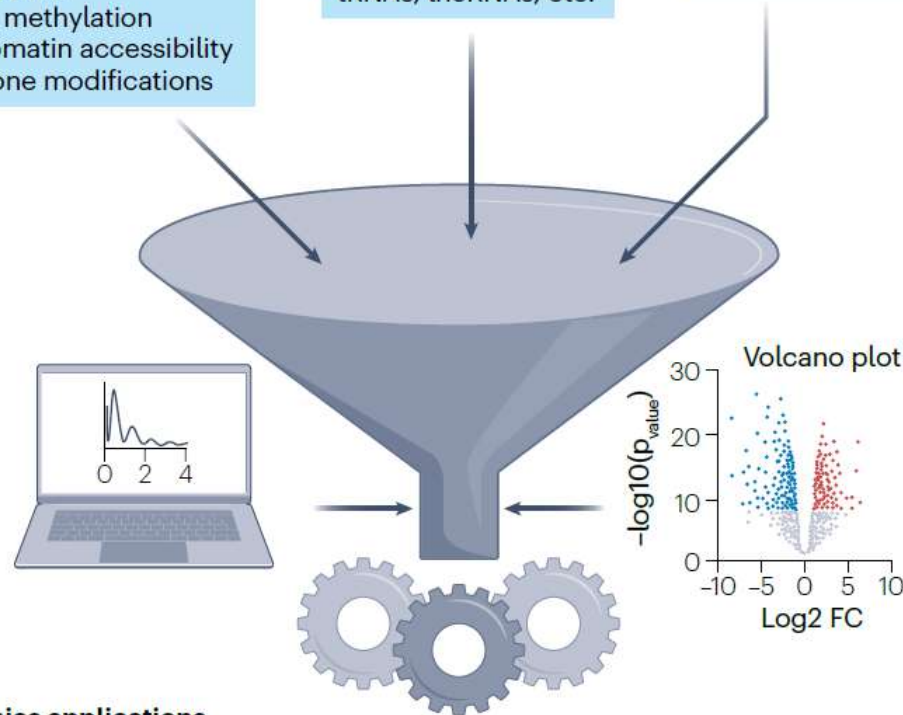
Genome, exome
SNP, CNV

Epigenome

- DNA methylation
- Chromatin accessibility
- Histone modifications

Transcriptome, epitranscriptome
mRNAs, microRNAs, tRNAs, lncRNAs, etc.

Proteome, phosphoproteome, metabolome



Single-cell multi-omics applications

Discovery of novel cell types

Tissue and tumour heterogeneity

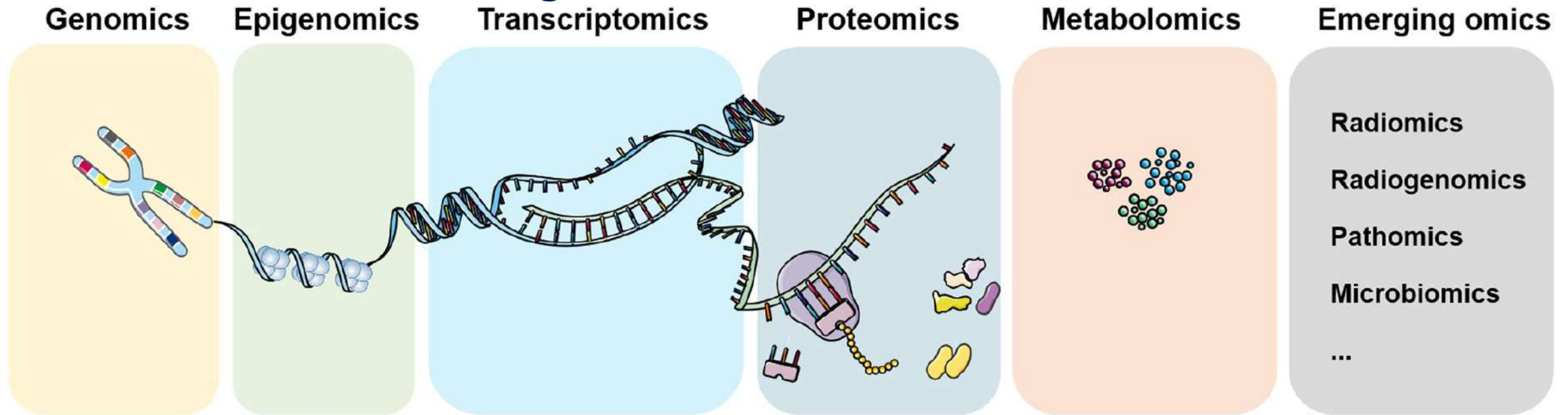
Atlas generation

Biomarker discovery

Insights into complex diseases

Novel pathways and networks

Multi-omics technologies



The information provided by omics technologies

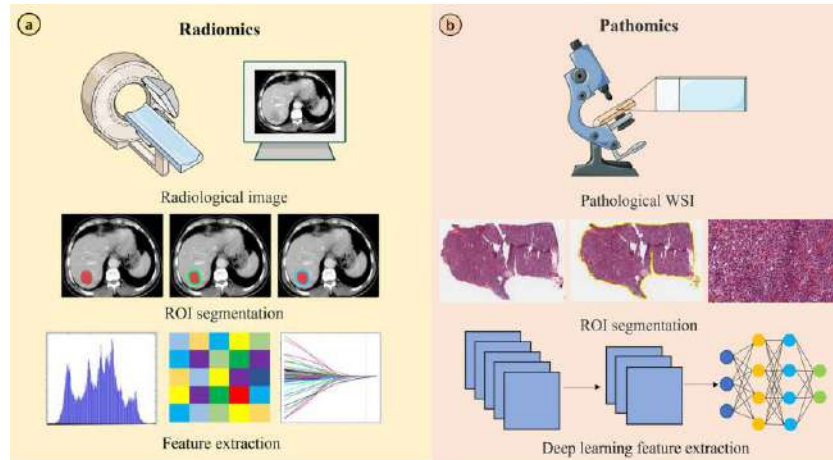
- | | | | | | |
|---|--|---|--|--|---|
| <ul style="list-style-type: none"> • point mutations • small insertions/deletions • genomic rearrangements • viral-genome insertions • structural variants • copy-number variants | <ul style="list-style-type: none"> • DNA modifications • histone modifications and variants • nucleosome occupancy • chromatin interactions • chromatin domains | <ul style="list-style-type: none"> • gene expression • noncoding RNAs • alternative splicing • alternative polyadenylation • gene fusions • allele-specific expression • RNA editing • endogenous retrotransposon transcription | <ul style="list-style-type: none"> • identification and quantitation of proteins • protein modifications | <ul style="list-style-type: none"> • identification and quantitation of metabolites • drug metabolism and toxicity • cancer metabolic reprogramming • immunometabolism | <ul style="list-style-type: none"> • cell composition, cell morphology, and spatial context • quantitative features from digital images • microenvironment information |
|---|--|---|--|--|---|

Representative techniques

- | | | | | | |
|--|---|---|--|--|---|
| <ul style="list-style-type: none"> • WGS • WES | <ul style="list-style-type: none"> • WGBS • ChIP-seq • MeRIP-Seq • ATAC-seq • 3C and derivatives | <ul style="list-style-type: none"> • Microarray • RNA-seq | <ul style="list-style-type: none"> • MS-based proteomics technology • SMPS | <ul style="list-style-type: none"> • NMR spectroscopy • MS-based metabolomics technology | <ul style="list-style-type: none"> • PET/CT, MRI, Dermoscopic images, Mammograms, H&E • WMS, 16S rRNA gene sequencing |
|--|---|---|--|--|---|

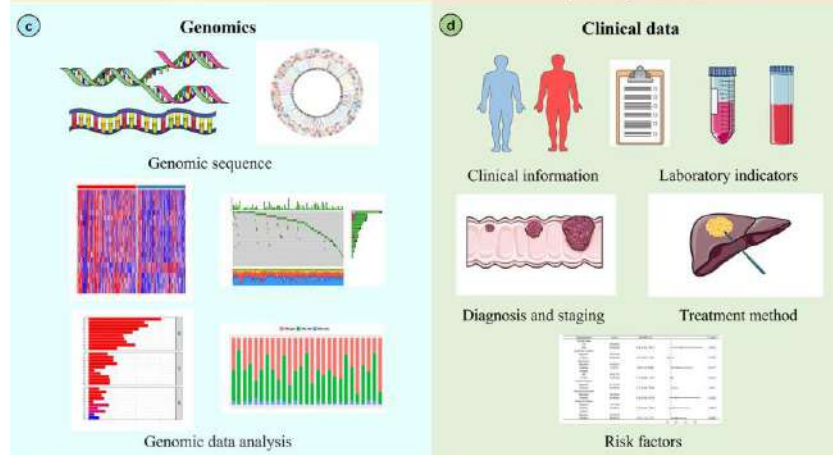
Multi-omics technologies have been developed to profile genome sequences, epigenetic features, transcription expression, protein and metabolite abundance, and more. Additional information from an individual can be captured by emerging omics technologies, such as radiomics, pathomics, and microbiomics. The corresponding representative technologies are enumerated. Whole-genome sequencing (WGS), whole-exome sequencing (WES), whole-genome bisulfite sequencing data (WGBS), chromatin immunoprecipitation sequencing (ChIP-seq), methylated RNA immunoprecipitation sequencing (MeRIP-seq), assay for transposase-accessible chromatin using sequencing (ATAC-seq), chromosome conformation capture (3C) technology, mass spectrometry (MS)-based proteomics technology, single-molecule protein sequencing (SMPS) technologies, Nuclear magnetic resonance (NMR) spectroscopy, mass spectrometry (MS)-based metabolomics technology, positron emission tomography(PET)/computed tomography (CT) scans, magnetic resonance imaging (MRI), Hematoxylin and eosin staining (H&E), and whole metagenome sequencing (WMS).

A Radiomics process mainly includes image acquisition, region of interest (ROI) segmentation, radiomics feature extraction



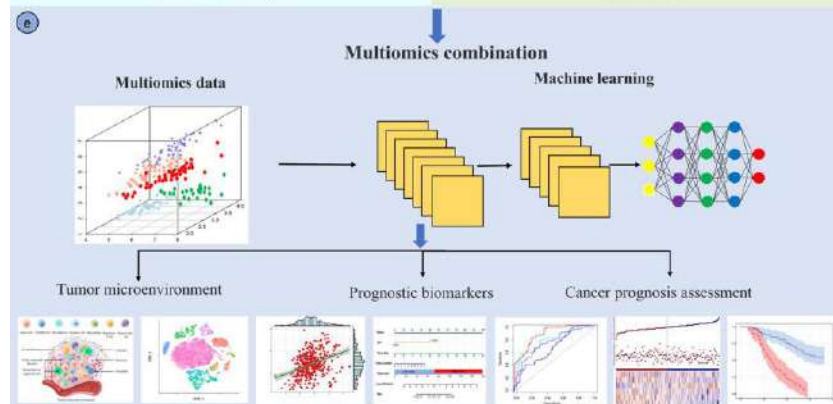
B Pathomics process includes acquisition of pathological images, whole slide images (WSI) ROI segmentation, extraction features of deep learning

C Genomics process mainly includes acquisition of genomics data, analysis of genomics data, etc.



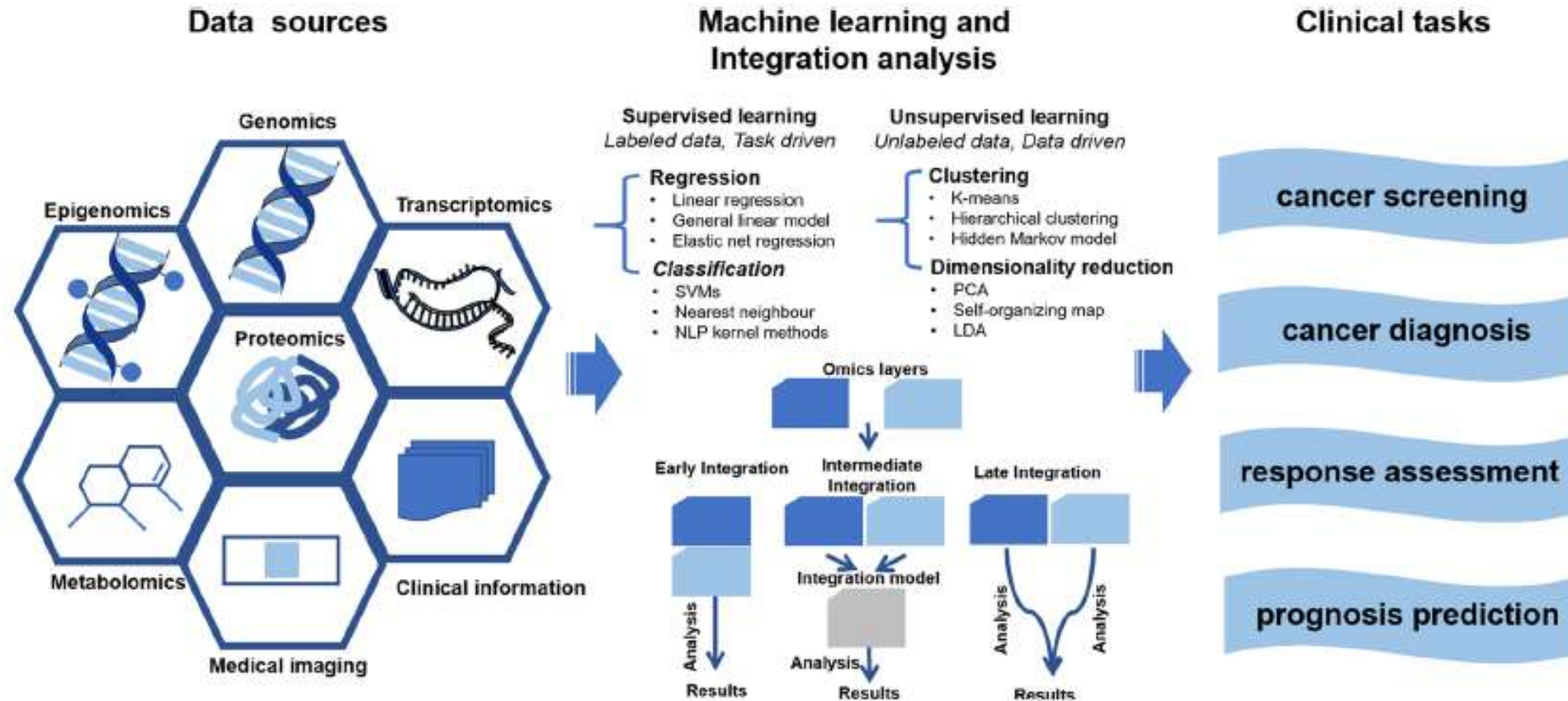
D Clinical data include demographic information, laboratory examination, diagnosis and staging, treatment method, etc.

E Multiomics combinations. Multiomics data are fused by machine learning methods to construct combinatorial models for tumor microenvironment exploration and cancer prognosis evaluation, and to develop comprehensive prognosis biomarkers



Multi-omic combinations

Artificial intelligence support of multi-level biomedical data integration workflow



The data acquired in multi-omics studies can be integrated with other complementary modality data, including medicinal imaging, and clinical information (left panel). ML methods can be divided broadly into supervised learning and unsupervised learning. Representative ML methods are listed (middle panel). Data integration strategies for multi-omics include early, intermediate, and late integration (middle panel). AI-based multi-omics analysis has the potential to aid precision medicine. Key applications of AI-based multi-omics analysis in cancer precision medicine are shown (right panel). Support vector machines (SVMs), natural language processing (NLP) kernel methods, principal component analysis (PCA), and latent dirichlet allocation (LDA).

Conclusions

- Biomarkers testing according to level of evidence (ESCAT)
- Testing in non squamous and light or non smokers squamous lung carcinomas
- Advanced stage and nowadays for early stages
- NGS approaches with panel of genes
- RNA seq preferred as testing for gene fusions
- Perspective of integration of multi-omics and IA

ESMO DEEP DIVE: LUNG CANCER

PRECISION ONCOLOGY IN LUNG CANCER - DIAGNOSTICS

Harnessing liquid biopsy for diagnosis, monitoring

Christian Rolfo, MD, PhD, MBA, Dr.hc.

Professor of Medicine

Director Division Medical Oncology

Associate Director for Early Phase Clinical Trials

The James Comprehensive Cancer Center, The Ohio State University

ESMO WEBINAR SERIES

ESMO GOOD SCIENCE
BETTER MEDICINE
BEST PRACTICE



Disclosures



- ◆ Advisory Board: AstraZeneca, Daiichi Sankyo, Regeneron and Novocure; Bristol-Myers Squibb (BMS), Novartis, Invitae, Guardant Health, COR2ED, Bayer, Boehringer Ingelheim, Abbvie, Invitae, Janssen, EMD Serono
- ◆ Research Grant: Astra Zeneca, Thermo Fisher, Oncohost, Lung Cancer Research Foundation, National Foundation for Cancer Research, and U54 (National Institute of Health)
- ◆ Research collaboration: GuardantHealth, Foundation Medicine, Roche Diagnostics, EMD Serono
- ◆ Scientific advisory board member of Imagene
- ◆ Leadership roles: in International Society of Liquid Biopsy, The European School of Oncology, International Association for Study of Lung Cancer, and Oncology Latin American Association.
- ◆ Editor role: Editor in chief of CROH and Honorary editor at Journal of Liquid Biopsy Elsevier. Editorial board Lung Cancer and ILCN (IASLC).

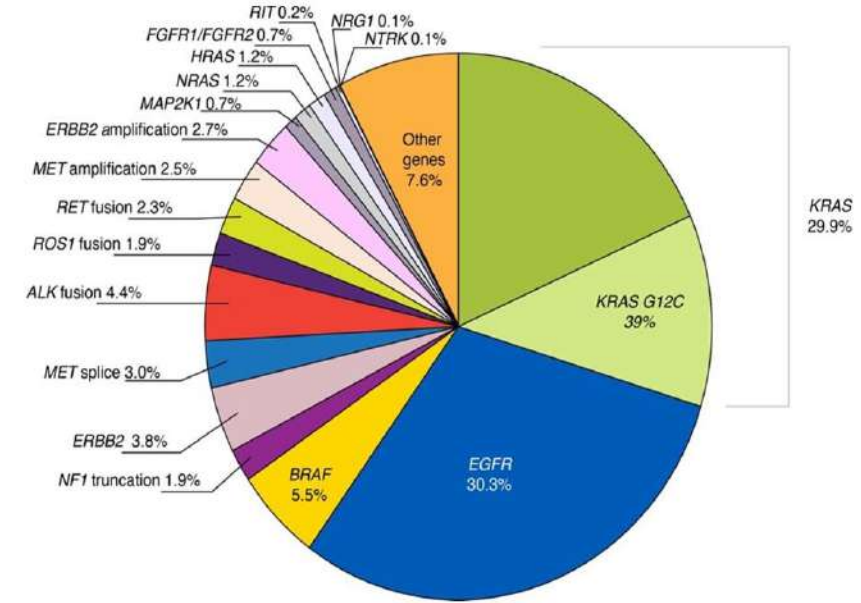
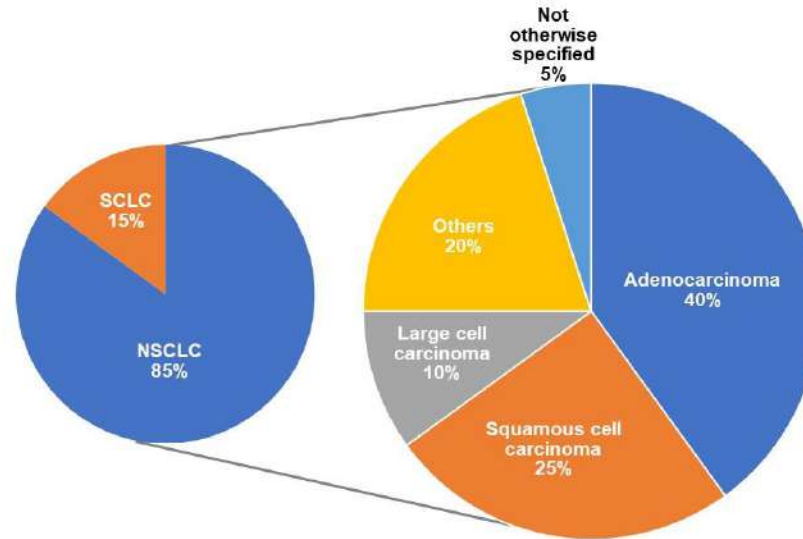
WHY TESTING?

SMALL MOLECULES HAVE TRANSFORMED
HOW WE TREAT HUMAN CANCER



- 20,000 genes
- 3,000 estimated as druggable
- <700 targeted by approved drugs

Landscape of Lung Cancer



PD-L1 expression
(Tumor mutational burden)


Galbraith S. ASCO 2024

Molecular profiling across histologies in lung cancer across 80.000 pts with lung cancer



Study design and patients selection

This was an observational retrospective analysis of:

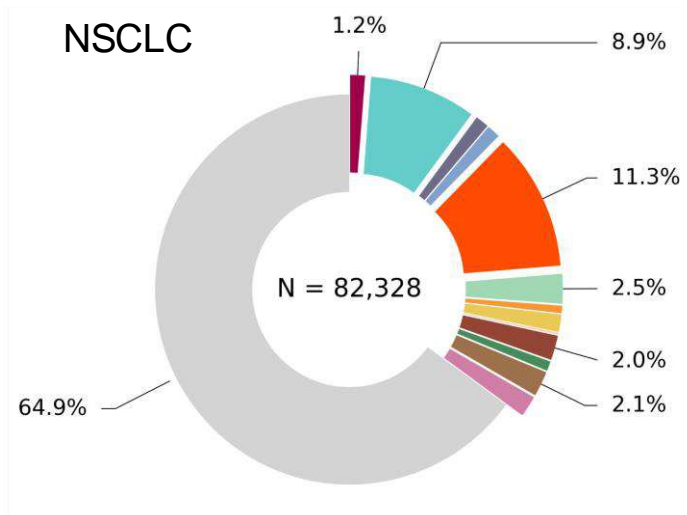


Patients tested at Foundation Medicine Inc laboratory (Cambridge, MA) between 2014 and 2022 (n = 82,328)

- Histologically confirmed non-small cell lung cancer of any age, sex and race.
- Tissue tested using FoundationOne/Foundation CDx (F1/F1CDx).
- +/- Clinical and demographic data Western Institutional Review Board Protocol No. 20152817.

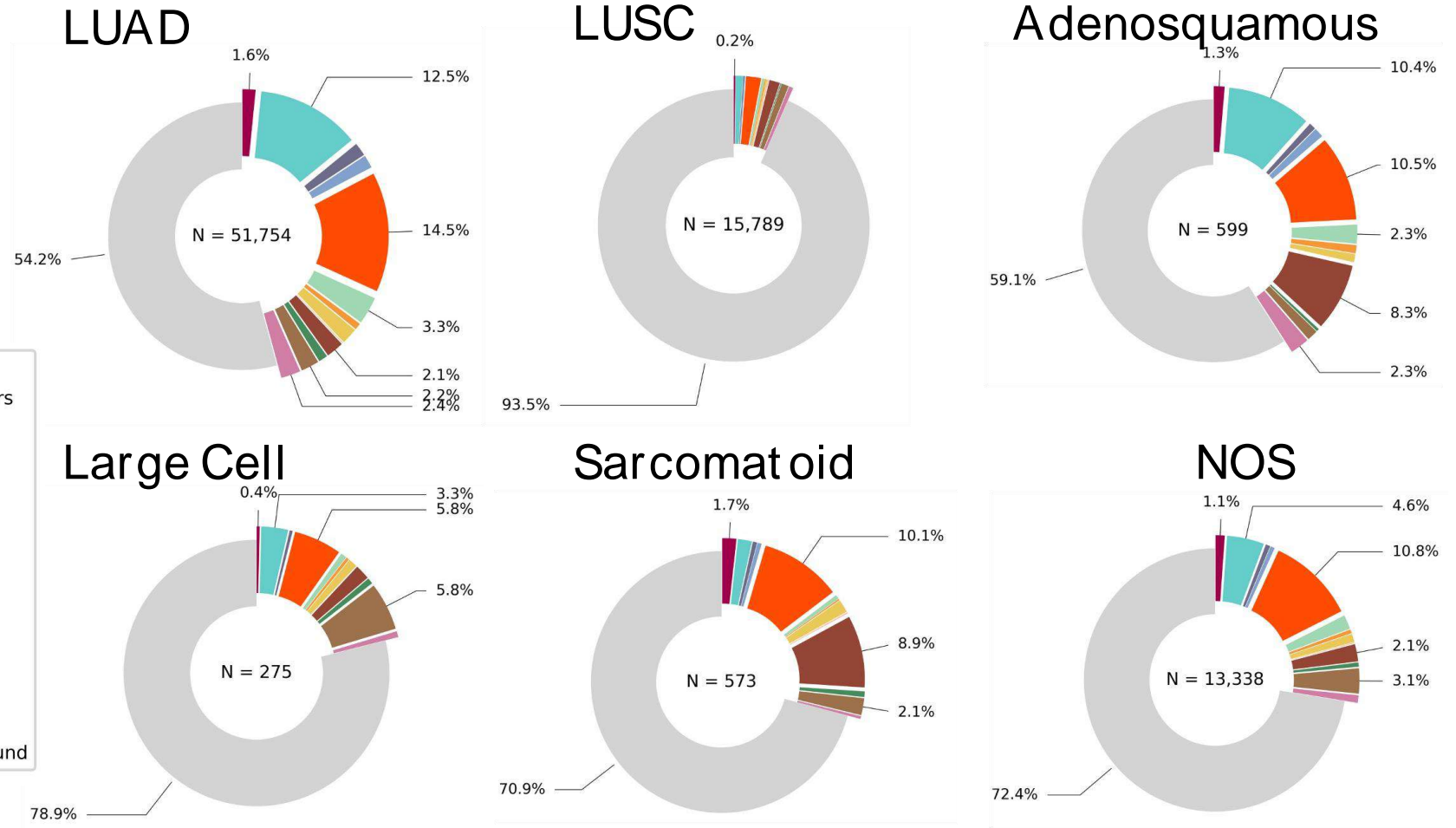
Biomarkers

| | |
|---|--|
| ■ | Multiple Actionable Genomic Biomarkers |
| ■ | EGFR ex19 deletion and L858R |
| ■ | EGFR S7681, L861Q, or G719X |
| ■ | EGFR ex20 insertion |
| ■ | KRAS G12C |
| ■ | ALK RE |
| ■ | ROS1 RE |
| ■ | BRAF V600E |
| ■ | NTRK1/2/3 RE |
| ■ | MET ex14 skipping |
| ■ | RET RE |
| ■ | MET amplification |
| ■ | ERBB2 mutation |
| ■ | No Actionable Genomic Biomarkers Found |



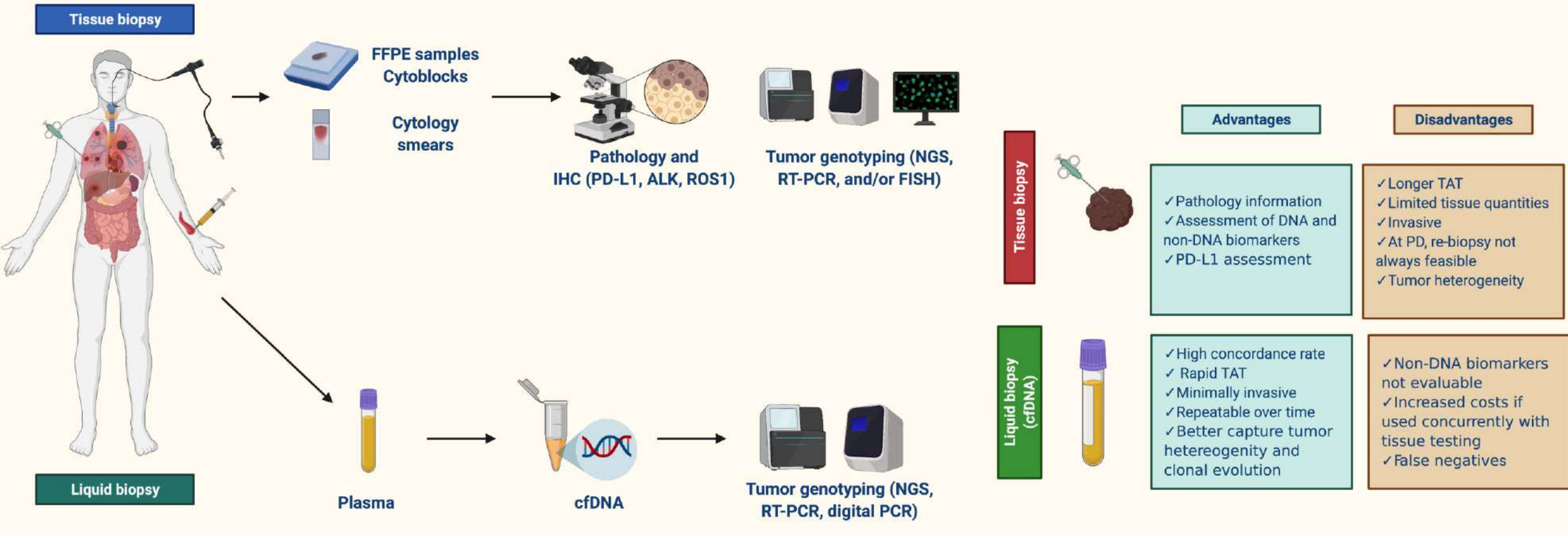
Russo A (Rolfo C), et al. ASCO 2024

Distribution of genomic alterations across histologies



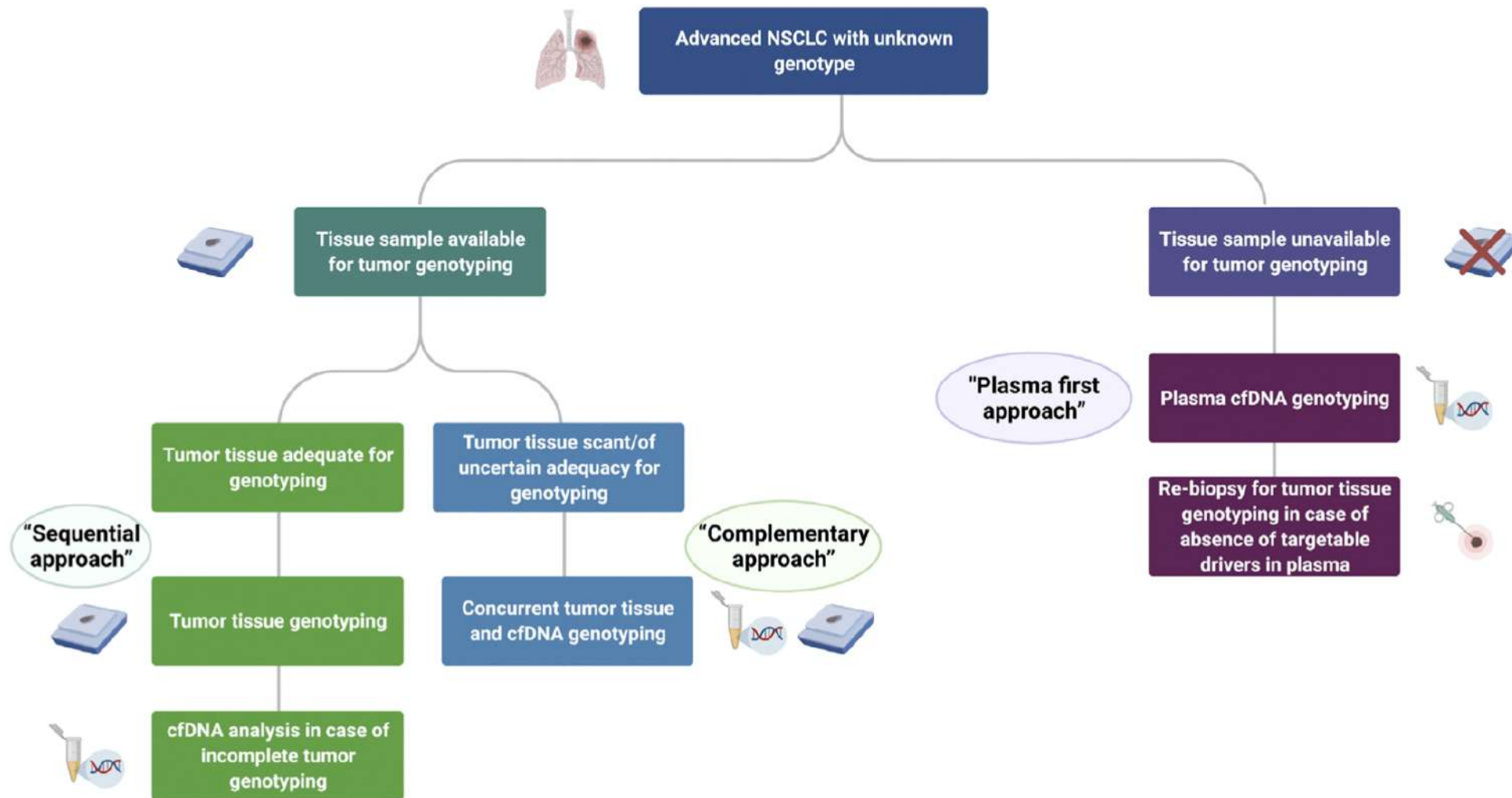
Russo A (Rolfo C), et al. ASCO 2024

Tissue vs. Liquid biopsy



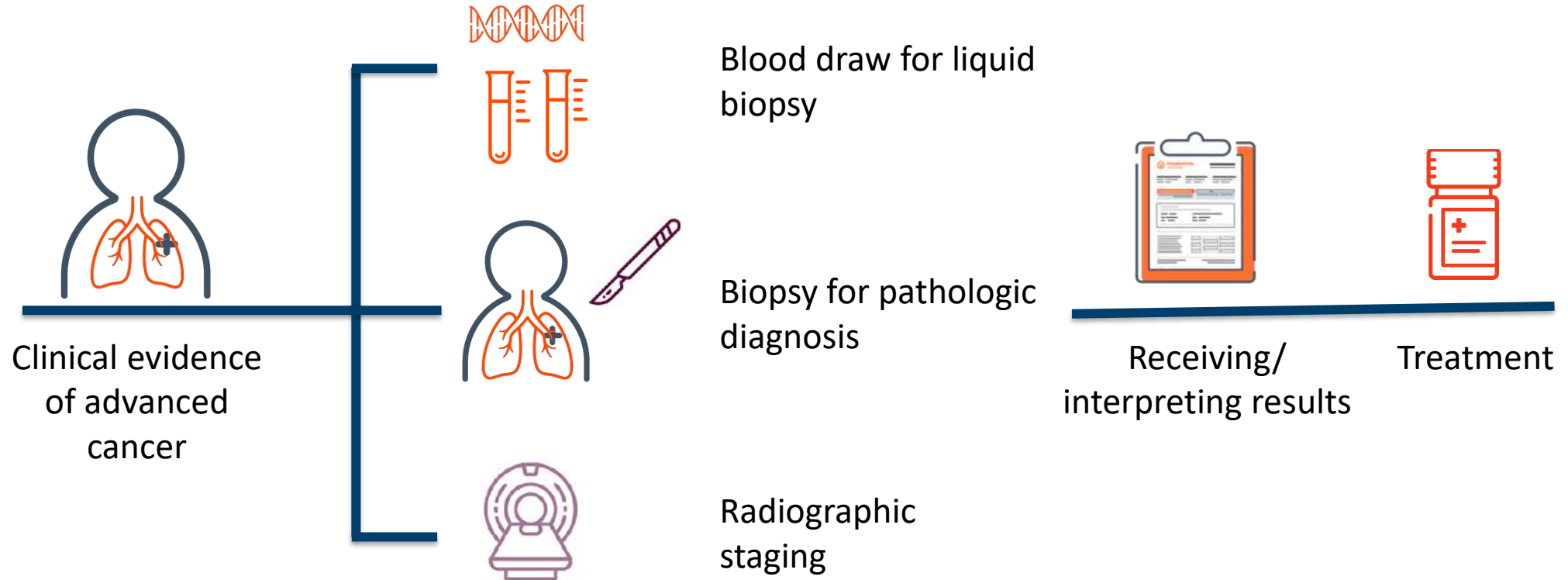
Rolfo C, et al. *J Thorac Oncol* 2021;16(10):1647-1662.

Diagnostic algorithm for liquid biopsy use in treatment-naive advanced/metastatic NSCLC



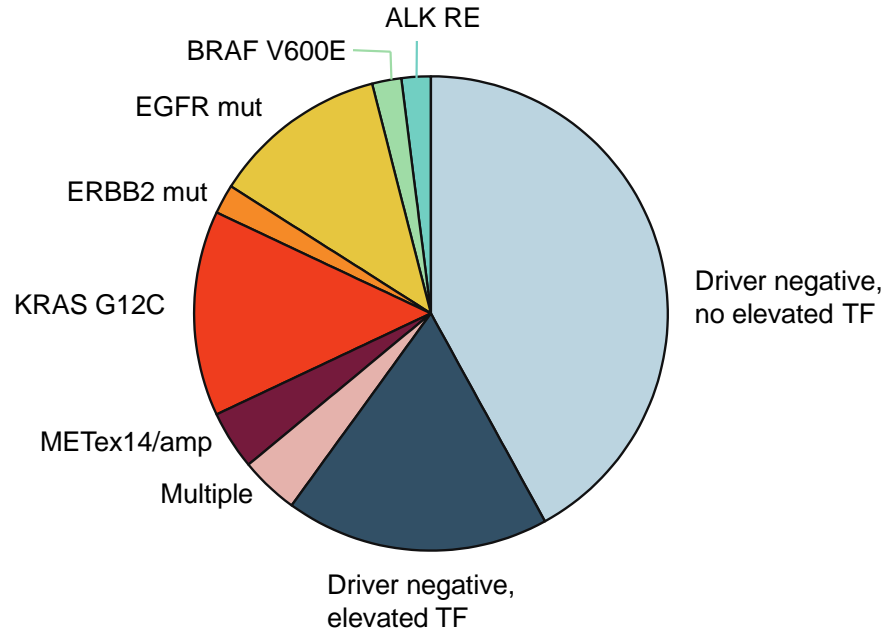
Expedited diagnostic odyssey

stacking diagnostic steps may be able to shorten the diagnostic odyssey

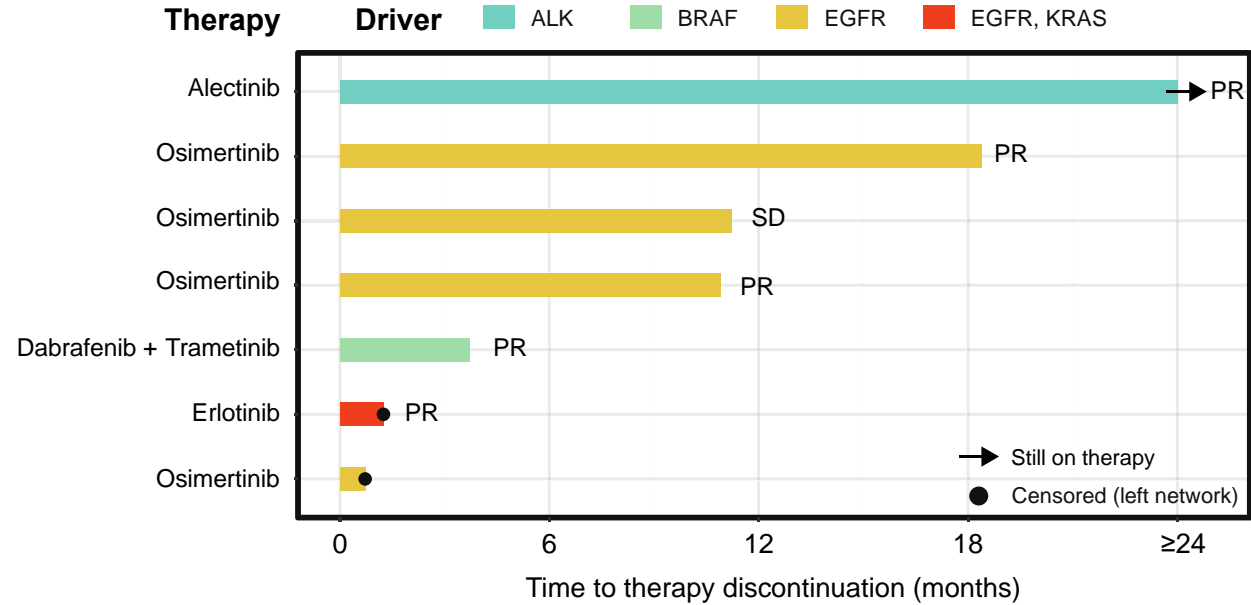


Russo A. et al (Rolfo C.) JCO PO, Feb 2024

36% of early LBX samples were positive for an actionable NCCN driver



9 (18%) of patients were NCCN driver negative with estimated tumor fraction (TF) $\geq 10\%$ (presumed true negatives)



7 driver+ patients received a 1L matched targeted therapy with a median TTD of 11 months and real-world response (PR) in majority of patients

Russo A. et al (Rolfo C.) JCO PO, Feb 2024

ctDNA tumor fraction informs the relative benefit from reflex to TBx CGP

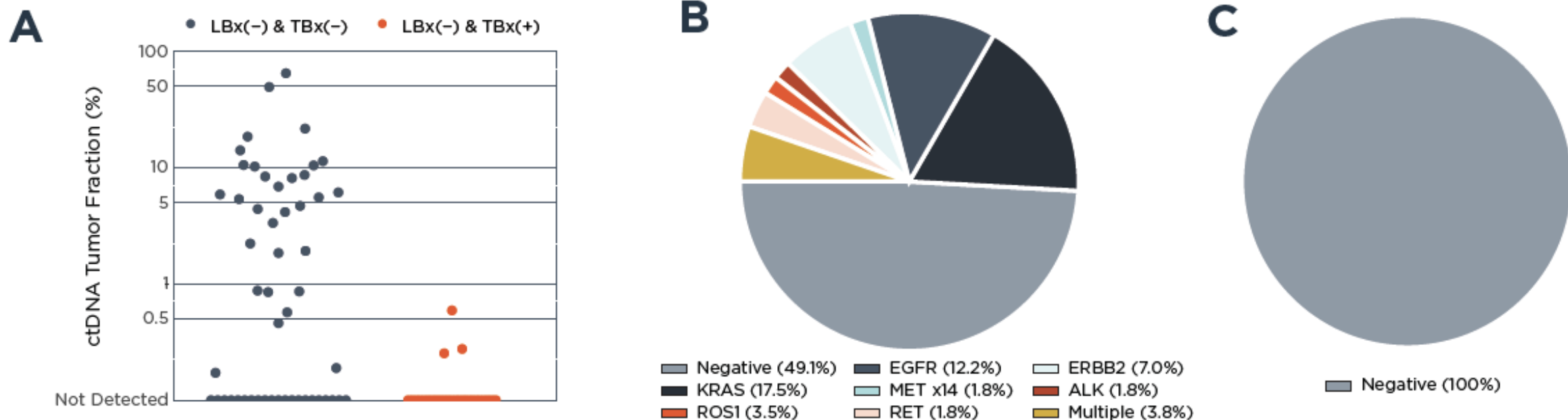
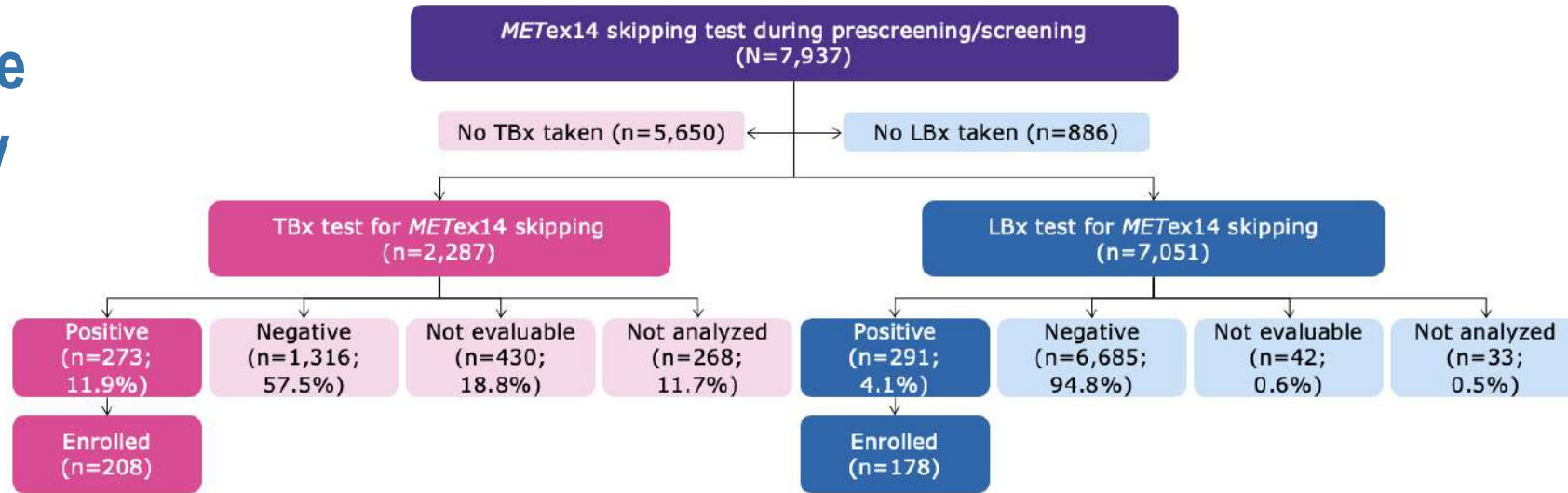


Figure 7: A) 24/81 (30%) patients with reflex TBx after negative LBx had a LBx TF $\geq 1\%$ and, given high NPV for driver alterations, might have avoided reflex to confirmatory TBx. **B)** Amongst patients with TF $< 1\%$, 51% (29/57) of patients had a driver mutation detected on TBx reflex while **C)** no driver mutations (0%) were seen for patients with TF $\geq 1\%$

Liquid biopsies (lhx) and tissue biopsies (tbx) for identifying *MET* exon 14 skipping (*metex14*) in advanced NSCLC: analyses from the phase II VISION study of tepotinib



Prognostic Value of Liquid Biopsy

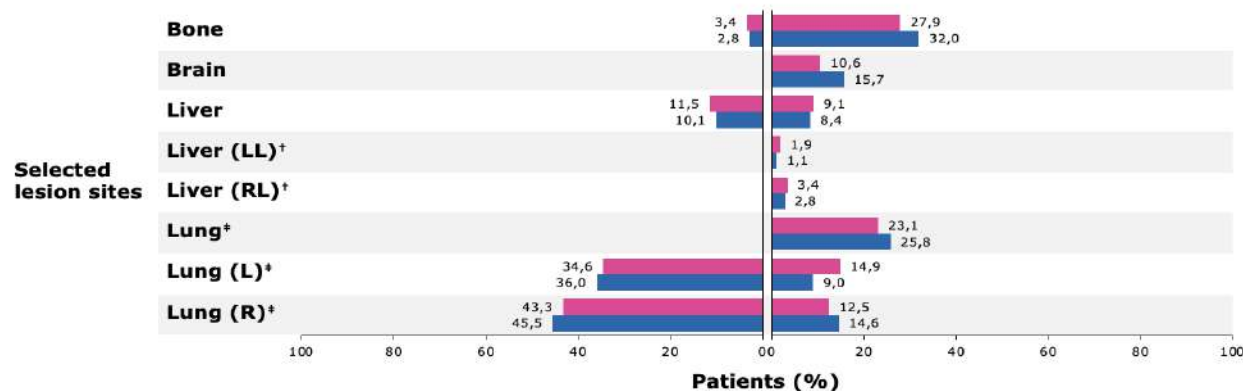


| | T+/L _{N/A} | T+/L- | T+/L+ | T-/L+ | T _{N/A} /L+ |
|------------------------|---------------------|---------------|--------------|------------|----------------------|
| Overall (n=313) | n=28 (8.9%) | n=106 (33.9%) | n=74 (23.6%) | n=6 (1.9%) | n=98 (31.3%) |
| 1L (n=164) | n=17 (10.4%) | n=52 (31.7%) | n=42 (25.6%) | n=3 (1.8%) | n=50 (30.5%) |
| +2L (n=149) | n=11 (7.4%) | n=54 (36.2%) | n=32 (21.5%) | n=3 (2.0%) | n=48 (32.2%) |

Rolfo C. ASCO 2023

| Median SOLD per RECIST v1.1, mm (range) | |
|---|-------------------|
| T+ (n=208) | 55.2 (10.2–267.5) |
| L+ (n=178) | 67.1 (11.6–227.8) |

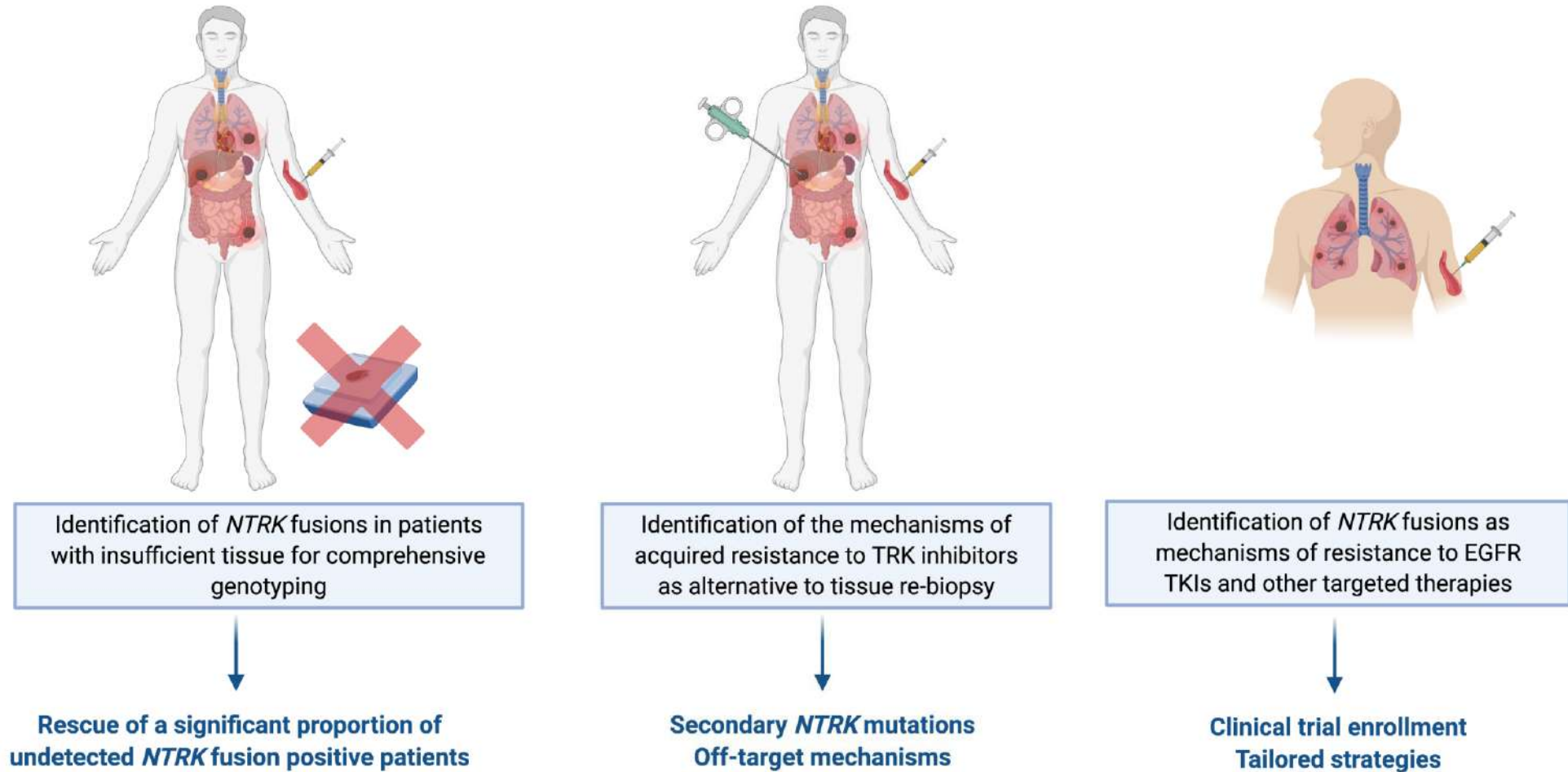
| Number of lesions* | Target lesions | | | Non-target lesions | | |
|--------------------|----------------|-------|-------|--------------------|-------|-------|
| | ≥3 | 2 | 1 | ≥3 | 2 | 1 |
| T+ | 18,8% | 30,3% | 51,0% | 21,6% | 32,2% | 36,1% |
| L+ | 27,5% | 28,7% | 43,8% | 15,7% | 34,3% | 42,7% |



| IRC | 1L | | +2L | |
|--------------------------|----------------------|----------------------|----------------------|----------------------|
| | T+/L- (n=52) | T+/L+ (n=42) | T+/L- (n=54) | T+/L+ (n=32) |
| ORR, % (95% CI) | 57.7 (43.2, 71.3) | 64.3 (48.0, 78.4) | 44.4 (30.9, 58.6) | 53.1 (34.7, 70.9) |
| mDOR, months (95% CI) | ne (10.4, ne) | 19.4 (7.6, ne) | 12.6 (5.1, 20.8) | 9.9 (4.4, 15.4) |
| mPFS, months (95% CI) | 22.1 (14.8, ne) | 12.1 (7.8, 49.7) | 13.8 (8.2, 24.9) | 8.2 (5.5, 13.7) |
| mOS, months (95% CI) | 32.7 (15.3, ne) | 28.5 (14.2, ne) | 20.8 (15.6, 32.5) | 19.8 (10.0, 26.5) |

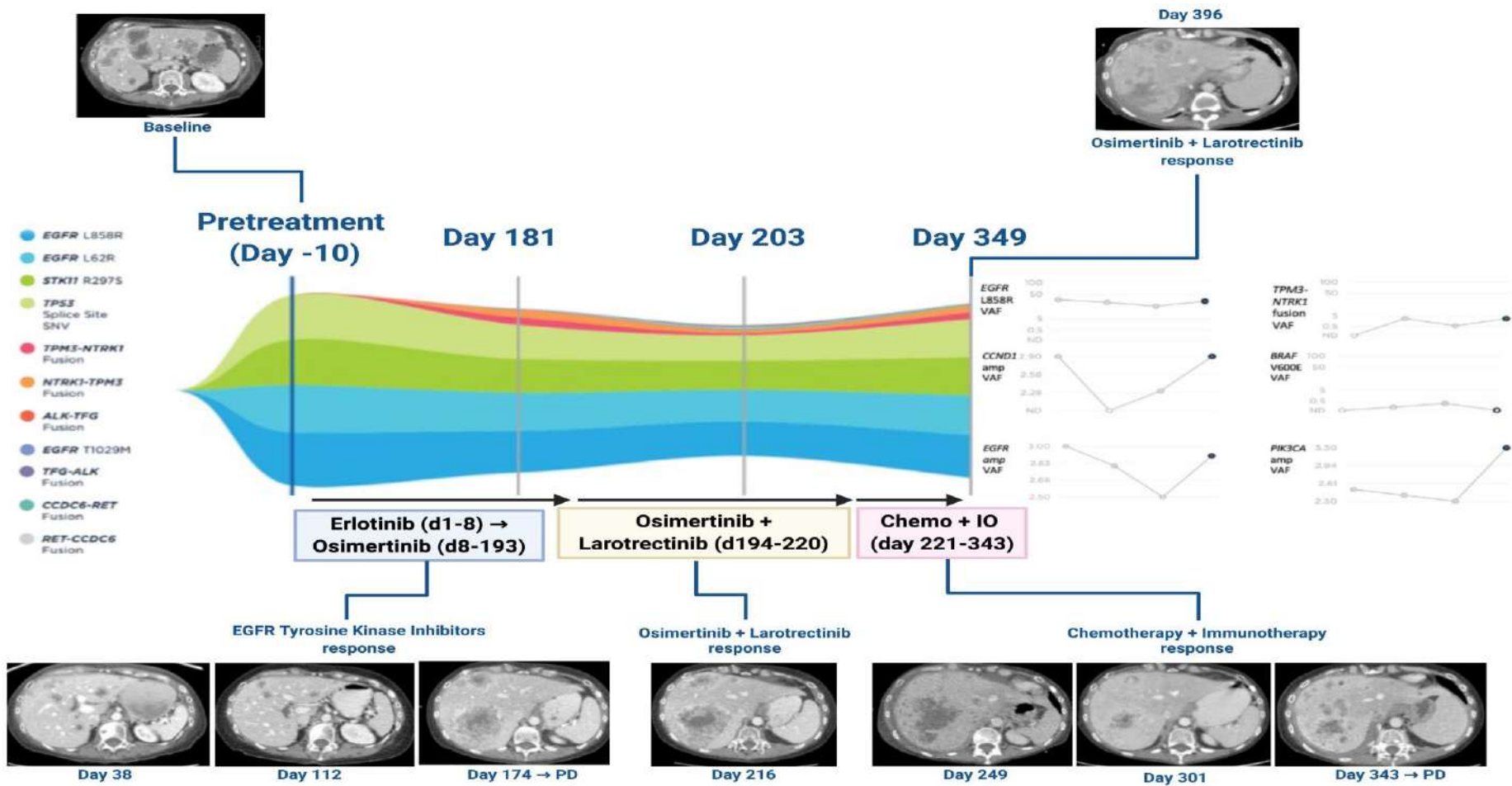
Rolfo C. ASCO 2023

Potential utility of liquid biopsy in *NTRK* fusion-positive NSCLCs



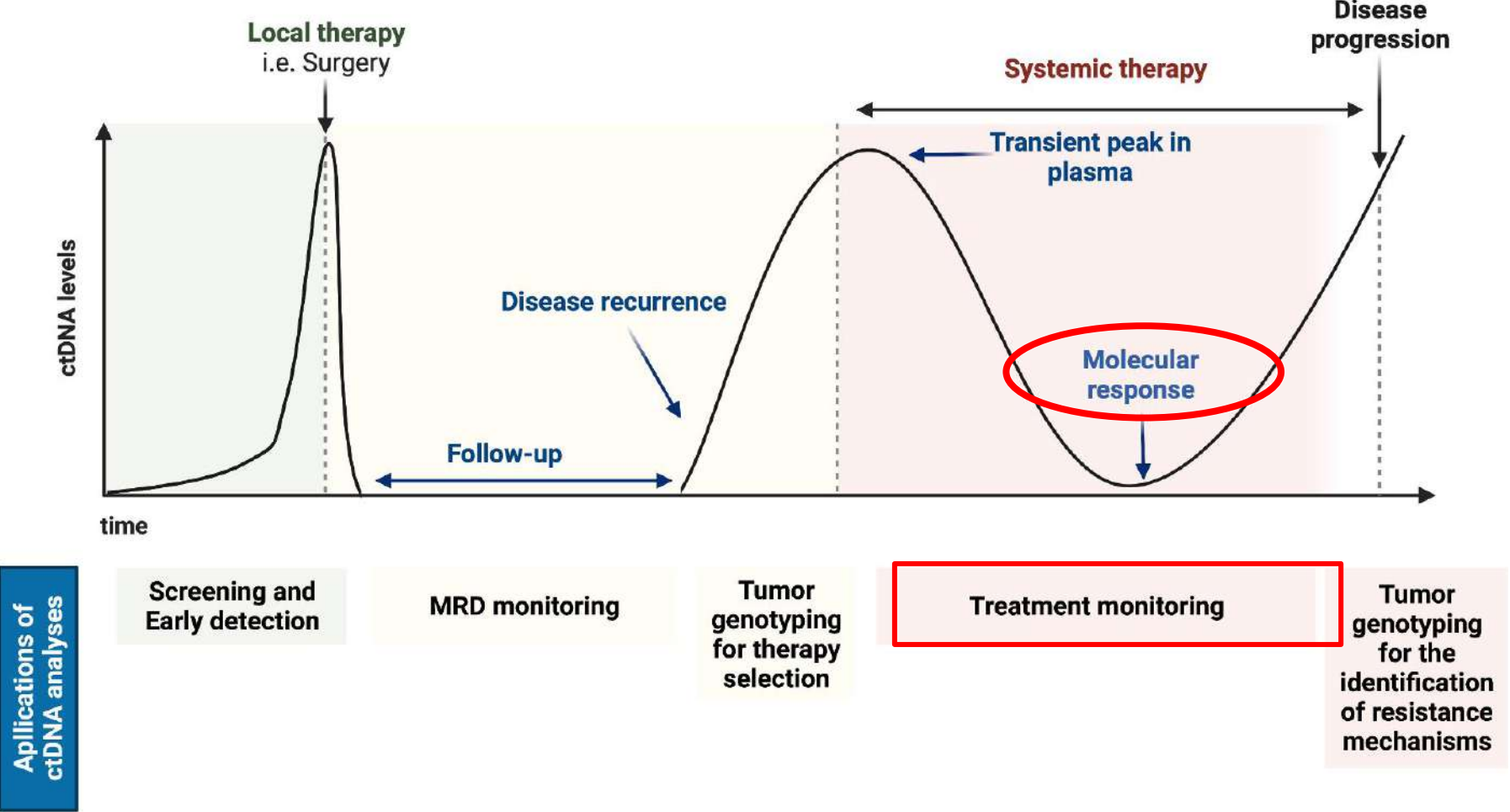
Rolfo C, et al, Br J Cancer 2021

NTRK fusions as mechanism of resistance



Rolfo C, et al, Br J Cancer 2021

Potential clinical applications of LB in lung cancer management



Malapelle U, et al. Lung Cancer. 2022;172:53-64.

Liquid biopsy can capture the dynamic evolution of resistance mechanisms to EGFR TKIs



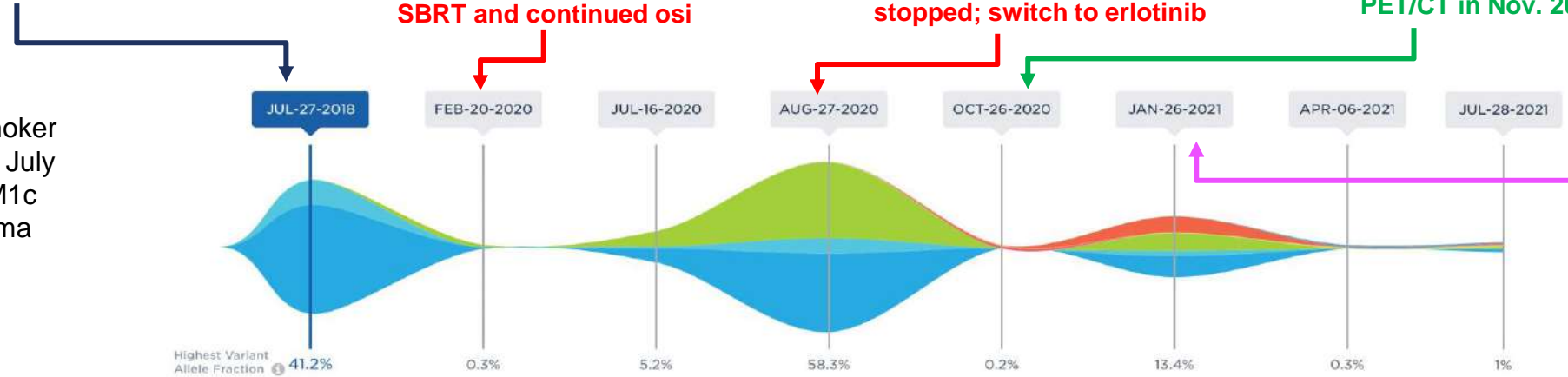
Osimertinib start with an intracranial CR and extracranial PR

52-year-old never smoker female; diagnosed in July 2018 with cT4 N3 M1c lung adenocarcinoma (stage IVB).

After 18 mos oligo-PD (LN mets) → SBRT and continued osi
 Further disease progression → osimertinib stopped; switch to erlotinib

PET/CT in Nov. 2020: CR

Erlotinib discontinuation → platinum-based chemo start



| Genetic Alteration | % cfDNA or amplification | | | | | | | | |
|--------------------|--------------------------|------|------|-------|------|-------|------|------|--|
| EGFR E746_A750del | 41.2% | 0.2% | 4.7% | 58.3% | ND | 13.4% | ND | 1% | |
| EGFR C797S | ND | 0.3% | 5.2% | 55.6% | ND | 10.7% | ND | 0.7% | |
| ARID1A Q456Q | ND | ND | ND | 0.2% | ND | 0.2% | 0.3% | 0.6% | |
| EGFR T790M | ND | ND | ND | ND | ND | 9.6% | ND | 0.4% | |
| TP53 C275Y | ND | ND | ND | ND | ND | ND | 0.1% | 0.2% | |
| ARID1A F1728F | ND | ND | ND | ND | ND | ND | 0.3% | 0.2% | |
| TP53 S127F | 6.5% | ND | 0.4% | 7.6% | ND | 2.6% | ND | 0.2% | |
| BRAF Amplification | 2.2% | ND | ND | ND | ND | ND | ND | ND | |
| CDK6 Amplification | 2.2% | ND | ND | ND | ND | ND | ND | ND | |
| EGFR Amplification | 3.4% | ND | ND | 4.2% | ND | ND | ND | ND | |
| NTRK2 L699L | - | - | - | - | 0.2% | ND | ND | - | |
| EGFR N338N | ND | ND | ND | ND | 0.1% | ND | ND | ND | |
| FGFR1 V795I | ND | ND | ND | ND | ND | ND | 0.1% | ND | |

Russo A (Rolfo C) et al. Clin Lung Cancer 2023

A sample of ctDNA report with key elements

Hypermutable phenotype
A hypermutable phenotype caused by defective DNA mismatch repair

Sequencing Depth
The number of times each DNA fragment is read during sequencing; the smaller the panel, the greater the depth

Germline vs Somatic
Follow-up germline testing may be required to distinguish between germline and somatic findings; considered more likely to be germline if VAF approximately 50% (the low VAF represented here suggests a subclonal somatic mutation)²⁴

Breadth of Coverage
The number of genes sequenced (all reports will usually have a full list of genes sequenced; the more genes covered, the lower the depth)

| | | | |
|-----------------------------------|--------------------|-----------------------------|--------------------------------|
| TUMOR TYPE Lung adenocarcinoma | COUNTRY CODE TW | REPORT DATE 28 July 2021 | ORDER TEST # ORD-1147354-01 |
| PATIENT | | | |
| DISEASE Lung adenocarcinoma | | | |
| NAME | | | |
| DATE OF BIRTH | | | |
| SEX | | | |
| MEDICAL RECORD # | | | |
| PHYSICIAN | | | |
| ORDERING PHYSICIAN Si, Wu-Choi | | | |
| SPECIMEN | | | |
| DATE OF COLLECTION 19 July 2021 | | | |
| SEQUENCING DEPTH (2000X) | | | |

Biomarker Findings

Blood Tumor Mutational Burden - 3 Muts/Mb
 Microsatellite status - MSI-High Not Detected
 Tumor Fraction - 25%

| Genomic Findings | VAF (%) |
|--------------------|---------|
| EGFR L858R | 20 |
| EGFR T790M | 15 |
| EGFR amplification | NA |
| TP53 Q192* | 4 |
| BRCA2 | 1 |
| DNMT3A | 1.5 |

9 Therapies with Clinical Benefit 4 Therapies with Lack of Response 20 Clinical Trials

Tumor Fraction
Tumor fraction can be an indicator of the robustness of the report

VAF
The approximate percentage of ctDNA present in a ctDNA sample; take into consideration when interpreting VAFs

SNVs
A single nucleotide change in DNA

InDels
Insertion and/or deletion of nucleotides into/from DNA

CNAs
Increase or loss in the number of copies of a particular gene

REs
Movement of DNA sequences across the genome that may lead to gene fusions

CHIP
an age-related source of biological noise, due to hematopoietic cell variations that can falsely appear as ctDNA variations

BIOMARKER FINDINGS

Blood Tumor Mutational Burden-3 Muts/Mb

10 Tris

Microsatellite status: MSI-high not detected

Tumor fraction, 25%

MSI-High not detected
No evidence of MSI in this sample

Tumor fraction is an estimate of the percentage of ctDNA present in a ctDNA sample based on observed aneuploid instability.

BREADTH OF COVERAGE

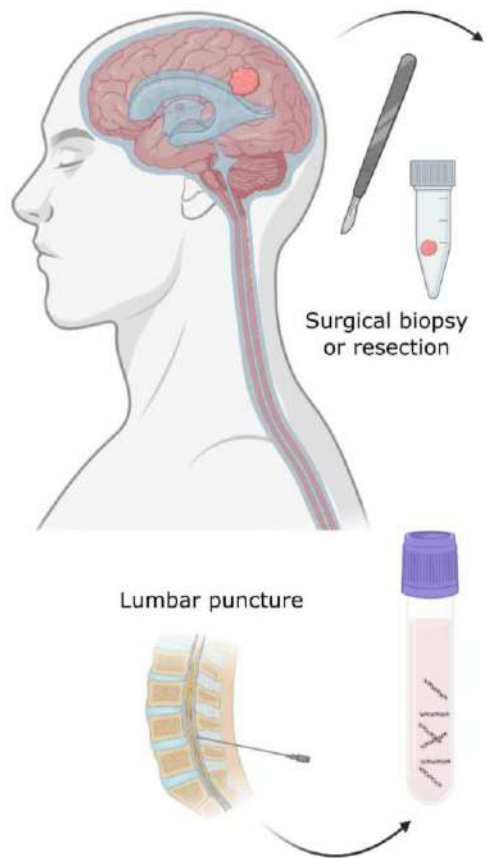
The number of genes sequenced (all reports will usually have a full list of genes sequenced; the more genes covered, the lower the depth)

Longitudinal changes in VAF of genomic alterations over time

Krebs MG (Rolfo C) et al. JAMA oncol 2022

Special situations: brain metastasis in TKI resistance

CSF demonstrates superiority of cell-free DNA over cell pellet genomic DNA for molecular profiling



Solid biopsy (tumour specimen)

Advantages
Allow histological diagnosis

Limitations
Very invasive and risky procedure
Sometimes not feasible due to tumour anatomical location
Not representative of tumour heterogeneity
Static snapshot

Liquid biopsy (CSF ctDNA)

Advantages
Less-invasive and easier to obtain than a tumour biopsy
CSF obtained as SOC for some patients
Concordance with tissue characterisation
Representative of intratumour and interlesion heterogeneity
Longitudinal real-time monitoring

Limitations
No histological characterisation
Lack of standardisation
Contraindications for lumbar puncture
Limited sensitivity

Effective in establishing CNS involvement:

- >97% cases with mutational profile matching primary tumor
- >70% of cases profiled had mutations

New findings compared with primary tumor in >17% demonstrating:

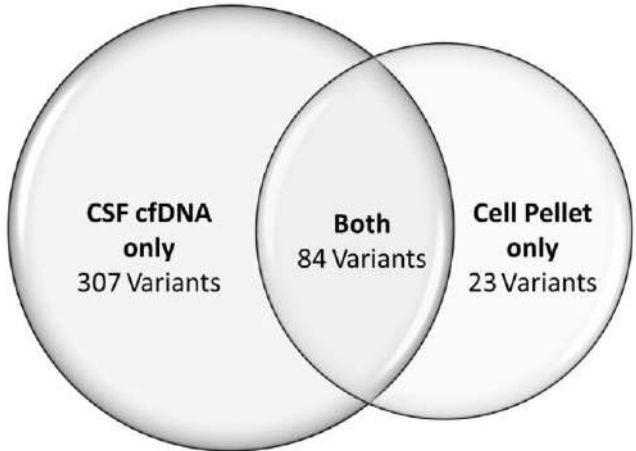
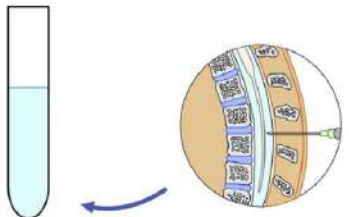
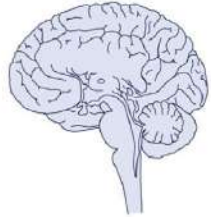
- Resistance mechanisms
- Clonal evolution
- New primary diagnoses

More informative than tumor cell profiling from same CSF sample

- Higher success
- More variants detected
- Higher variant frequency

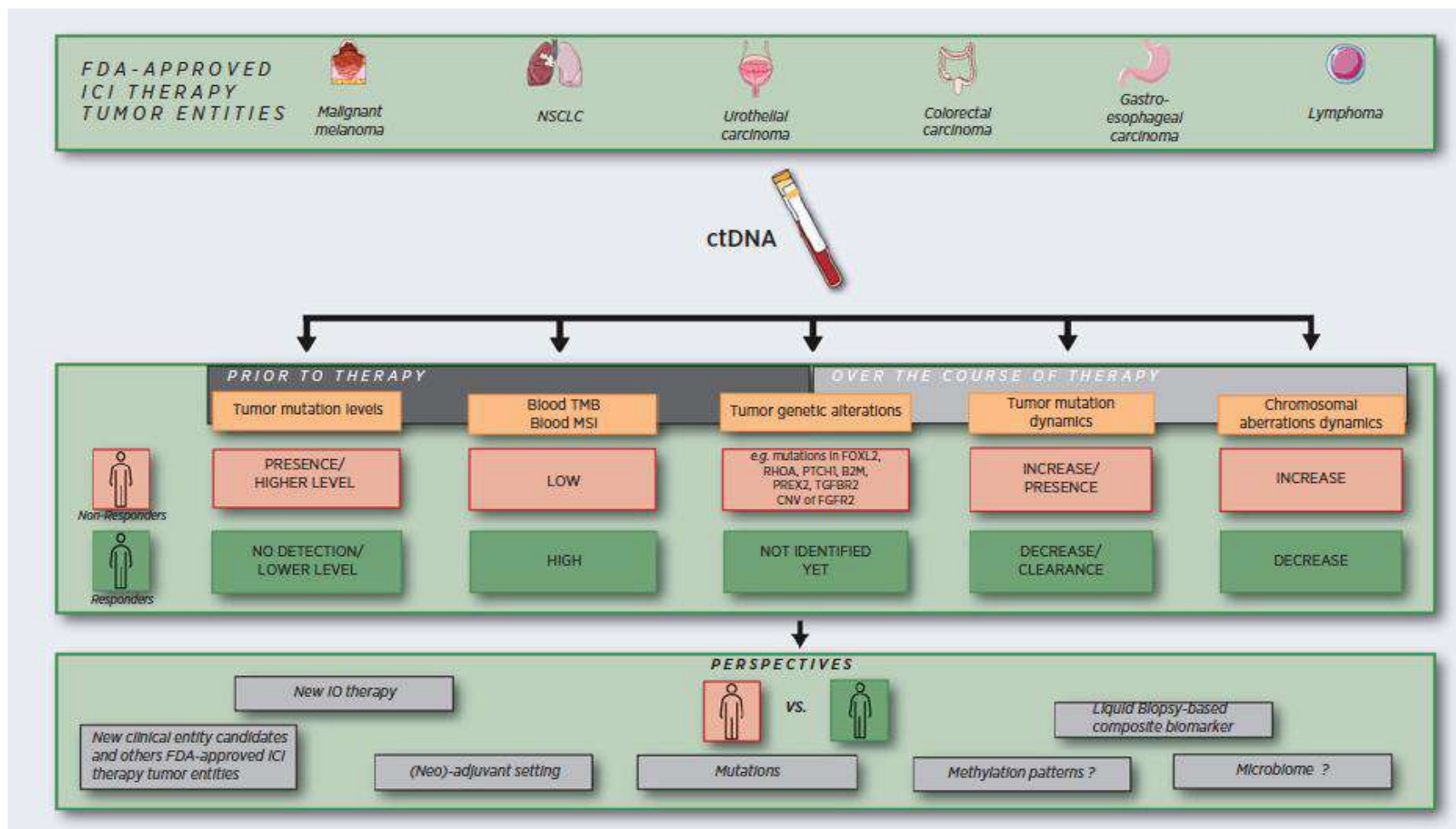
Meaningful results generated even in samples with:

- Very Low input DNA
- Normal CSF cytology



Escudero et al, *Cancers* 2021, 13(9), 1989; Bale et al (Arcila M.) *J Mol Diagn* . 2021 Jun;23(6):742-752

Use of liquid biopsy in immunotherapy

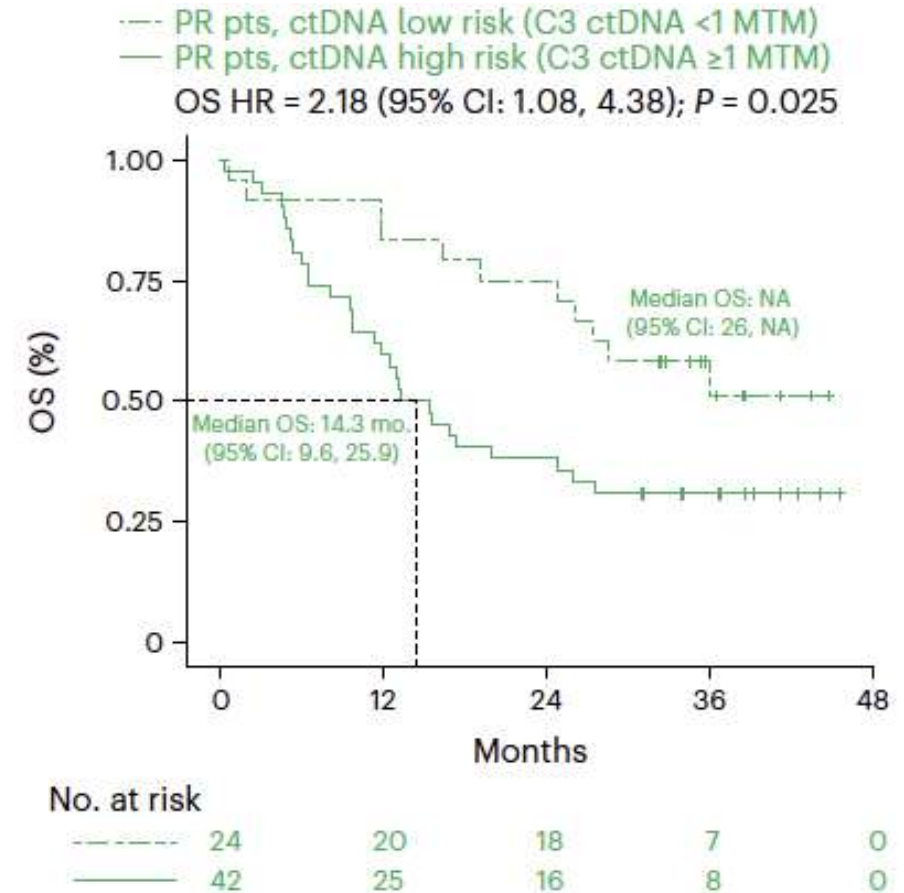
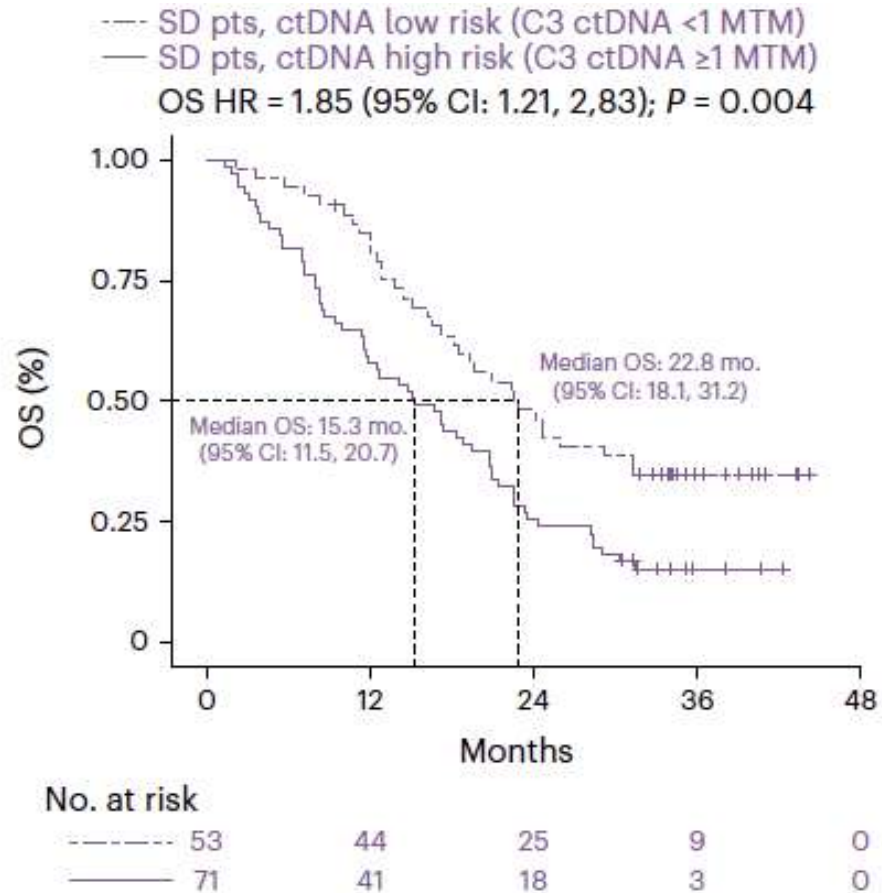


Stadler J, et al. Cancer Res 2022

ESMO DEEP DIVE: LUNG CANCER

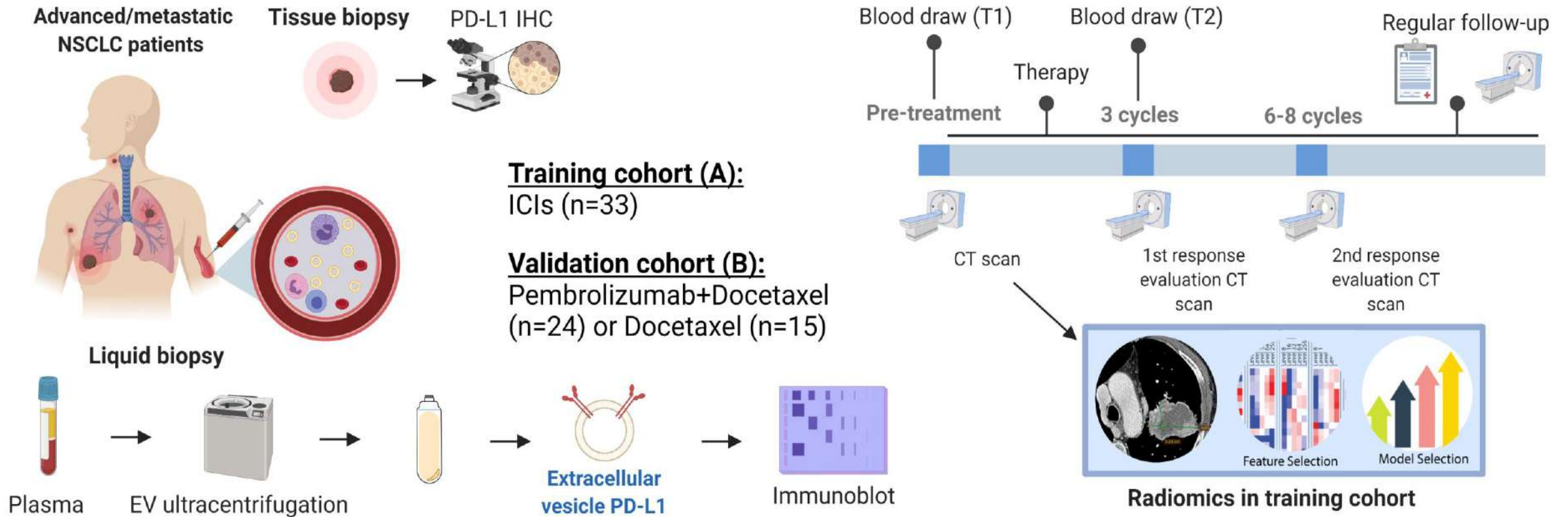
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Lack of ctDNA Clearance Is Associated with Poorer Outcomes



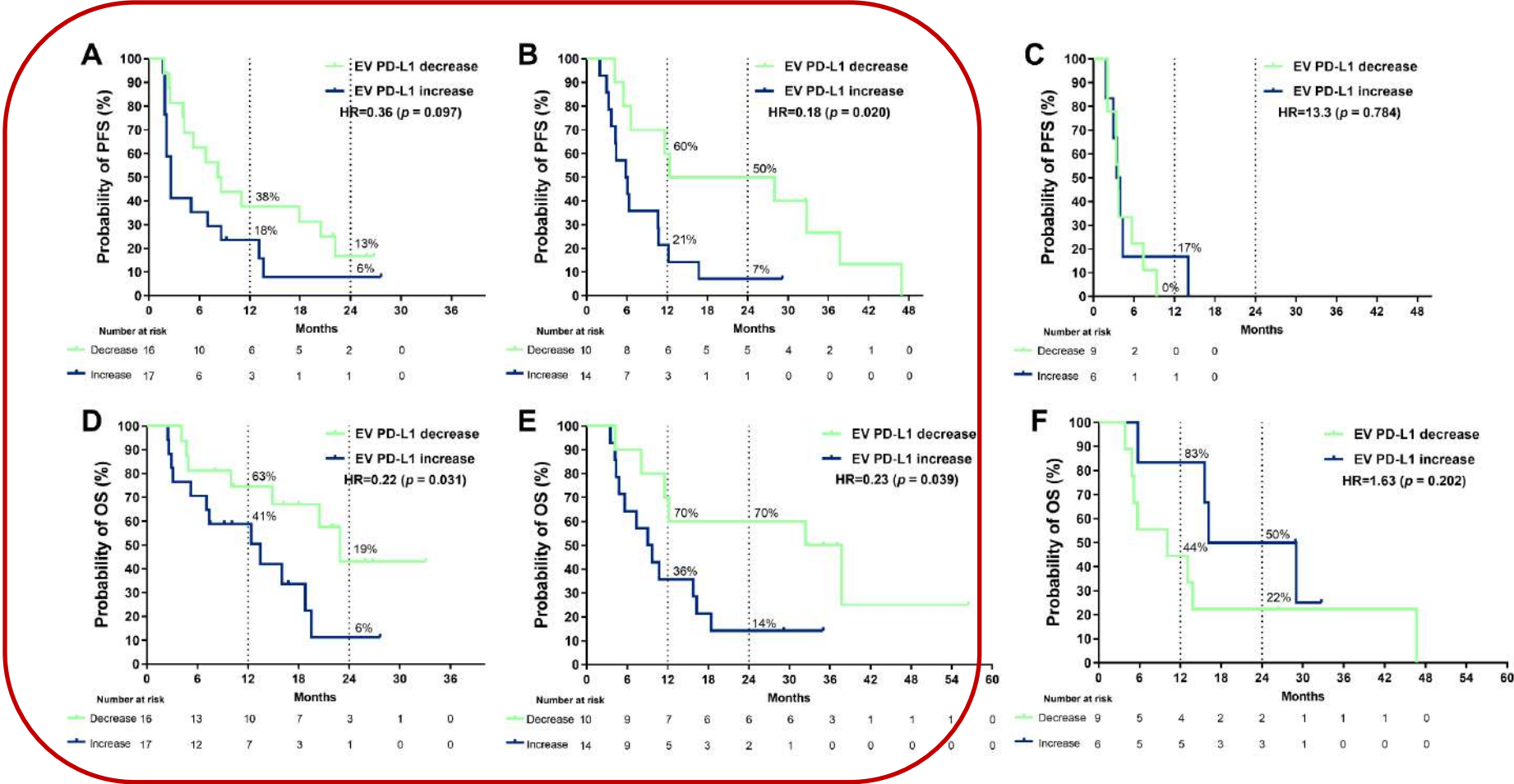
Assaf ZJF, et al. Nat Med. 2023;29(4):859-868.

EV PD-L1 as a predictor for immunotherapy response and outcome



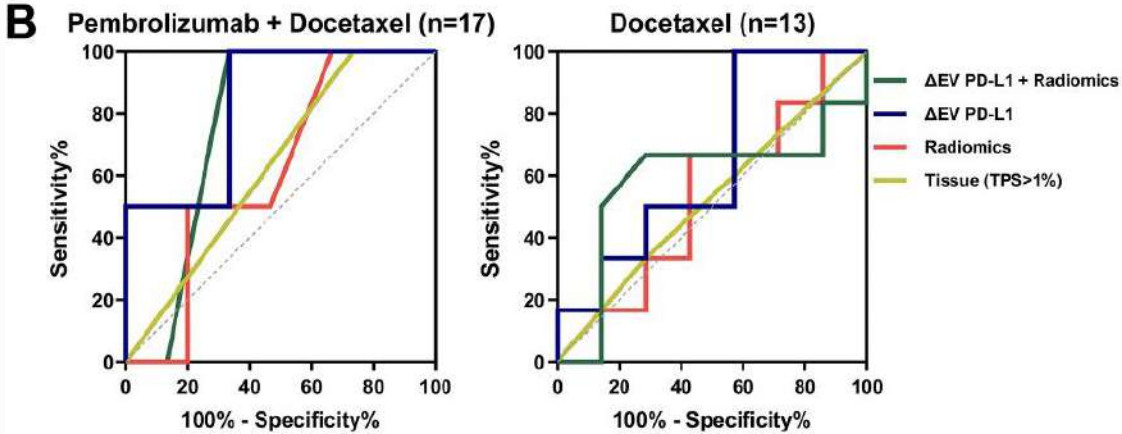
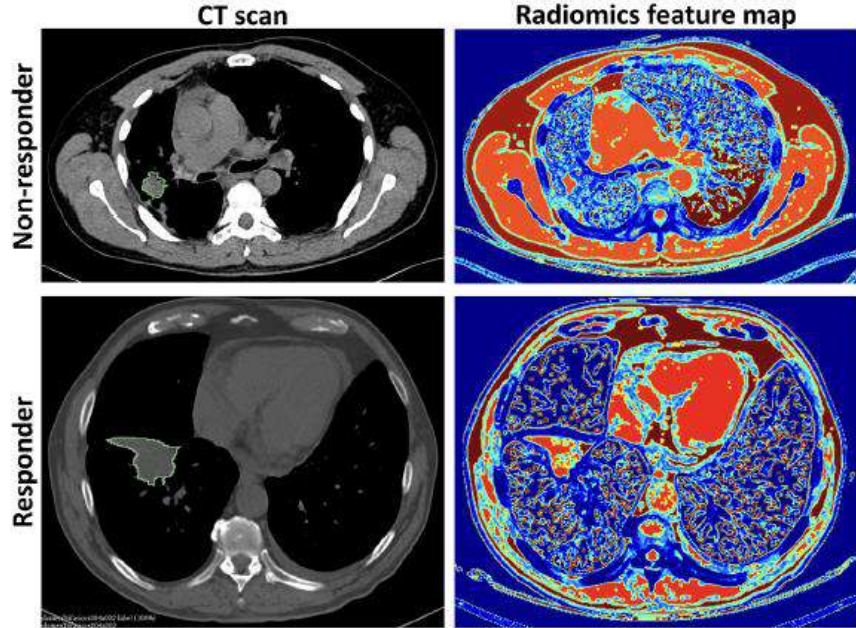
de Miguel-Perez et al. (Roflo) J Exp Clin Cancer Res (2022) 41:186

EV PD-L1 and survival outcomes



de Miguel-Perez et al. (Rolfo) J Exp Clin Cancer Res (2022) 41:186

Validation of a multiomic model of plasma extracellular vesicle PD-L1 and radiomics for prediction of response to immunotherapy in NSCLC

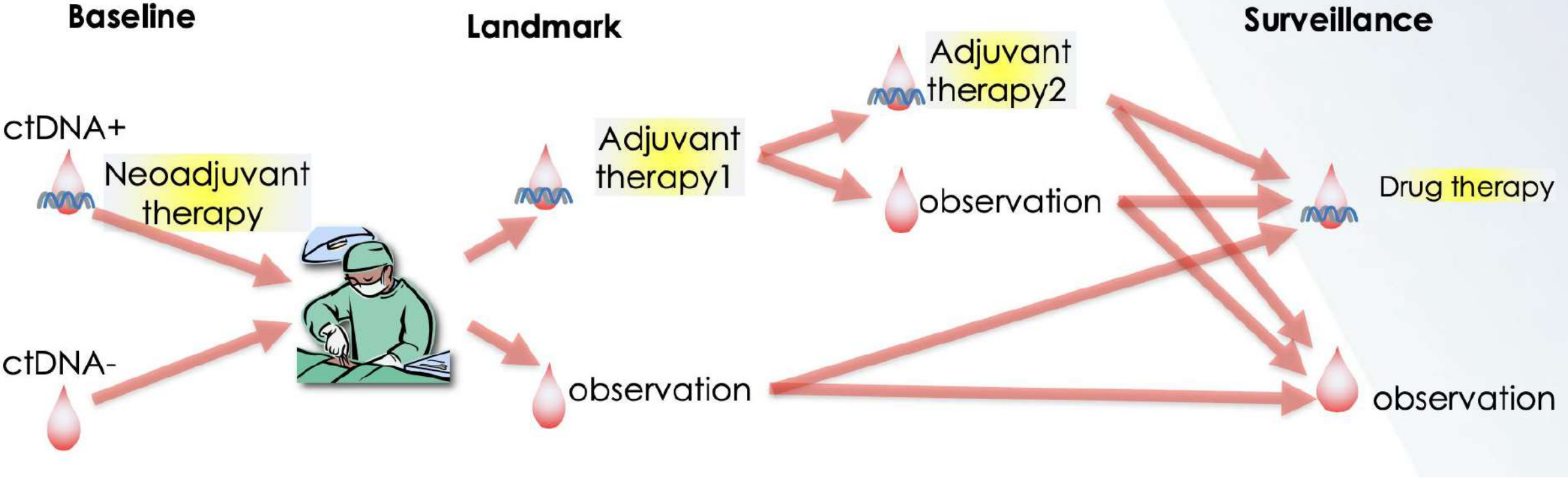


| Predictive models for Durable RECIST response (21±3 weeks) | | | | | | | | |
|--|----------------------------------|-------------|-------------|----------|------------------|-------------|-------------|----------|
| Cohort: | Pembrolizumab + Docetaxel (n=17) | | | | Docetaxel (n=13) | | | |
| Biomarker | AUC | Sensitivity | Specificity | Accuracy | AUC | Sensitivity | Specificity | Accuracy |
| ΔEV PD-L1 + Radiomics | 76.7% | 100% | 66.7% | 70.6% | 53.6% | 42.9% | 83.3% | 69.2% |
| ΔEV PD-L1 | 83.3% | 50% | 100.0% | 94.1% | 54.8% | 28.6% | 83.3% | 61.5% |
| Radiomics | 61.7% | 50% | 80% | 76.5% | 57.1% | 42.9% | 83.3% | 53.8% |
| Tissue PD-L1 ≥1% | 63.3% | 100% | 26.7% | 35.3% | 52.4% | 28.6% | 66.6% | 46.2% |

This multiparametric model showed high sensitivity and specificity at identifying non-responders to ICIs and outperformed tissue PD-L1, being directly correlated with tumor change.

de Miguel-Perez et al. (Rolfo) Journal of Experimental & Clinical Cancer Research (2024) 43:81

DNA guided perioperative management in the future



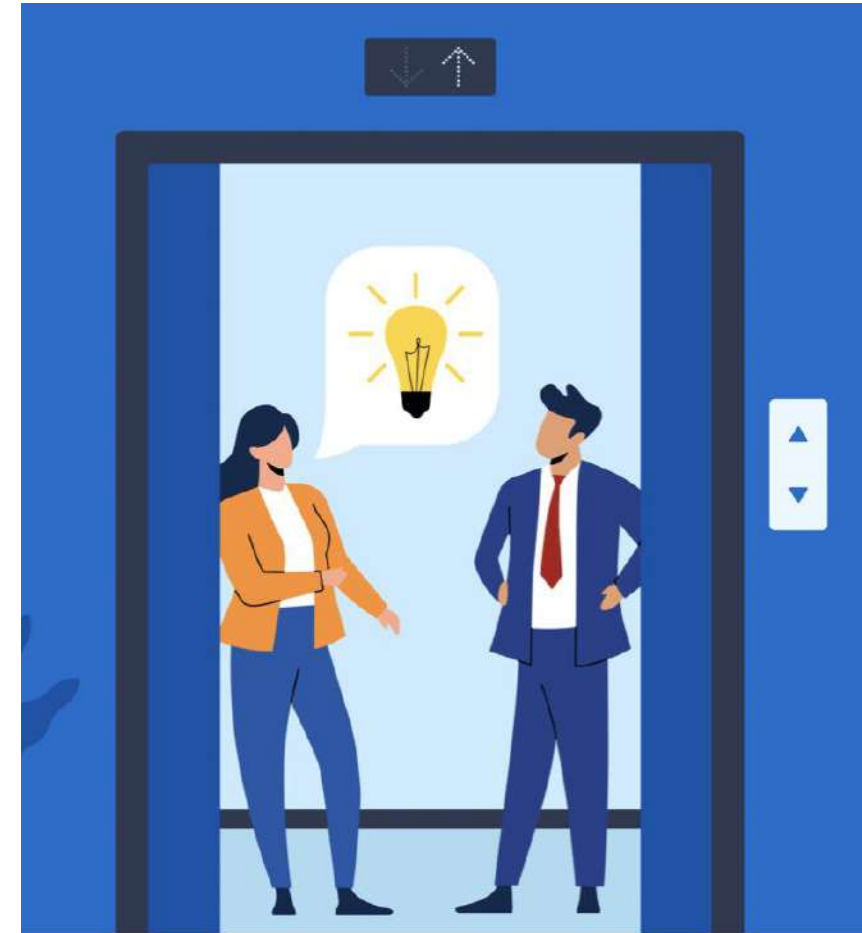
Modified from Soh, Hamada Fujino, and Mitsudomi, Cancers (Basel) . 2021

Take home message... my elevator pitch

- Liquid Biopsy is an important tool for diagnosis and monitoring.
- A good opportunity to incorporate LB in immunotherapy assesment
- Integrating liquid biopsy in clinical trials is a necessity
- Use of analytes beyond ctDNA opening the door for biology understanding.

Christian Rolfo, MD, PhD, MBA, Dr.hc.

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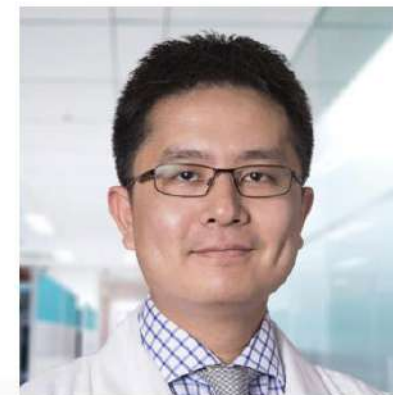
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The James



THE OHIO STATE UNIVERSITY
COMPREHENSIVE CANCER CENTER



The James



THE OHIO STATE UNIVERSITY
COMPREHENSIVE CANCER CENTER

ACKNOWLEDGEMENTS TEAM AND COLLABORATIONS

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UNIVERSITÀ
DEGLI STUDI
DI MESSINA



@ChristianRolfo
@RolfoLab

The James



ESMO DEEP DIVE: LUNG CANCER

PRECISION ONCOLOGY IN LUNG CANCER – DIAGNOSTICS

Beyond Xray vision – novel imaging technologies and radiomics

Raquel Perez-Lopez

Team Leader – Radiomics Group Vall d'Hebron Institute of Oncology (VHIO). Barcelona (Spain)

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BETTER MEDICINE
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DECLARATION OF INTERESTS

Raquel Perez-Lopez

Employment: Vall d'Hebron Institute of Oncology and Vall d'Hebron Hospital

Research Funding: CRIS Foundation, FERO Foundation, LaMarató Foundation, La Caixa Foundation, Carlos III Instituto de Investigacion, European Commission Horizon Program.

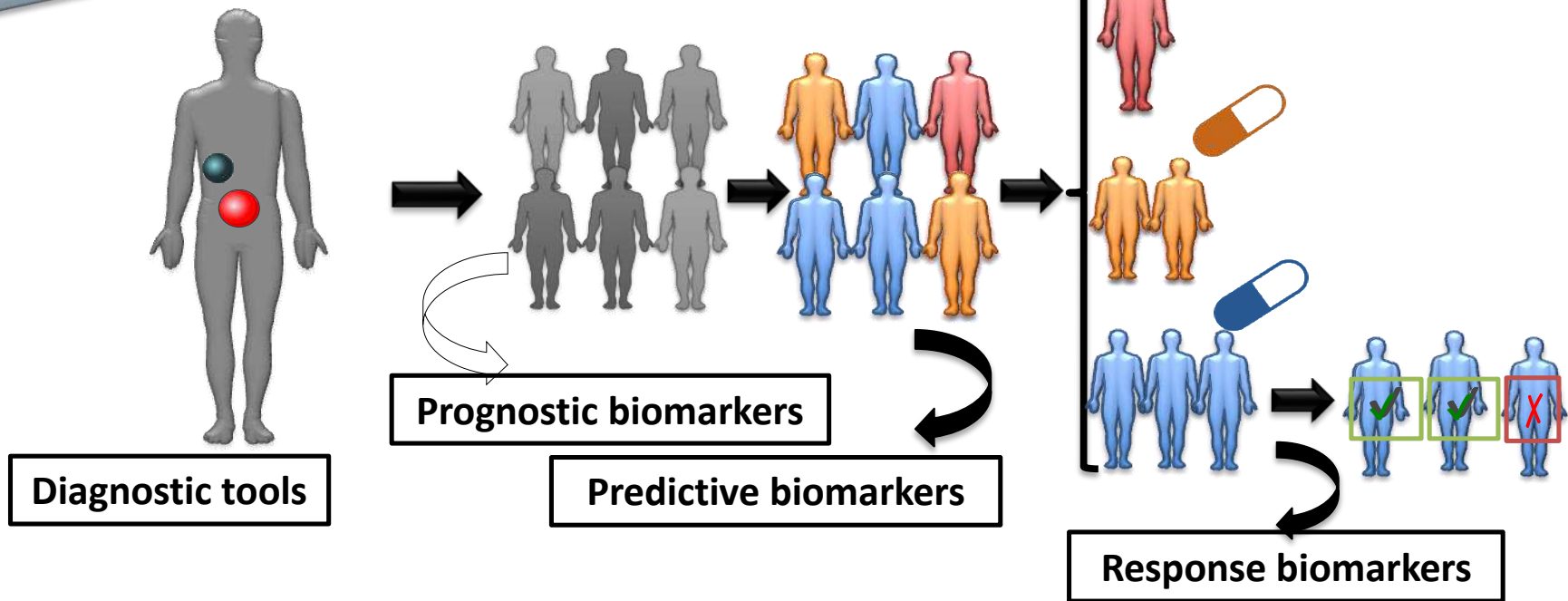
Grant support: AstraZeneca, Roche

Steering committee / advisor role: AstraZeneca, Roche

Medical imaging

Medical imaging

Medical imaging



Diagnostic tools

Prognostic biomarkers

Predictive biomarkers

Response biomarkers

Clinical Decision Support (CDS) systems

Computer-Aided Diagnosis (CAD) systems

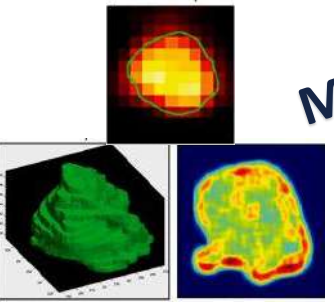
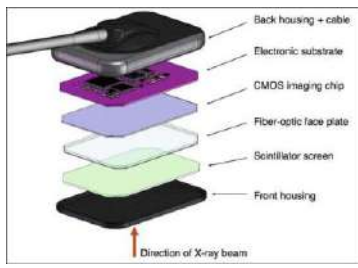
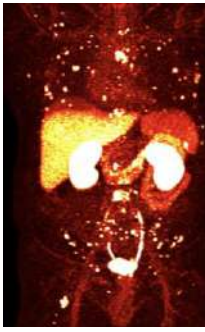
Therapeutic Decision Support (TDS) systems



X-ray era

Digital era

Radiomics era



Machine learning
Deep-learning
 CNNs
 Transformers
 Foundation models



XIX century

XX century

XXI century

1895
(X-ray)

1950
(PET)

1973
(MRI)

2000

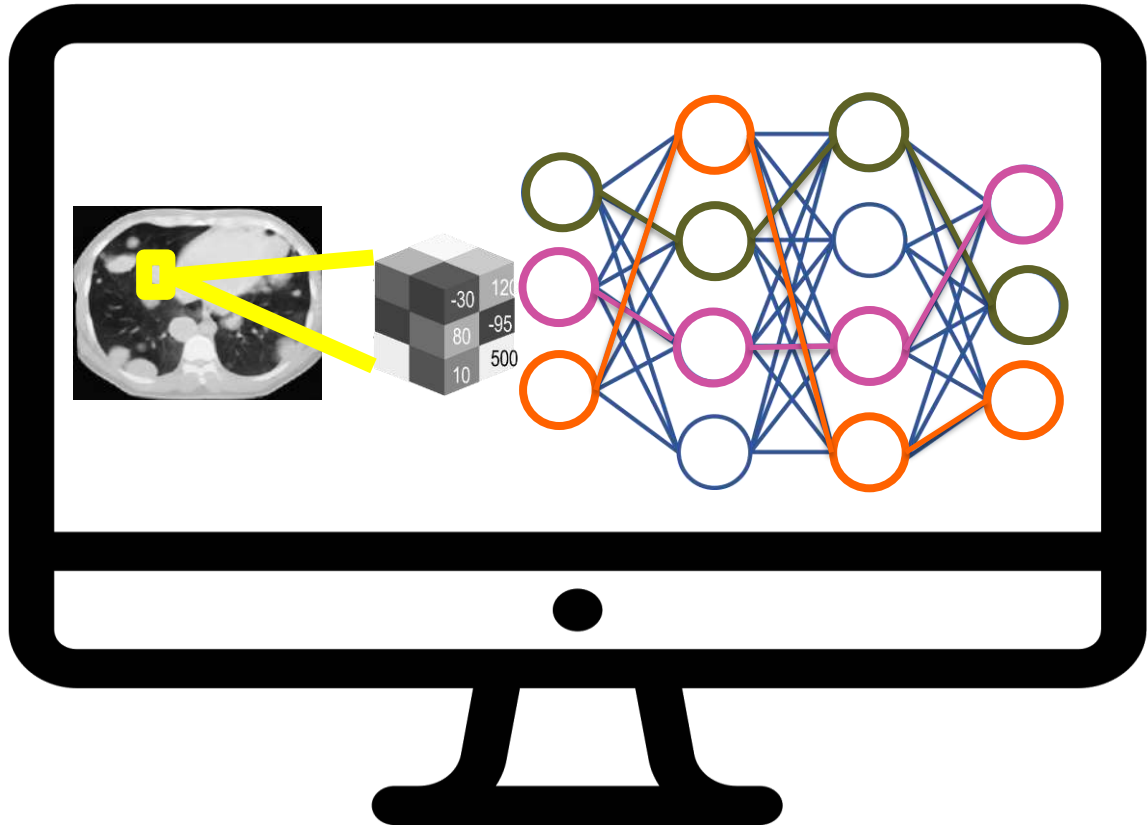
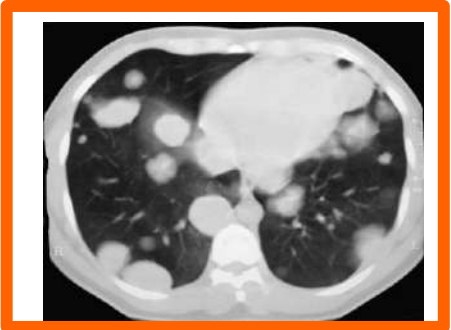
2010

2012

2025



DIAGNOSIS



Tuberculosis
COVID-19

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DETECTION & DIAGNOSIS

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LUNG CANCER SCREENING PROGRAM

Inclusion criteria



| | Recruitment period | Inclusion criteria | Primary comparison | Key outcomes |
|--------------------------|--------------------|--|---|--|
| Randomised trials | | | | |
| NLST ² | 2002-04 | Age 55-74 years; ≥30 pack-year smoking history; currently smoke or quit <15 years ago | Annual low-dose CT vs chest radiography for 3 years (n=53 454) | 20% reduction in lung cancer-related mortality with low-dose CT |
| NELSON ³ | 2003-06 | Age 50-74 years; >15 cigarettes per day for >25 years or >10 cigarettes per day for >30 years; currently smoke or quit ≤10 years ago | Low-dose CT at baseline and in year 1, year 3, and year 5-5 vs no screen (n=15 789) | 24% reduction in lung cancer-related mortality with low-dose CT in men |

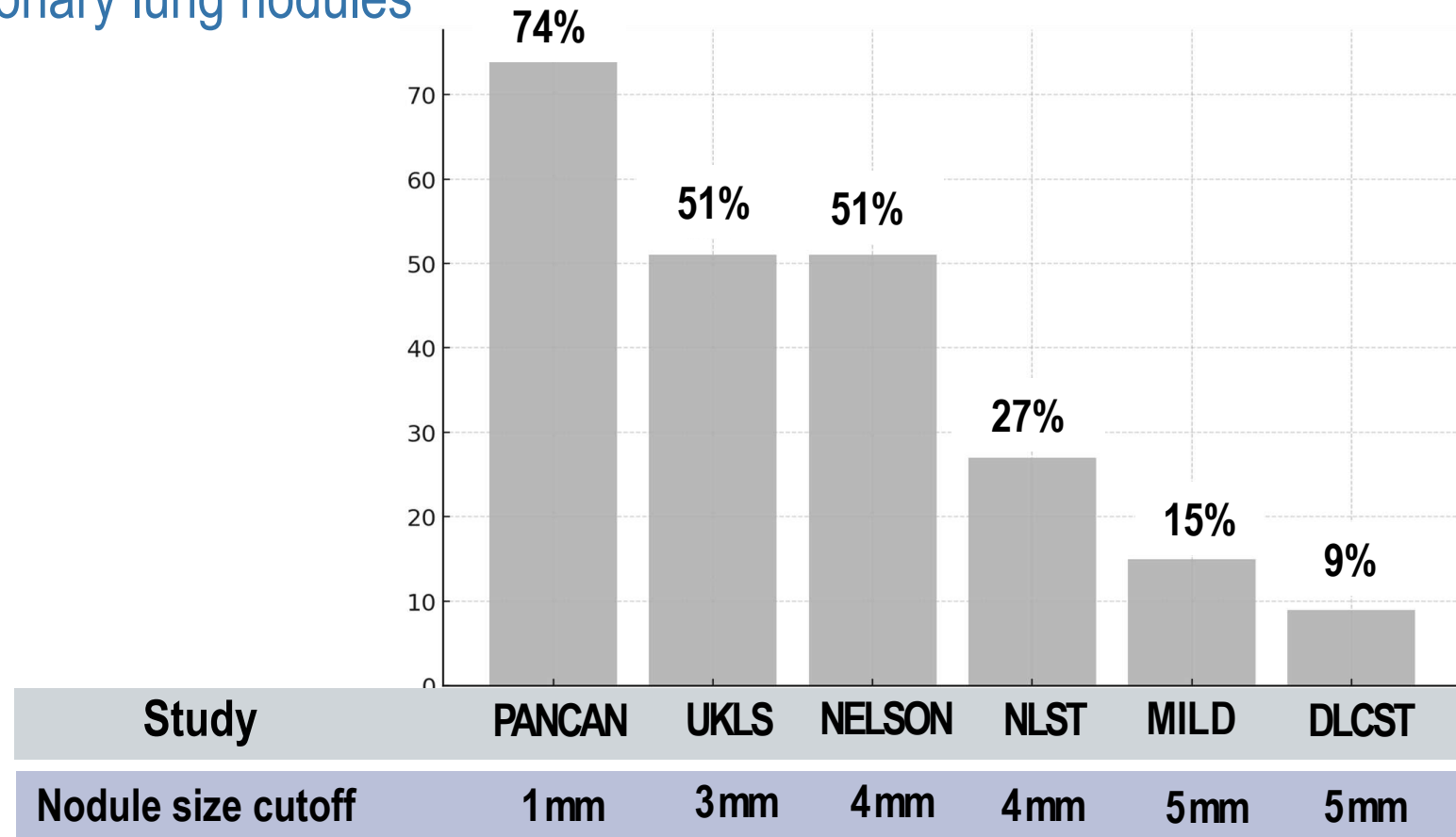
Field JK, et al. *Lancet Reg Health Eur* 2021. PMID: 34806061

de Koning HJ, et al. *N Engl J Med* 2020. PMID: 31995683

NSLT Research Team. *N Engl J Med* 2011. PMID: 21714641

LUNG CANCER SCREENING PROGRAM

Prevalence of pulmonary lung nodules



Field JK, et al. *Lancet Reg Health Eur* 2021. PMID: 34806061

de Koning HJ, et al. *N Engl J Med* 2020. PMID: 31995683

NSLT Research Team. *N Engl J Med* 2011. PMID: 21714641

LUNG CANCER SCREENING PROGRAM

Lung nodule malignancy risk

Lung-RADS® v2022



American College of Radiology™



| Lung-RADS | Category Descriptor | Findings | Management |
|-----------|--|---|--|
| 0 | Incomplete Estimated Population Prevalence: ~1% | Prior chest CT examination being located for comparison (see note 9) Part or all of lung nodules cannot be evaluated Findings suggestive of an inflammatory or infectious process (see note 10) | Comparison to prior chest CT; Additional lung cancer screening CT imaging needed; 1-3 month LDCT |
| 1 | Negative Estimated Population Prevalence: 39% | No lung nodules OR Nodule with benign features: • Complete, central, popcorn, or concentric ring calcifications OR • Fat-containing | |
| 2 | Benign - Based on imaging features or indolent behavior Estimated Population Prevalence: 45% | Juxtapleural nodule: • < 10 mm (524 mm ³) mean diameter at baseline or new AND • Solid; smooth margins; and oval, lentiform, or triangular shape Solid nodule: • < 6 mm (< 113 mm ³) at baseline OR • New < 4 mm (< 34 mm ³) Part solid nodule: • < 6 mm total mean diameter (< 113 mm ³) at baseline Non solid nodule (GGN): • < 30 mm (< 14,137 mm ³) at baseline, new, or growing OR • ≥ 30 mm (≥ 14,137 mm ³) stable or slowly growing (see note 7) Airway nodule, subsegmental - at baseline, new, or stable (see note 11) Category 3 lesion that is stable or decreased in size at 6-month follow-up CT OR Category 4B lesion proven to be benign in etiology following appropriate diagnostic workup | 12-month screening LDCT |
| 3 | Probably Benign - Based on imaging features or behavior Estimated Population Prevalence: 9% | Solid nodule: • ≥ 6 to < 8 mm (≥ 113 to < 268 mm ³) at baseline OR • New 4 mm to < 6 mm (34 to < 113 mm ³) Part solid nodule: • ≥ 6 mm total mean diameter (≥ 113 mm ³) with solid component < 6 mm (< 113 mm ³) at baseline OR • New < 6 mm total mean diameter (< 113 mm ³) Non solid nodule (GGN): • ≥ 30 mm (≥ 14,137 mm ³) at baseline or new Atypical pulmonary cyst: (see note 12) • Growing cystic component (mean diameter) of a thick-walled cyst Category 4A lesion that is stable or decreased in size at 3-month follow-up CT (excluding airway nodules) | 6-month LDCT |

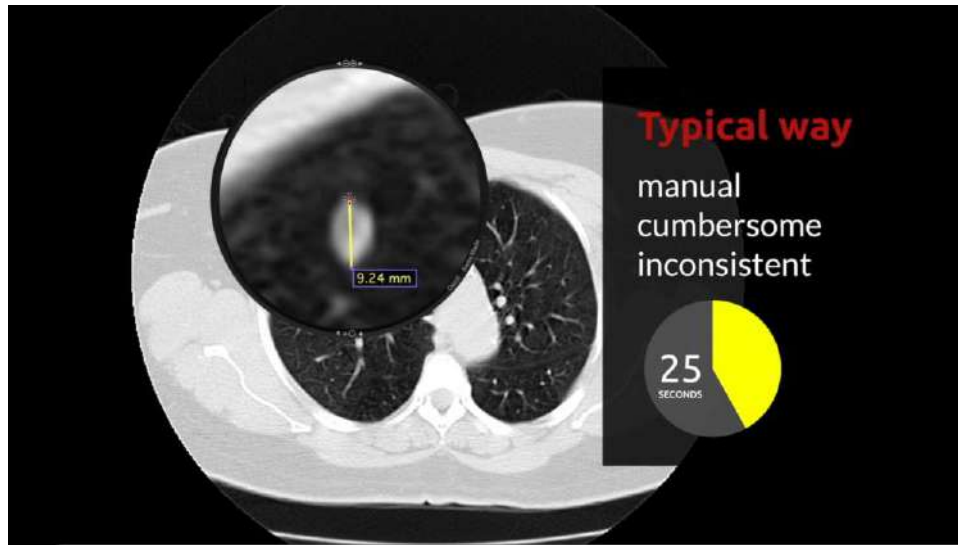
| | | | |
|----|---|---|---|
| 4A | Suspicious Estimated Population Prevalence: 4% | Solid nodule: • ≥ 8 to < 15 mm (≥ 268 to < 1,767 mm ³) at baseline OR • Growing < 8 mm (< 268 mm ³) OR • New 6 to < 8 mm (113 to < 268 mm ³) Part solid nodule: • ≥ 6 mm total mean diameter (≥ 113 mm ³) with solid component ≥ 6 mm to < 8 mm (≥ 113 to < 268 mm ³) at baseline OR • New or growing < 4 mm (< 34 mm ³) solid component Airway nodule, segmental or more proximal - at baseline (see note 11) Atypical pulmonary cyst: (see note 12) • Thick-walled cyst OR • Multilocular cyst at baseline OR • Thin- or thick-walled cyst that becomes multilocular | 3-month LDCT; PET/CT may be considered if there is a ≥ 8 mm (≥ 268 mm ³) solid nodule or solid component |
| 4B | Very Suspicious Estimated Population Prevalence: 2% | Airway nodule, segmental or more proximal - stable or growing (see note 11) Solid nodule: • ≥ 15 mm (≥ 1,767 mm ³) at baseline OR • New or growing ≥ 8 mm (≥ 268 mm ³) Part solid nodule: • Solid component ≥ 8 mm (≥ 268 mm ³) at baseline OR • New or growing ≥ 4 mm (≥ 34 mm ³) solid component Atypical pulmonary cyst: (see note 12) • Thick-walled cyst with growing wall thickness/nodularity OR • Growing multilocular cyst (mean diameter) OR • Multilocular cyst with increased loculation or new/increased opacity (nodular, ground glass, or consolidation) Slow growing solid or part solid nodule that demonstrates growth over multiple screening exams (see note 8) | Referral for further clinical evaluation Diagnostic chest CT with or without contrast; PET/CT may be considered if there is a ≥ 8 mm (≥ 268 mm ³) solid nodule or solid component; tissue sampling; and/or referral for further clinical evaluation Management depends on clinical evaluation, patient preference, and the probability of malignancy (see note 13) |
| 4X | Estimated Population Prevalence: < 1% | Category 3 or 4 nodules with additional features or imaging findings that increase suspicion for lung cancer (see note 14) | |
| S | Significant or Potentially Significant Estimated Population Prevalence: 10% | Modifier: May add to category 0-4 for clinically significant or potentially clinically significant findings unrelated to lung cancer (see note 15) | As appropriate to the specific finding |

American College of Radiology Committee on Lung-RADS®. Lung-RADS Assessment Categories 2022.

Available at <https://www.acr.org/-/media/ACR/Files/RADS/Lung-RADS/Lung-RADS-2022.pdf>

LUNG CANCER SCREENING PROGRAM

AI-aided lung nodule malignancy risk assessment



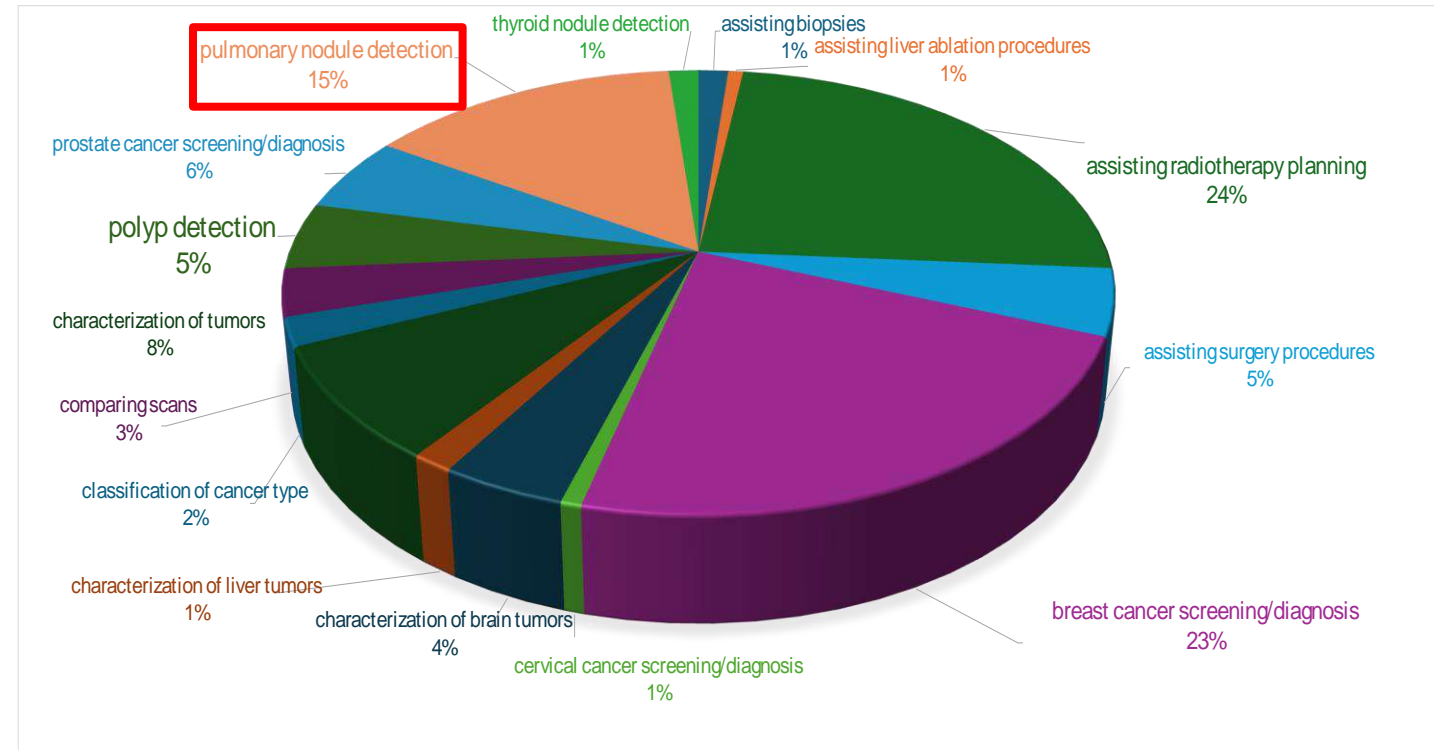
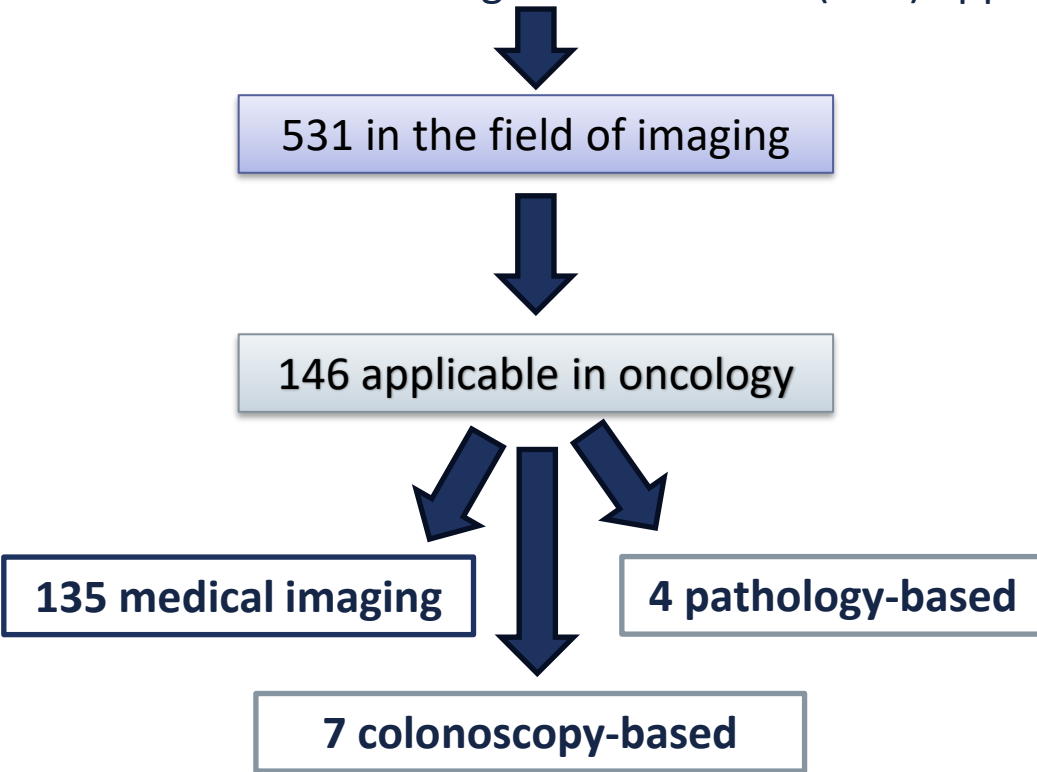
Quanyang W, et al. *Cancer Med.* 2024. PMID: 38581113

TABLE 4 The latest artificial intelligence-based methods for classifying benign and malignant nodules.

| Year | Authors | Data source | Method | Quality index | Quality index value |
|------|--------------------------------------|-----------------|---|-------------------------|---------------------|
| 2016 | Petousis et al. ¹¹² | NLST dataset | DBNs Including three expert-driven DBNs and two DBNs derived from structure learning methods | AUC | >0.75 |
| 2018 | Filho et al. ⁷⁷ | LIDC-IDRI | Topology-based phylogenetic diversity indices are proposed for features engineering and selection. Feature data are fed to 2D CNNs | Accuracy AUC | 0.9263 0.934 |
| 2018 | Causey et al. ⁹⁰ | LIDC-IDRI | Training 3D CNN models and collecting output features. A 3D CNN is then used for malignancy classification based on quantitative image features | AUC | 0.99 |
| 2018 | Dey et al. ⁹¹ | LIDC-IDRI | Performance comparison between 3D DCNN and 3D DenseNet variants | Accuracy AUC | 0.899 0.9459 |
| 2019 | Balagurunathan et al. ¹¹³ | NLST dataset | Optimal linear classifiers | AUC | 0.85 |
| 2019 | Al-Shabi et al. ¹¹⁴ | LIDC-IDRI | Deep Local-Global networks containing residual blocks and non-local blocks | AUC | 0.9562 |
| 2019 | Chen et al. ¹¹⁵ | LIDC-IDRI | Using Med3D models pre-trained on ResNets, initialize classification networks using Med3D models | Accuracy | 0.9192 |
| 2020 | Harsono et al. ⁸¹ | LIDC-IDRI | Integrated modified pre-trained inflated 3D ConvNets with FPN | AUC | 0.8184 |
| 2020 | Yang et al. ¹¹⁶ | LIDC-IDRI | Self-attention transformer based on 3D DenseNets and MIL algorithms | AUC | 0.932 |
| 2021 | Yu et al. ¹⁰³ | LIDC-IDRI | Res-trans networks | Accuracy AUC | 0.9292 0.9628 |
| 2021 | Halder et al. ¹¹⁷ | LIDC-IDRI | Two-path morphological 2D CNN | Accuracy AUC | 0.9610 0.9936 |
| 2019 | Xie et al. ⁷⁸ | LIDC-IDRI | MV-KBC model can learn 3-D lung nodule characteristics by decomposing a 3D nodule into nine fixed views | Accuracy AUC | 0.916 0.957 |
| 2018 | Zhu et al. ⁵⁰ | LIDC-IDRI | R-CNN-GBM | Accuracy | 0.9274 |
| 2019 | Nasrullah et al. ⁶¹ | LIDC-IDRI | CMixNet-GBM | Sensitivity | 0.94 |
| 2023 | Mikhael et al. ¹¹⁸ | NLST | 3D Resnet | AUC | 0.92 |
| 2023 | Bushara et al. ¹¹⁹ | LIDC | LCD-CapsNet | Accuracy AUC | 0.94 0.989 |
| 2023 | Irshad et al. ¹²⁰ | Exasens dataset | An IGWO-based DCNN model | Accuracy Sensitivity | 98.27% 97.67% |

FDA APPROVED AI-ENABLED MEDICAL DEVICES

692 U.S. Food and Drug Administration (FDA) approved AI-enabled medical devices



<https://radiology.healthregister.com/>

Joshi G et al, Electronics 2024. doi: 10.3390/electronics13030498

* Updated 19 October, 2023

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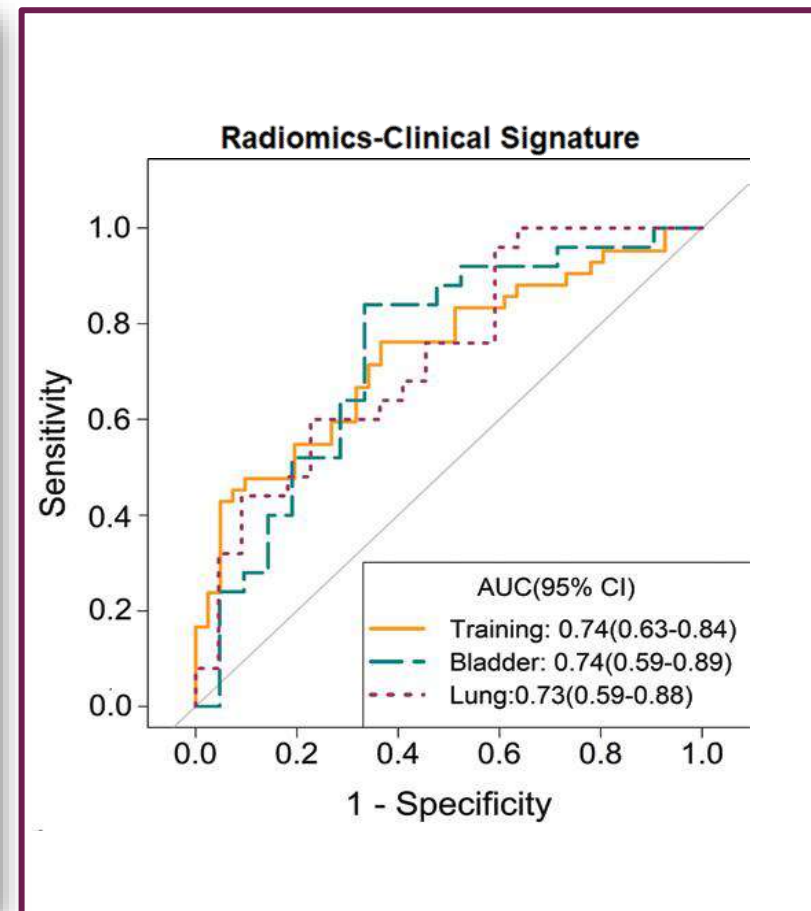
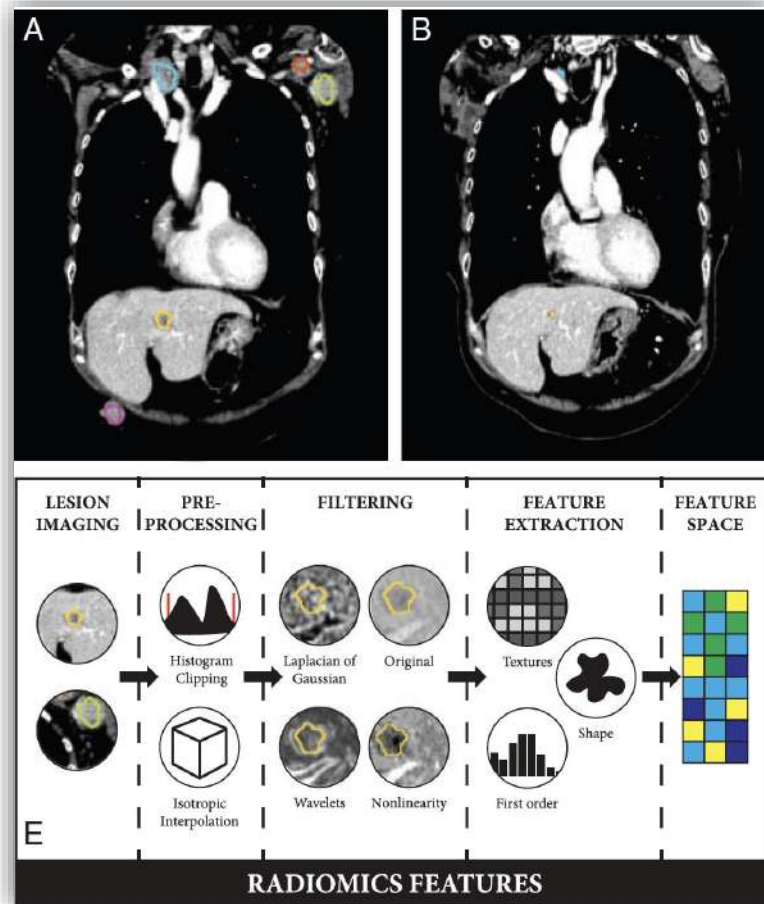
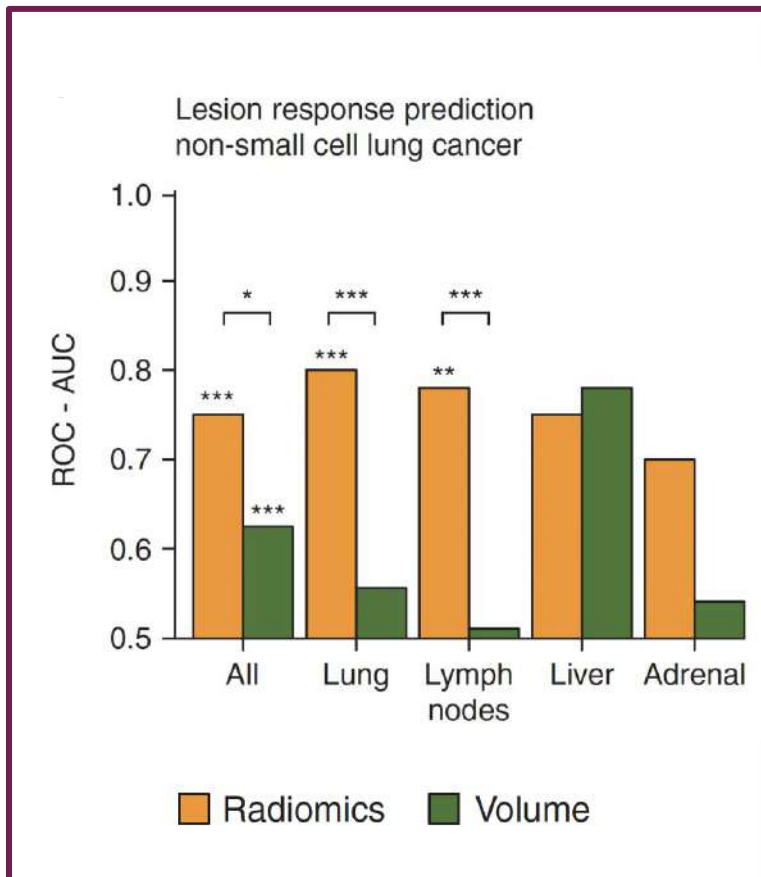
ENHANCED RESPONSE PREDICTIONS AND ON-TREATMENT EVALUATION

Computational analysis of standard imaging



IMMUNE-RADIOMICS SIGNATURES

End-to-end signature for response prediction

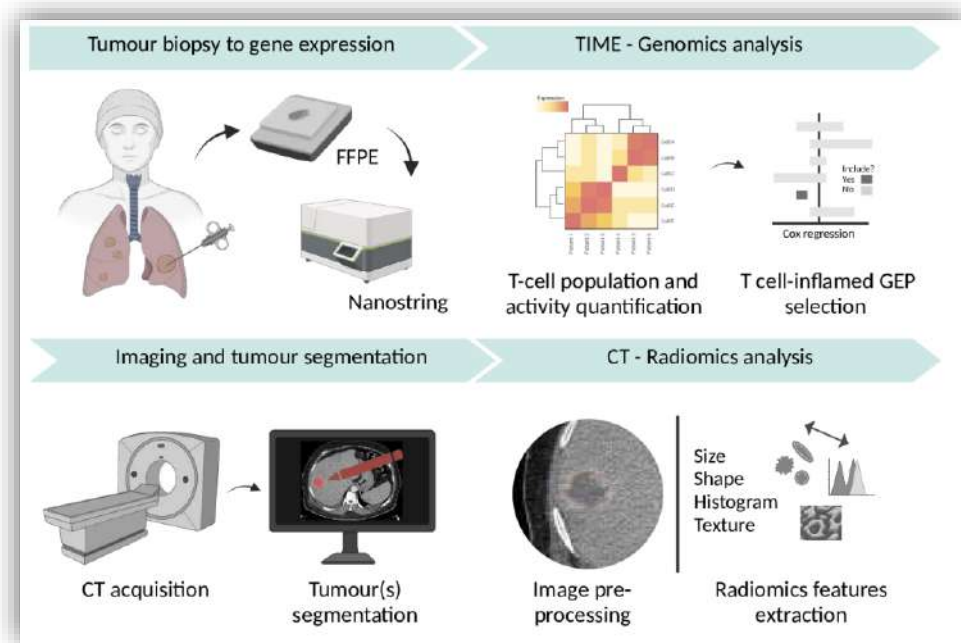


Trebeschi S, et al. Ann Oncol. 2019. PMID: 30895304

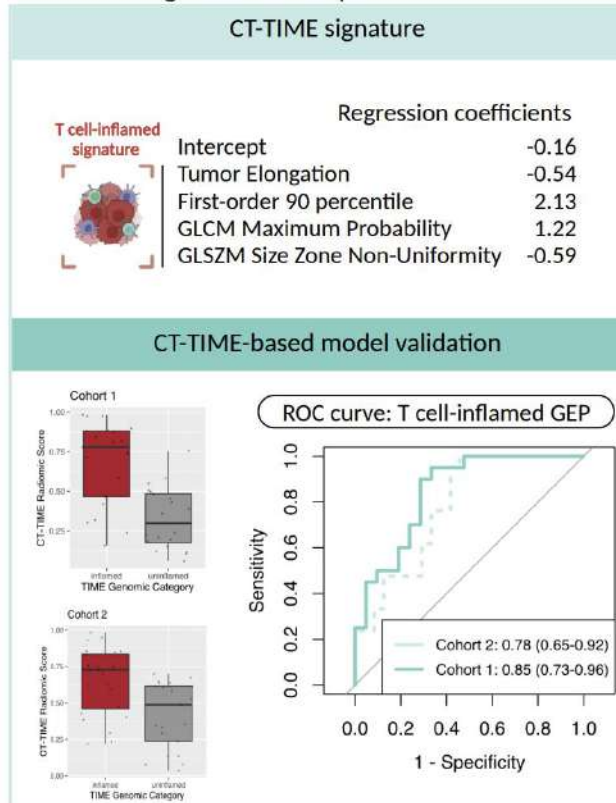
Ligero M, et al. Radiology. 2021. PMID: 33497314

IMMUNE-RADIOMICS SIGNATURES

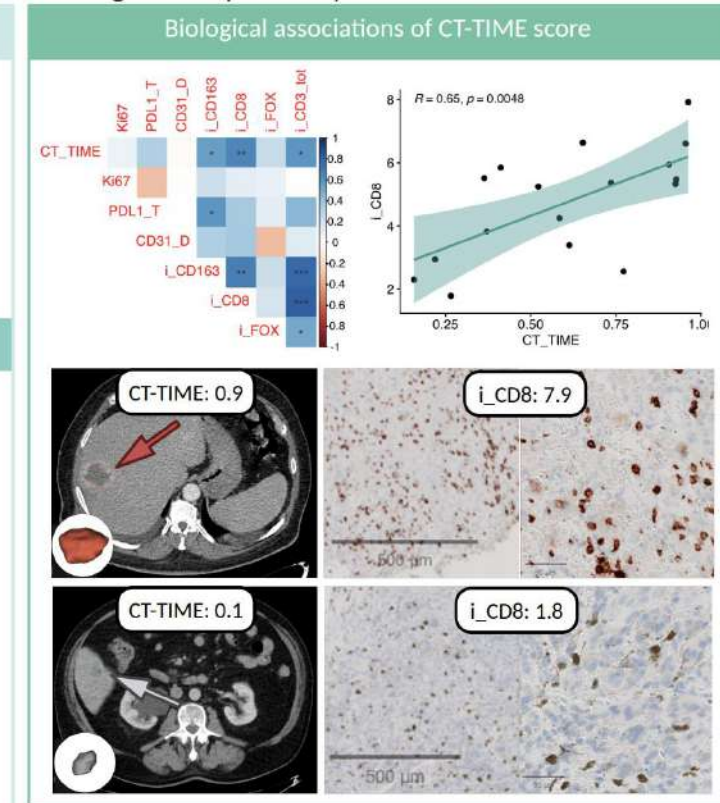
Biomarker prediction



A CT-TIME signature development and validation



B Biological interpretability of CT-TIME

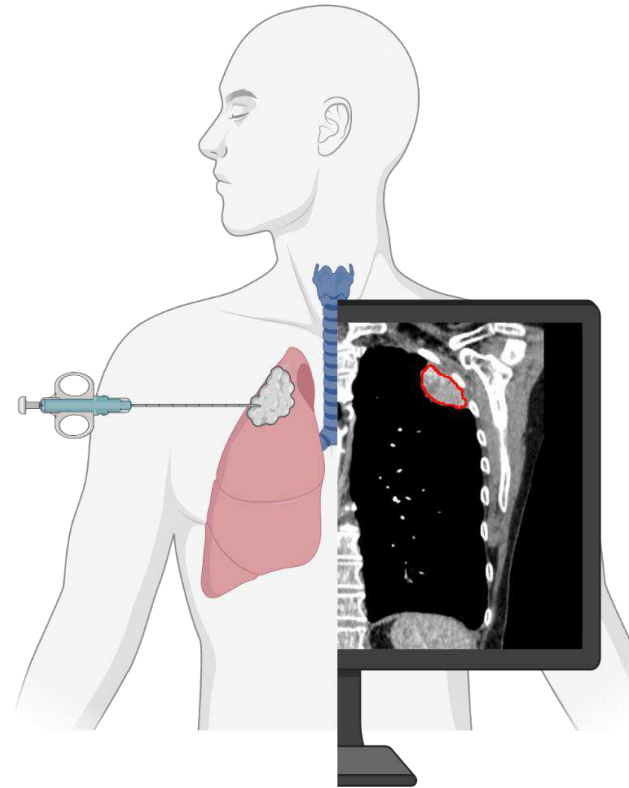
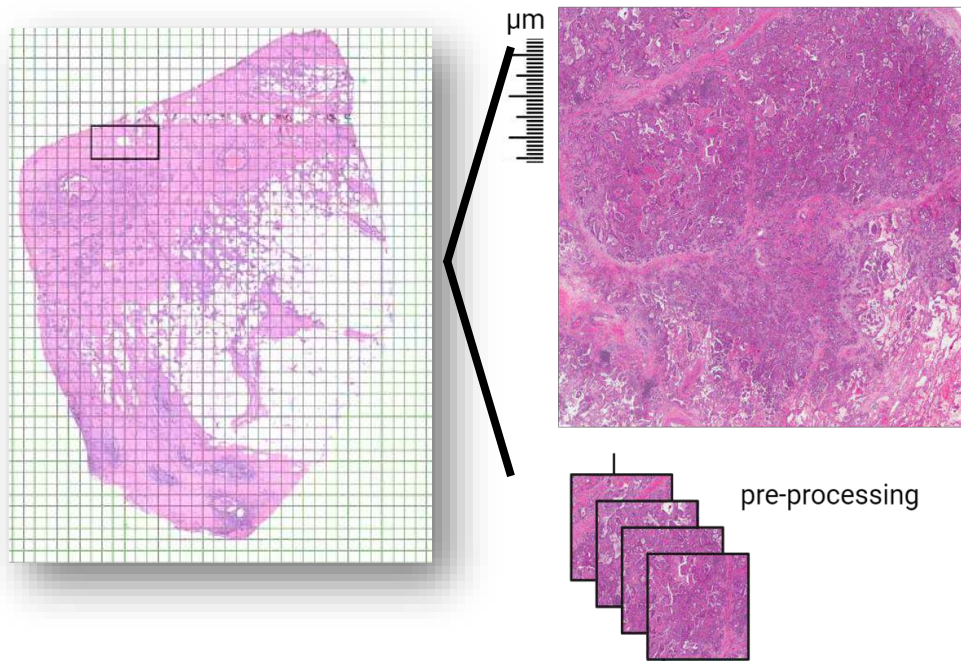


Bernatowicz K, et al. *J Immunother Cancer*. 2025. PMID: 39800381

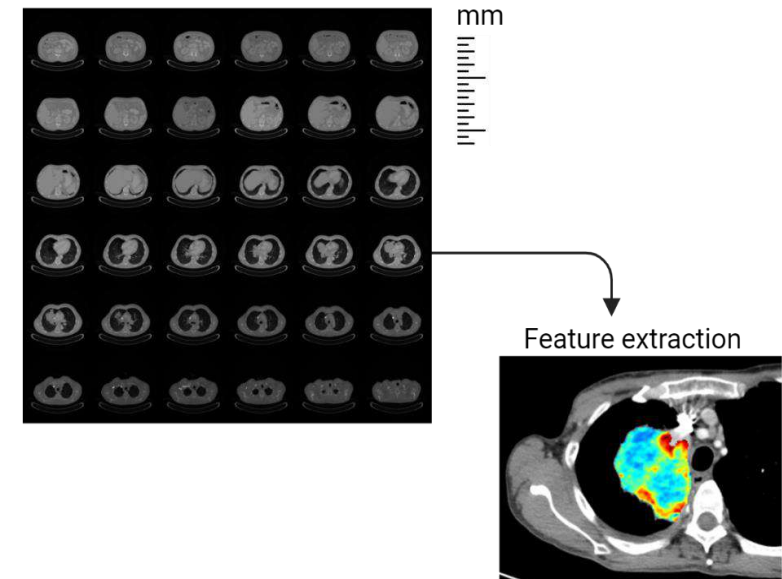
Pathology and radiology provide valuable information at different scales



Histopathology images



Radiology images

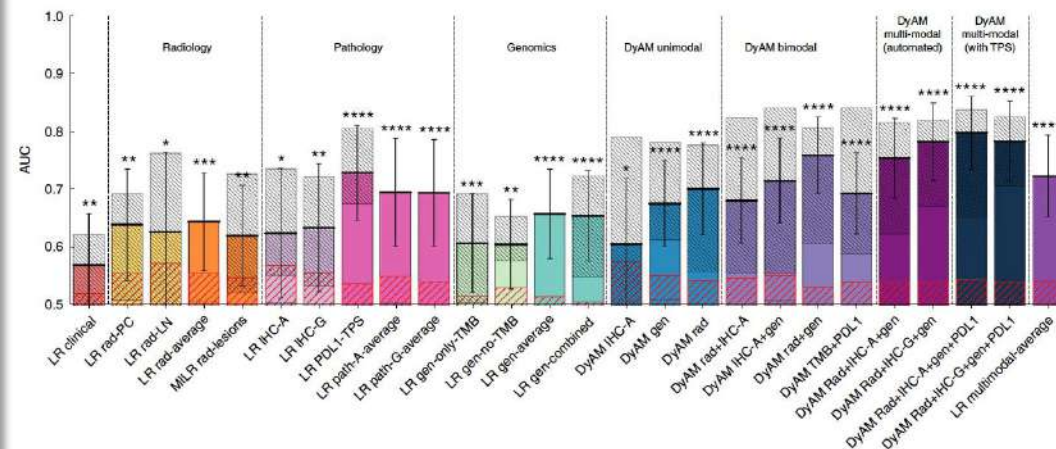
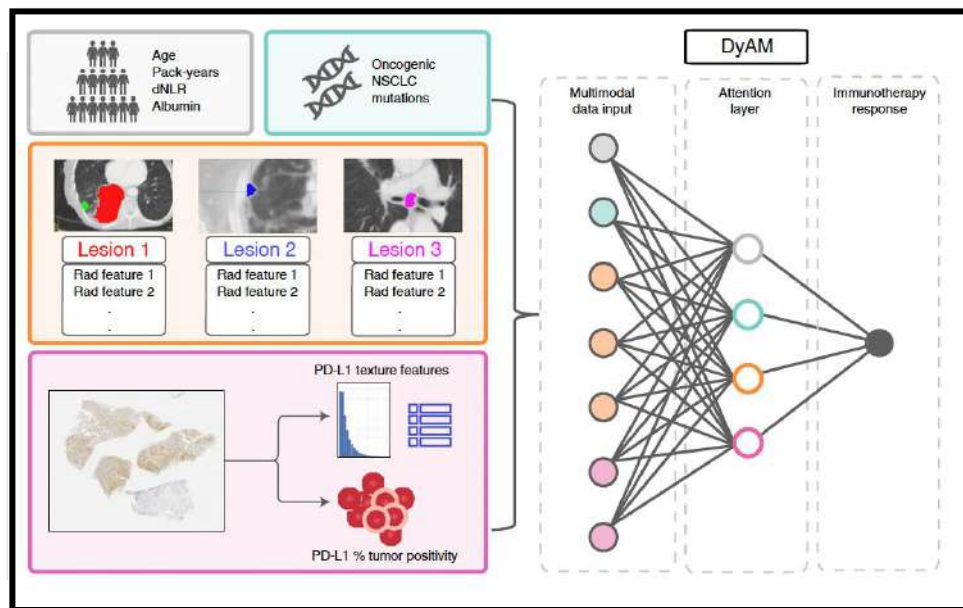
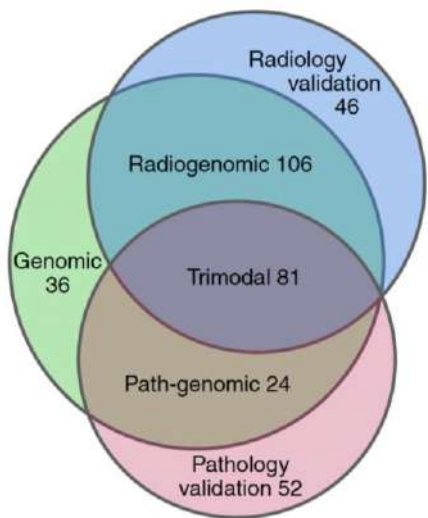


Perez-Lopez R, et al .Nat Rev Cancer. 2024. PMID: 38755439
Ghaffari Laleh N, et al. Clin Cancer Res. 2023. PMID: 36083132



Multimodal data integration improves AI-model performance

Response prediction to PD-L1 inhibitors in NSCLC



Vanguri RS, et al. Nat Cancer. 2022.PMID: 36038778

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ENHANCED RESPONSE PREDICTIONS AND ON-TREATMENT EVALUATION

Advanced imaging techniques

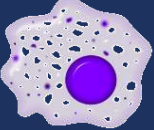


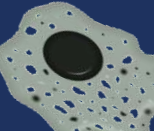

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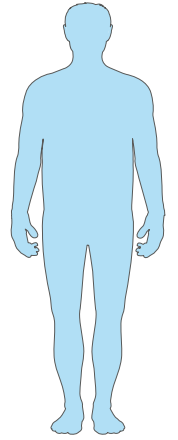
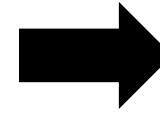
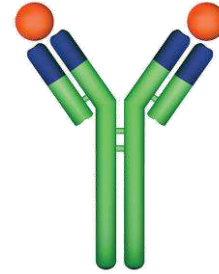
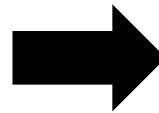
ESMO GOOD SCIENCE
BETTER MEDICINE
BEST PRACTICE



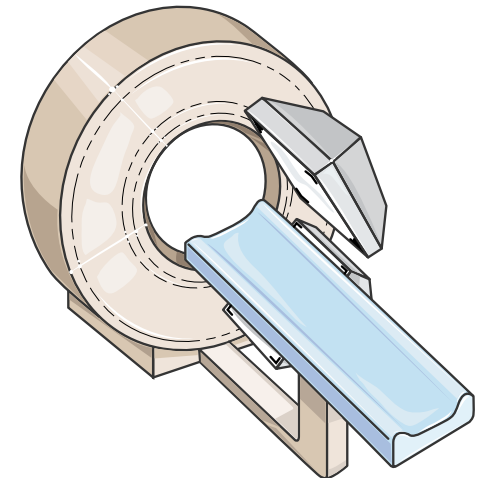
Imaging of immune cell dynamics with immunotracers



-  Macrophages: CD206, PD-L1, S100A9
-  T-lymphocytes: CD4, CD8, CD69, PD-1
-  B-lymphocytes: CD19, CD20
-  Phagocytes: CD163
-  Tumor cells: PD-L1

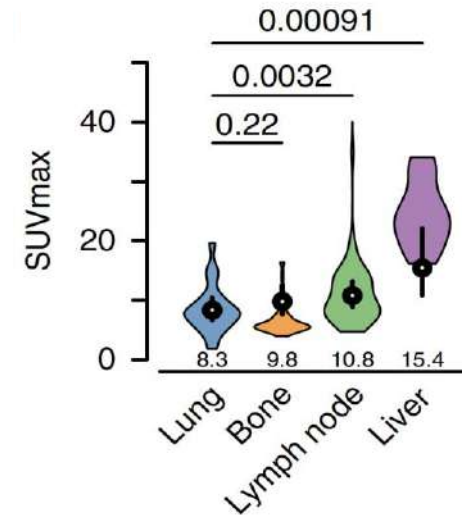
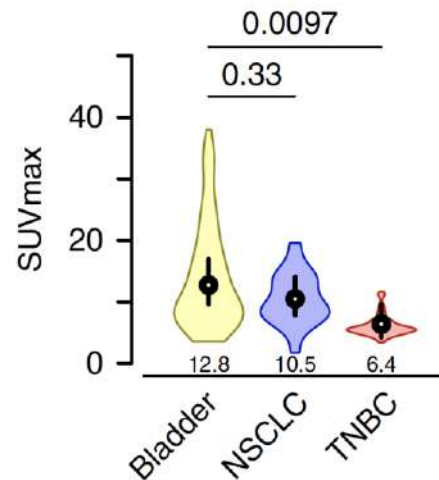
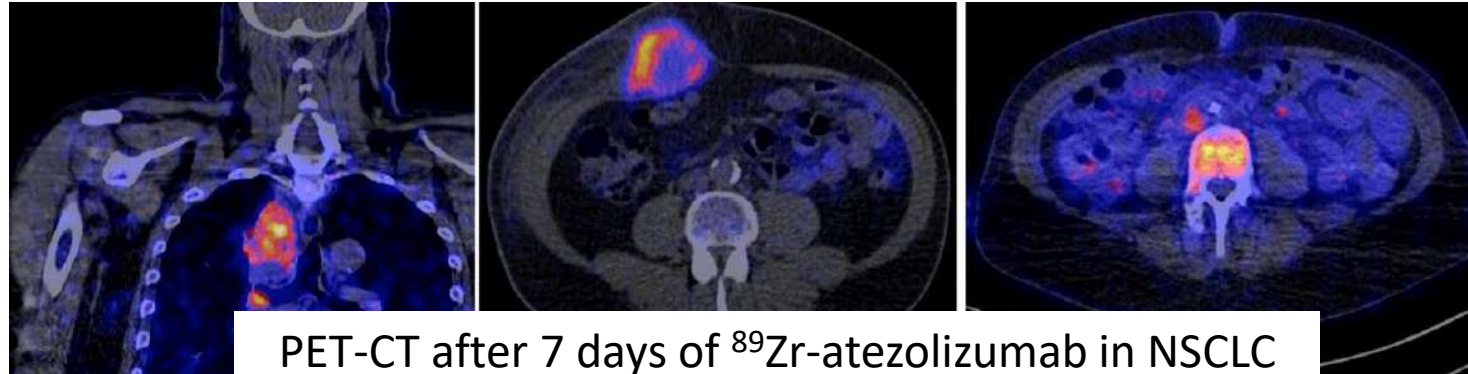


^{11}C , ^{18}F , ^{68}Ga , ^{89}Zr



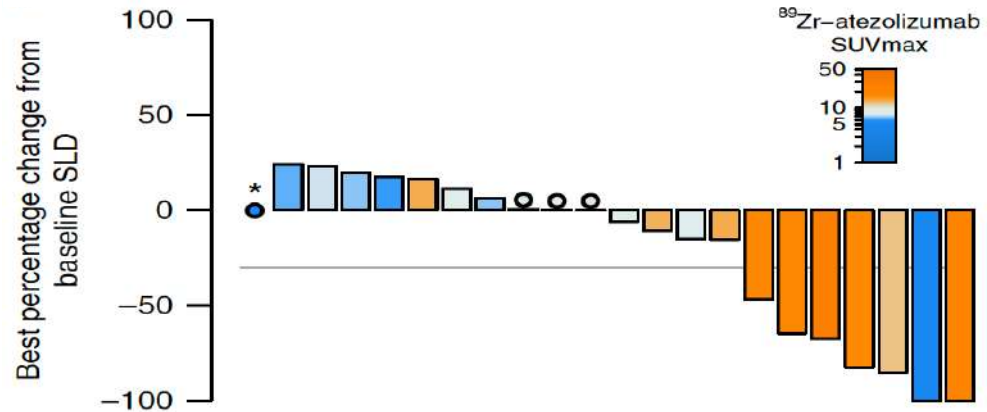
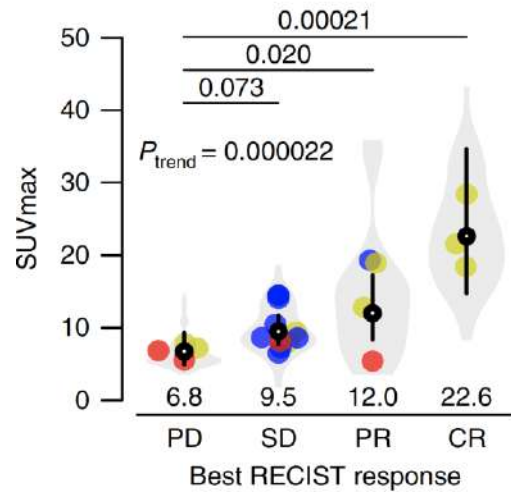
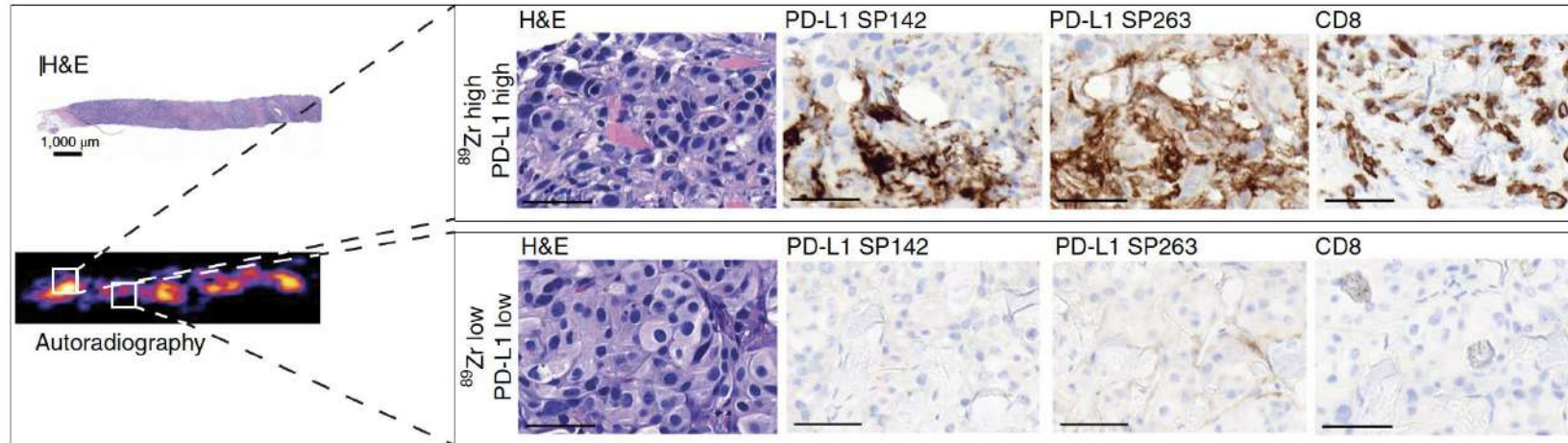
⁸⁹Zr-atezolizumab PET-CT for assessing response to PD-L1 inhibition

22 patients with
non-small cell lung cancer (NSCLC), bladder cancer or triple-negative breast cancer (TNBC)



Bensch F, et al. Nat Med. 2018. PMID: 30478423

⁸⁹Zr-atezolizumab PET-CT for assessing response to PD-L1 inhibition

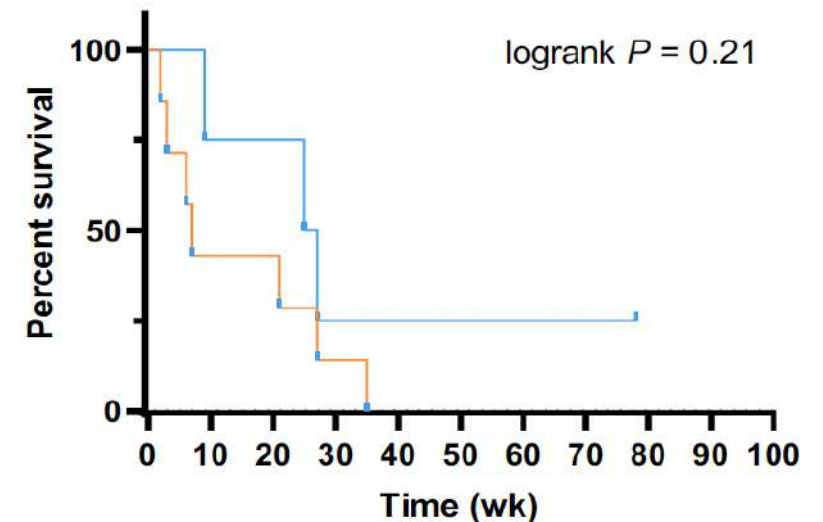
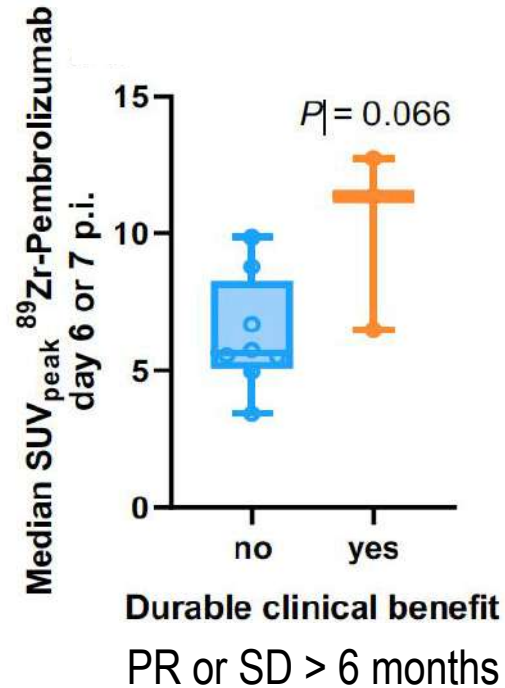
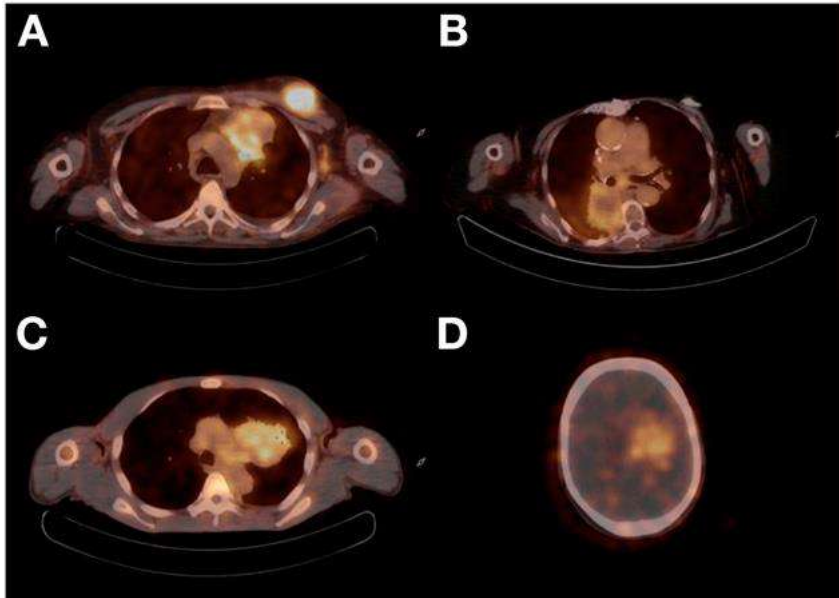


Bensch F, et al. Nat Med. 2018. PMID: 30478423

⁸⁹Zr-pembrolizumab PET-CT for assessing response to PD-1 inhibition

12 NSCLC patients eligible for pembrolizumab monotherapy as first- or later-line therapy

SUV_{peak} on day 6-7, when tumour uptake was the highest and blood-pool activity was the lowest (best tumour-to-background ratio)



Niemeijer AN, et al. *J Nucl Med.* 2022. PMID: 34272316

Take home messages

- AI-driven tools **enhance lung nodule detection** and size measurements.
- AI models are advancing the ability to **distinguish between benign and malignant nodules**. These improvements are expected to further refine risk assessment in the near future, reducing unnecessary follow-ups and invasive biopsies.
- AI and advanced imaging techniques may help immunotherapy selection by providing non-invasive, real-time insights into the tumour microenvironment.



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<https://vhio.net/pf/radiomics-group/>

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